

Journal of Integrated OMICS

a methodological journal

Editors-in-Chief

Carlos Lodeiro-Espiño

Florentino Fdez-Riverola

Jens Coorssen

Jose-Luís Capelo-Martínez

JIOMICS

Journal of Integrated OMICS

Focus and Scope

Journal of Integrated OMICS, JIOMICS, provides a forum for the publication of original research papers, preliminary communications, technical notes and critical reviews in all branches of pure and applied "-omics", such as genomics, proteomics, lipidomics, metabolomics or metallomics. The manuscripts must address methodological development. Contributions are evaluated based on established guidelines, including the fundamental nature of the study, scientific novelty, and substantial improvement or advantage over existing technology or method. Original research papers on fundamental studies, and novel sensor and instrumentation development, are especially encouraged. It is expected that improvements will also be demonstrated within the context of (or with regard to) a specific biological question; ability to promote the analysis of molecular mechanisms is of particular interest. Novel or improved applications in areas such as clinical, medicinal and biological chemistry, environmental analysis, pharmacology and materials science and engineering are welcome.

Editors-in-Chief

Carlos Lodeiro-Espiño, University NOVA of Lisbon, Portugal

Florentino Fdez-Riverola, University of Vigo, Spain

Jens R. Coorssen, University of Western Sydney, NSW, Australia

Jose-Luís Capelo-Martínez, University NOVA of Lisbon, Portugal

Regional editors

ASIA

Gary Xiao

Director of Functional Genomics and Proteomics Laboratories at
Osteoporosis Research Center, Creighton University Omaha, Nebraska, USA

Yogeshwer Shukla

Proteomics laboratory at Indian Institute of Toxicology Research (Council of
Scientific and Industrial Research), Lucknow, India

AUSTRALIA AND NEW ZEALAND

Jens R. Coorssen

University of Western Sydney, NSW, Australia

Europe

Gilberto Igrejas

University of Trás-os-Montes and Alto Douro, Life Sciences and
Environmental School, Institute for Biotechnology and Bioengineering, Centre
of Genetics and Biotechnology
Department of Genetics and Biotechnology, 5001-801 Vila Real, Portugal

Martin von Bergen

UFZ, Helmholtz-Centre for Environmental Research, Department of
Proteomics, Permoserstr. 15, 04318 Leipzig, Germany

Jan Ottervald

Research and Development | Innovative Medicines Neuroscience, CNSP iMed
Science Södertälje, AstraZeneca, Sweden

North America

Randen Patterson

Center for Computational Proteomics, The Pennsylvania State University, USA

School of Medicine, The University of North Carolina at Chapel Hill, USA

Oscar Alzate

Associate Professor of Cell and Developmental Biology, Adjunct Associate Professor in Neurology, Director: Systems Proteomics Center

Yue Ge

US Environmental Protection Agency, Research Triangle Park, USA

South America**Eduardo Alves de Almeida**

Depto. de Química e Ciências Ambientais, IBILCE - UNESP, Brazil

Carlos H. I. Ramos

Chemistry Institute – UNICAMP, Brazil

Marco Aurélio Zezzi Arruda

University of Campinas - Unicamp

Associated editors**AFRICA****Saffaj Taouqif**

Centre Universitaire Régional d'Interface, Université Sidi Mohamed Ben Abdallah, route d'Imouzzar-Fès, Morocco

ASIA**Abdul Jaleel A**

Rajiv Gandhi Centre for Biotechnology, Thycad PO, Trivandrum, Kerala, India

Ali A. Ensafi

Isfahan University of Technology, Iran

Allison Stelling

Dresden, Germany

Amita Pal

Division of Plant Biology, Bose Institute, Kolkata, India

Ashish Gupta

Centre of Biomedical Magnetic Resonance, SGPGIMS Campus, Lucknow, India

Canhua Huang

The State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, PR China

Chaminda Jayampath Seneviratne

Oral Biosciences, Faculty of Dentistry, University of Hong Kong, Hong Kong

Cheolju Lee

Korea Institute of Science and Technology, Seoul, Korea

Chi Chiu Wang

Department of Obstetrics & Gynaecology, Chinese University of Hong Kong, Hong Kong

Chii-Shiarnng Chen

National Museum of Marine Biology and Aquarium, Checheng, Pingtung, Taiwan

Ching-Yu Lin

Institute of Environmental Health, College of Public Health, National Taiwan University, Taipei, Taiwan

Chantragan Srisomsap

Chulabhorn Research Institute, Bangkok, Thailand

Chen Han-Min

Department of Life Science, Catholic Fu-Jen University, Taipei, Taiwan

David Yew

Chinese University of Hong Kong, Shatin, N.T., Hong Kong

Debmalya Barh

Institute of Integrative Omics and Applied Biotechnology (IIOAB), India

Dwaipayan Bharadwaj

Genomics & Molecular Medicine Unit, Institute of Genomics & Integrative Biology (CSIR), Mall Road, Delhi, India

Eiji Kinoshita

Department of Functional Molecular Science, Graduate School of Biomedical Sciences, Hiroshima University, Japan

Eun Joo Song

Molecular Recognition Research Center, Korea Institute of Science & Technology, Seoul, Korea

Fan Chen

Institute of Genetics and Developmental Biology, Chinese Academy of Sciences (CAS), China

Feng Ge

Institute of Hydrobiology, Chinese Academy of Sciences, China

Ganesh Chandra Sahoo

BioMedical Informatics Center of Rajendra Memorial Research Institute of Medical Science (RMRIMS), Patna, India

Guangchuang Yu

Institute of Life & Health Engineering, Jinan University, Guangzhou, China

Gufeng Wang

Department of Chemistry, North Carolina State University, Raleigh, USA

Hai-Lei Zheng

School of Life Sciences, Xiamen University, China

Hee-bal Kim

Department of Food and Animal Biotechnology of the Seoul National University, Korea

Hsin-Yi Wu

Institute of Chemistry, Academia Sinica, Taiwan

Hitoshi Iwahashi

Health Research Institute, National Institute of Advanced Industrial Science and Technology (AIST), Japan

Hong-Lin Chan

National Tsing-Hua University, Taiwan

Hongying Zhong

College of Chemistry, Central China Normal University, Wuhan, P. R. China

Huan-Tsung Chang

Department of Chemistry, National Taiwan University, Taipei, Taiwan

HuaXu

Research Resources Center, University of Illinois, Chicago

Hui-Fen Wu

Department of Chemistry, National Sun Yat – Sen University, 70, Lien-Hai Road, 80424, Kaohsiung, Taiwan

Hye-Sook Kim

Faculty of Pharmaceutical Sciences, Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Okayama University, Japan

Hyun Joo An

ChungNam National University, Daejeon, Korea (South)

Ibrokhim Abdurakhmonov

Institute of Genetics and Plant experimental Biology Academy of Sciences of Uzbekistan, Uzbekistan

Isam Khalaila

Biotechnology Engineering Department, Ben-Gurion University, Israel

Jagannadham Medicharla

Senior Principal Scientist, CSIR-Centre for Cellular and Molecular Biology, Hyderabad, India

Jianghao Sun

Food Composition and Method Development Lab, U.S. Dept. of Agriculture, Agricultural Research Services, Beltsville, USA

Jong Won Yun

Dept. of Biotechnology, Kyungsan, Kyungbuk 712-714, Republic of Korea

Juan Emilio Palomares-Rius

Forestry and Forest Products Research Institute, Tsukuba, Japan

Jung Min Kim

Liver and Immunology Research Center, Daejeon Oriental Hospital of Daejeon University, Republic of Korea

Kazuaki Kakehi

School of Pharmacy, Kinki University, Kowakae 3-4-1, Higashi-Osaka, 577-8502, Japan

Kazuki Sasaki

Department of Molecular Pharmacology, National Cerebral and Cardiovascular Center, Japan

Ke Lan

West China School of Pharmacy, Sichuan University, Chengdu, China

Kelvin Leung

Department of Chemistry, Hong Kong Baptist University, Hong Kong

Kobra Pourabdollah

Razi Chemistry Research Center (RCRC), Shahreza Branch, Islamic Azad University, Shahreza, Iran

Kohji Nagano

Chugai Pharmaceutical Co. Ltd., Japan

Koji Ueda

Laboratory for Biomarker Development, Center for Genomic Medicine, RIKEN, Tokyo, Japan

Krishnakumar Menon

Amrita Center for Nanosciences and Molecular Medicine, Amrita Institute of Medical Sciences, Kochi, Kerala, India

Lakshman Samaranayake

Dean, And Chair of Oral Microbiology, University of Hong Kong, Hong Kong

Lal Rai

Molecular Biology Section, Centre of Advanced Study in Botany, Banaras Hindu University, Varanasi-221005, India

Lei Zhou

Singapore Eye Research Institute, Singapore

Li Jianke

Institute of Apicultural Research, Chinese Academy of Agricultural Science, Beijing, China, HKSAR, PR China

Ling Zheng

College of Life Sciences, Wuhan University, China

Luk John Moonching

National University of Singapore, Singapore

Mahdi Ghasemi-Varnamkhasti

Department of Agricultural Machinery Engineering, Faculty of Agriculture, Shahrekord University, Shahrekord, Iran

Manjunatha Kini

Department of Biological Sciences, National University of Singapore, Singapore

Masahiro Sugimoto

Graduate School of Medicine and Faculty of Medicine, Kyoto University Medical Innovation Center, Japan

Masaya Miyazaki

National Institute of Advanced Industrial Science and Technology, 807-1 Shuku, Tosu, Saga 841-0052, Japan

Ming-Fa Hsieh

Department of Biomedical Engineering, Chung Yuan Christian University, Taiwan

Mingfeng Yang

Key Laboratory of Urban Agriculture of Ministry of Agriculture P. R. China Beijing University of Agriculture, China

Mo Yang

Interdisciplinary Division of Biomedical Engineering, the Hong Kong Polytechnic University, Hong Kong, China

Mohammed Rahman

Center of Excellence for Advanced Materials Research (CEAMR), King Abdulaziz University, Jeddah, Saudi Arabia

Moganty Rajeswari

Department of Biochemistry, All India Institute of Medical Sciences, Ansari Nagar, New Delhi, India

Nam Hoon Cho

Dept. of Pathology, Yonsei University College of Medicine, Korea

Ningwei Zhao

Life Science & Clinical Medicine Dept. ; Shimadzu (China) Co., Ltd

Pei-Yuan Qian

Division of Life Science, Hong Kong University of Science and Technology, China

Peng Zhou

Center of Bioinformatics (COBI), Key Laboratory for NeuroInformation of Ministry of Education (KLNME), University of Electronic Science and Technology of China (UESTC)

Poh-Kuan CHONG (Shirly)

National University of Singapore, Singapore

Qian Shi

Institutes of Biomedical Sciences, Fudan University, Shanghai, China

Qionglin Liang

Tsinghua University, Beijing, China

Rakesh Mishra

Centre for Cellular and Molecular Biology, Hyderabad, India

Roger Beuerman

Singapore Eye Research Institute, Singapore

Sameh Magdeldin Mohamed

Niigata prefecture, Nishi-ku, Terao, Niigata, Japan

Sanjay Gupta

Advanced Centre for Treatment, Research and Education in Cancer (ACTREC), Tata Memorial Centre, Kharghar, Navi Mumbai, India

Sanjeeva Srivastava

Indian Institute of Technology (IIT) Bombay, India

Seiichi Uno

Education and Research Center for Marine Resources and Environment, Faculty of Fisheries, Kagoshima University, Japan

Sen-Lin Tang

Biodiversity Research Center, Academia Sinica, Taipei, Taiwan

Setsuko Komatsu

National Institute of Crop Science, Japan

Shaojun Dai

Alkali Soil Natural Environmental Science Center, Key Laboratory of Saline-alkali Vegetation Ecology Restoration in Oil Field, Ministry of Education, Northeast Forestry University, P.R. China

Shipin Tian

Institute of Botany, Chinese Academy of Sciences, China

Songping Liang

Hunan Normal University, Changsha City, China

Steven Shaw

Department of Obstetrics and Gynecology, Chang Gung Memorial Hospital, Linkou, Taiwan

Suresh Kumar

Department of Applied Chemistry, S. V. National Institute of Technology, Gujarat, India

Tadashi Kondo

National Cancer Center Research Institute, Japan

Taesung Park

National Research Laboratory of Bioinformatics and Biostatistics at the Department of Statistics Seoul National University, Korea

Toshihide Nishimura

Department of Surgery I, Tokyo Medical University, Tokyo, Japan

Vishvanath Tiwari

Department of Biochemistry, Central University of Rajasthan, India

Wei Wang

School of Medical Sciences, Edith Cowan University, Perth, Australia

Weichuan Yu

Department of Electronic and Computer Engineering and Division of Biomedical Engineering, The Hong Kong University of Science and Technology, Clear Water Bay, Kowloon, Hong Kong, China

Wei-dong Zhang

Lab of Natural Products, School of Pharmacy, Second Military Medical University, Shanghai, China

Wenxiong Lin

School of Life Sciences, Fujian Agriculture and Forestry University, China

William Chen Wei Ning

School of Chemical and Biomolecular Engineering Nanyang Technological University, Singapore

Xiao LiWang

Division of Cardiovascular Diseases, Mayo Clinic, Rochester, MN

Xiao Zhiqiang

Key Laboratory of Cancer Proteomics of Chinese Ministry of Health, Xiangya Hospital, Central South University, 87 Xiangya Road, Changsha, Hunan 410008, P.R. China

Xiaoping Wang

Key Laboratory of Molecular Biology & Pathology, State Bureau of Chinese

Medicine, China

Xuanxian Peng

School of Life Sciences, Sun Yat-sen University, Guangzhou, China

Yang Liu

Department of Chemistry, Tsinghua University, Beijing, China

YasminAhmad

Peptide and Proteomics Division Defence Institute of Physiological and Allied Research (DIPAS), DRDO, Ministry of Defence, Timarpur, Delhi-54, India

Yin Li

Institute of Microbiology, Chinese Academy of Sciences, Beijing, China

Yong Song Gho

Department of Life Science, POSTECH, Pohang, Korea

Yoon-E Choi

Chonbuk National University, Iksan-si, South Korea

Yoon-Pin Lim

Department of Biochemistry, National University of Singapore, Singapore

Young-Gyu Ko

College of Life Sciences and Biotechnology, Korea University, Korea

Young-Suk Kim

Department of Food Science and Engineering, College of Engineering, Ewha Womans University, Seoul, Korea

Youngsoo Kim

Department of Biomedical Sciences, Seoul National University College of Medicine, Seoul, Republic of Korea

Youxiong Que

National Research & Development Center for Sugarcane, China Agriculture Research System(CARS), Fujian Agriculture & Forestry University, Republic of China

Yu-Chang Tyan

Department of Medical Imaging and Radiological Sciences, Kaohsiung Medical University, Kaohsiung, Taiwan

Yu Wang

Department of Pharmacology and Pharmacy, the University of Hong Kong, China

Yu Xue

Department of Systems Biology, College of Life Science and Technology Huazhong University of Science and Technology, Wuhan, China

Yulan Wang

State Key Laboratory of Magnetic Resonance and Atomic and Molecular Physics, Wuhan Centre for Magnetic Resonance, Wuhan Institute of Physics and Mathematics, The Chinese Academy of Sciences , China

Zhengwei Yuan

The key laboratory of health ministry for congenital malformation, Shengjing Hospital, China Medical University

Zhiqiang Gao

Department of Chemistry, National University of Singapore

AUSTRALIA AND NEW ZEALAND

Bruno Catimel

Epithelial laboratory, Ludwig Institute for Cancer Research, Post Office Royal Melbourne Hospital, Australia

Daniel Cozzolino

Barley Research Laboratory, School of Agriculture, Food and Wine, University of Adelaide, Australia

David Beale

CSIRO Land and Water, Hightett, Australia

Emad Kiriakous

Queensland University of Technology (QUT), Brisbane, Australia

Joëlle Coumans-Moens

School of Science and Technology, School of Medicine, University of New England, Australia

Marc Wilkins

University of New South Wales, Sydney, Australia

Maurizio Ronci

Mawson Institute, University of South Australia, Mawson Lakes, Australia

Michelle Hill

University of Queensland, Australia

Michelle Colgrave

CSIRO Livestock Industries, St Lucia, Australia

Nicolas Taylor

ARC Centre of Excellence in Plant Energy Biology & Centre for Comparative Analysis of Biomolecular Networks (CABiN), University of Western Australia, Perth, Australia

Peter Hoffmann

Institute for Photonics & Advanced Sensing (IPAS), School of Chemistry and Physics, University of Adelaide, Australia

Stefan Clerens

Protein Quality &Function, AgResearch Ltd Christchurch, New Zealand

Peter Solomon

Research School of Biology College of Medicine, Biology and Environment, Australian National University, Australia

Phoebe Chen

Department of Computer Science and Computer Engineering, La Trobe University, Melbourne, Australia

Richard Christopherson

School of Molecular Bioscience, University of Sydney, Australia

Sham Nair

Department of Biological Sciences, Faculty of Science, Macquarie University, NSW, Australia

Sylvia Urban

School of Applied Sciences (Discipline of Applied Chemistry), RMIT University, Melbourne, Victoria, Australia

Valerie Wasinger

Bioanalytical Mass Spectrometry Facility, Mark Wainwright Analytical Centre, University of NSW, Australia

Wujun Ma

Centre for Comparative Genomics, Murdoch University, Australia

Yin Xiao

Institute of Health and Biomedical Innovation, Queensland University of Technology, Australia

EUROPE

AhmetKoc, PhD

Izmir Institute of Technology, Department of Molecular Biology & Genetics, Urla, Izmir, Turkey

Alejandro Gella

Department of Basic Sciences, Neuroscience Laboratory, Faculty of Medicine and Health Sciences, Universitat Internacional de Catalunya, Sant Cugat del Vallès-08195, Barcelona, Spain

Alessandro Pessione

Università degli Studi di Torino, Italy

Alexander Scherl

Proteomics Core Facility, Faculty of Medicine, University of Geneva, Geneva, Switzerland

Alfio Ferlito

ENT Clinic, University of Udine, Italy

Almudena Fernández Brieria

Dpt. Biochemistry Genetics and Immunology, Faculty of Biology –University of Vigo, Spain

Alfonsina D'Amato

Politecnico di Milano, Department of Chemistry, Materials and Chemical Engineering "GiulioNatta", Italy

Alfred Vertegaal

Molecular Cell Biology, Leiden University Medical Center, The Netherlands

Ali Mobasheri

School of Veterinary Medicine and Science, Faculty of Medicine and Health Sciences, University of Nottingham, Sutton Bonington Campus, Sutton Bonington, Leicestershire, United Kingdom

Andre Almeida

Instituto de Tecnologia Química e Biológica, Universidade Nova de Lisboa, Portugal

Andrea Matros

Leibniz Institute of Plant Genetics and Crop Plant Research (IPK-Gatersleben), Gatersleben, Germany

Andrei Turtoi

University of Liege, Metastasis Research Laboratory, GIGA-Cancer Bât. B23, Belgium

Angelo D'Alessandro

Università degli Studi della Tuscia, Department of Ecological and Biological Sciences, Viterbo, Italy

Angelo Izzo

Department of Experimental Pharmacology, University of Naples Federico II, Naples, Italy

Antonio Gnani

Department of Medical Basic Sciences, University of Bari "Aldo Moro", Bari, Italy

Ana Maria Rodríguez-Piñeiro

Institute of Biomedicine, University of Gothenburg, Sweden

Ana Varela Coelho

Instituto de Tecnologia Química e Biológica (ITQB) Universidade Nova de Lisboa (UNL), Portugal

Anna Maria Timperio

Dipartimento Scienze Ambientali Università della Tuscia Viterbo, Italy

André Nogueira Da Costa

Molecular Carcinogenesis Group, Section of Mechanisms of Carcinogenesis International Agency for Research on Cancer - World Health Organization (IARC-WHO), Lyon, France

Andreas Boehm

Steigerfurtweg 8a, D-97084 Würzburg, Germany

Andrea Scaloni

Proteomics and Mass Spectrometry Laboratory, ISPAAM, National Research Council, via Argine 1085, 80147 Napoli, Italy

Andreas Tholey

Division for Systematic Proteome Research, Institute for Experimental Medicine, Christian-Albrechts-University, Germany

Angel Manteca

Departamento de Biología Funcional and IUBA, Facultad de Medicina, Universidad de Oviedo, Spain

Angel P. Diz

Department of Biochemistry, Genetics and Immunology, Faculty of Biology, University of Vigo, Spain

Angela Bachi

Mass Spectrometry Unit DIBIT, San Raffaele Scientific Institute, Milano, Italy

Angela Chambery

Department of Life Science, Second University of Naples, Italy

Anna-Irini Koukkou

University of Ioannina, Department of Chemistry, Biochemistry Laboratory, Greece

António Sebastião Rodrigues

Departamento de Genética, Faculdade de Ciências Médicas, Universidade Nova de Lisboa, Portugal

Arkadiusz Kosmala

Laboratory of Cytogenetics and Molecular Biology, Institute of Plant Genetics, Polish Academy of Sciences, Poland

Arzu Umar

Department of Medical Oncology, Laboratory of Breast Cancer Genomics and Proteomics, Erasmus Medical Center Rotterdam Josephine Nefkens Institute, Rotterdam, The Netherlands

Baggerman Geert

ProMeta, Interfaculty Center for Proteomics and Metabolomics, Leuven,

Belgium

Bart De Spiegeleer

Ghent University, Belgium

Bart Devreese

Laboratory for Protein Biochemistry and Biomolecular Engineering,
Department for Biochemistry and Microbiology, Ghent University, Belgium

Bernard Corfe

Department of Oncology, University of Sheffield, Royal Hallamshire Hospital,
United Kingdom

Bernd Thiede

Biotechnology Centre of Oslo, University of Oslo, Blindern, Norway

Björn Meyer

Institut für Instrumentelle Analytik und Bioanalytik Hochschule Mannheim,
Germany

Bruno Baudin

Biochemistry Laboratory A, Saint-Antoine Hospital, Hôpitaux Universitaires
Est Parisien-APHP, Paris, France

Bruno Manadas

Center for Neuroscience and Cell Biology, University of Coimbra, Portugal

Cândido Pinto Ricardo

Instituto de Tecnologia Química e Biológica, Universidade Nova de Lisboa, Av.
da República-EAN, 2780-157 Oeiras, Portugal

Carla Pinheiro

Plant Sciences Division, Instituto de Tecnologia Química e Biológica (ITQB),
Universidade Nova de Lisboa, Portugal

Claudia Desiderio

Consiglio Nazionale delle Ricerche, Istituto di Chimica del Riconoscimento
Molecolare (UOS Roma), Italy

Claudio De Pasquale

SAG Department, University of Palermo, Italy

Carlos Gutiérrez Merino

Dept. Biochemistry and Molecular Biology University of Extremadura, Badajoz,
Spain

Cecilia Calado

Engineering Faculty Catholic University of Portugal, Rio de Mouro, Portugal

Celso Reis

Institute of Molecular Pathology and Immunology of the University of Porto,
IPATIMUP, Portugal

Celso Vladimiro Cunha

Medical Microbiology Department, Institute of Hygiene and Tropical
Medicine, New University of Lisbon, Portugal

Charles Steward

The Wellcome Trust Sanger Institute, Hinxton, United Kingdom

Chris Goldring

Department of Pharmacology and Therapeutics, MRC Centre for Drug Safety
Science, University of Liverpool, United Kingdom

Christian Lindermayr

Institute of Biochemical Plant Pathology, Helmholtz Zentrum München,
German Research Center for Environmental Health, Neuherberg, Germany

Christiane Fæste

Section for Chemistry and Toxicology Norwegian Veterinary Institute, Oslo,
Norway

Christer Wingren

Department of Immunotechnology, Lund University, Lund, Sweden

Christophe Cordella

UMR1145 INRA, Laboratoire de Chimie Analytique, Paris, France

Christophe Masselon

Laboratoire de Biologie a Grande Echelle (iRTSV/BGE), CEA Grenoble, France

Cosima Damiana Calvano

Universita' degli Studi di Bari, Dipartimento di Chimica, Bari, Italy

David Cairns

Section of Oncology and Clinical Research, Leeds Institute of Molecular

Medicine, Leeds, UK

Daniela Cecconi

Dip. diBiotecnologie, LaboratoriodiProteomica e Spettrometriadi Massa,
Universitàdi Verona, Verona, Italy

David Honys

Laboratory of Pollen Biology, Institute of Experimental Botany ASCR, Czech
Republic

David Sheehan

Dept. Biochemistry, University College Cork (UCC), Ireland

Deborah Penque

Departamento de Genética, Instituto Nacional de Saúde Dr Ricardo Jorge
(INSA, I.P.), Lisboa, Portugal

Dilek Battal

Mersin University, Faculty of Pharmacy, Department of Toxicology, Turkey

Domenico Garozzo

CNR ICTP, Catania, Italy

Ed Dudley

Institute of Mass Spectrometry, College of Medicine Swansea University,
Singleton Park, Swansea, Wales, UK

Edoardo Saccenti

University of Amsterdam, Netherlands Metabolomics Centre, The Netherlands

Elena Gonzalez

Complutense University of Madrid, Dept. Biochemistry and Molecular Biology
IV, Veterinary Faculty, Madrid, Spain

Elia Ranzato

Dipartimento di Scienze e Innovazione Tecnologica, DiSIT, University of
Piemonte Orientale, Alessandria, Italy

Elisa Bona

Università del Piemonte Orientale, DISIT, Alessandria, Italy

Elke Hammer

Interfaculty Institute for Genetics and Functional Genomics, Ernst-Moritz-
Arndt Universität, Germany

Enrica Pessione

University of Torino, Life Sciences and Systems Biology Department, Torino,
Italy

Eva Rodríguez Suárez

Proteomics Core Facility - CIC bioGUNE, Parque tecnologico de Bizkaia, Spain

Federica Pellati

Department of Life Sciences, University of Modena and Reggio Emilia, Italy

Ferdinando Cerciello

Laboratory of Molecular Oncology, Clinic of Oncology, University Hospital
Zürich, Switzerland

Fernando J. Corrales

Division of Hepatology and Gene Therapy, Proteomics Unit, Center for
Applied Medical Research (CIMA), Pamplona, Spain

Florian Szabados

Dept. of Medical Microbiology, Ruhr-University Bochum, Germany

Francesco Salii

University of Milano Bicocca, Italy

Francisco J Blanco

Platform of Proteomics, Proteo-Red-ISCHII INIBIC-Hospital Universitario A
Coruña, Spain

Francisco Javier Fernández Acero

Laboratory of Microbiology, Marine and Environmental Sciences Faculty,
University of Cádiz, Pol. Río San Pedro s/n, Puerto Real, Cádiz, Spain

Francisco Torrens

InstitutUniversitari de CiènciaMolecular, Universitat de València, Spain

François Fenaille

CEA, IBiTecS, Service de Pharmacologie et DImmunoanalyse (SPI), France

Frederic Silvestre

University of Namur, Belgium

Fulvio Magni

Department of Health Science, Monza, Italy

Georgios Theodoridis

Department of Chemistry, Aristotle University, Greece

Germain Rousselet

Laboratoire Réparation et Transcription dans les cellules Souches (LRTS), CEA/DSV/IRCM, Fontenay aux Roses, France

German Bou

Servicio de Microbiología-INIBIC, Complejo Hospitalario Universitario La Coruña, Spain

Gianfranco Mamone

Proteomic and Biomolecular Mass Spectrometry Centre, Institute of Food Science CNR, Italy

Gianfranco Romanazzi

Department of Environmental and Crop Sciences, Marche Polytechnic University, Italy

Gianluigi Mauriello

Department of Food Science, University of Naples Federico II Naples, Italy

Giorgio Valentini

Università degli Studi di Milano, Dept. of Computer Science, Italy

Giuseppe Palmisano

Department of Biochemistry and Molecular Biology
University of Southern Denmark, Odense M, Denmark

Helen Gika

Chemical Engineering Department, Aristotle University of Thessaloniki, Greece

Hugo Miguel Baptista Carreira dos Santos

REQUIMTE-FCT Universidade NOVA de Lisboa, Portugal

Ignacio Casal

Functional Proteomics Laboratory, Centro de Investigaciones Biológicas (CSIC), Madrid, Spain

Ignacio Ortea

European Commission, Joint Research Center, Institute for Reference Materials and Measurements, Geel, Belgium

Iñaki Álvarez

Institut de Biotecnologia i Biomedicina Vicent Villar Palasí, Universitat Autònoma de Barcelona, Barcelona

Isabel Marcelino

Instituto de Tecnología Química e Biológica, Oeiras, Portugal

Isabel Liste

Area de Biología Celular y del Desarrollo, Instituto de Salud Carlos III, Madrid, Spain

Isabelle Fournier

University Lille Nord de France, Fundamental & Applied Biological Mass Spectrometry - EA 4550, Villeneuve d'Ascq, France

Jacek Z. Kubiak

CNRS UMR 6061, University of Rennes 1, Institute of Genetics and Development of Rennes, Rennes, France

Jane Thomas-Oates

Centre of Excellence in Mass Spectrometry and Department of Chemistry, University of York, Heslington, UK

Jatin Burniston

Muscle Physiology and Proteomics Laboratory, Research Institute for Sport and Exercise Sciences, Liverpool John Moores University, Tom Reilly Building, Liverpool, United Kingdom

Jean-Paul Issartel

INSERM U836, Grenoble Institut des Neurosciences, La Tronche, France

Jens Allmer

Molecular Biology and Genetics, Izmir Institute of Technology, Urla, Izmir, Turkey

Jerry Thomas

Technology Facility, Department of Biology, University of York, UK

Jesús Jorrín Novo

Agricultural and Plant Biochemistry, Proteomics Research Group, Department of Biochemistry and Molecular Biology, Córdoba, Spain

Jesus Mateos Martín

Osteoarticular and Aging Research Lab, Proteomics Unit INIBIC-Complejo Hospitalario Universitario de A Coruña, A Coruña, Spain

Joan Cerdà

Laboratory IRTA, Institute of Marine Sciences (CSIC), Passeig marítim 37-49, 08003 Barcelona, Spain

Joan Claria

Department of Biochemistry and Molecular Genetics, Hospital Clínic of Barcelona, Spain

João Rodrigues

Instituto de Higiene e Medicina Tropical, Universidade Nova de Lisboa, Portugal

Joaquim ROS

Dept. Ciències Mèdiques Bàsiques. IRB Lleida. University of Lleida, Spain

Joerg Reinders

AG Proteomics, Institute of Functional Genomics, University Regensburg, Germany

Johan Palmfeldt

Research Unit for Molecular Medicine, Aarhus University Hospital, Skejby, Aarhus, Denmark

Jose Andrés Fernández González

Universidad del País Vasco, Facultad de Ciencia y Tecnología, Spain

Jose Câmara

University of Madeira, Funchal, Portugal

Jose Cremata Alvarez

Department of Carbohydrate Chemistry, Center for Genetic Engineering and Biotechnology, Havana, Cuba

Jose Luis Martín-Ventura

IIS-FJD-UAM, Madrid, Spain

José Manuel Bautista

Departamento de Bioquímica y Biología Molecular IV, Universidad Complutense de Madrid, Spain

Jose Manuel Palma

Departamento de Bioquímica, Biología Celular y Molecular de Plantas Estación Experimental del Zaidín, CSIC, Granada, Spain

José Moreira

Danish Center for Translational Breast Cancer Research, Denmark

Juraj Gregan

Max F. Perutz Laboratories, University of Vienna, Austria

Karin Stensjö

Department of Photochemistry and Molecular Science, Ångström laboratory, Uppsala University, Sweden

Kathleen Marchal

CMPG/Bioinformatics, Dep Microbial and Molecular Systems, Leuven, Germany

Kay Ohlendieck

Department of Biology, National University of Ireland, Maynooth, Co. Kildare, Ireland

Keiryn Bennett

CeMM - Center for Molecular Medicine of the Austrian Academy of Sciences Vienna, Austria

Kjell Sergeant

Centre de Recherche Public-Gabriel Lippmann, Department 'Environment and Agro-biotechnologies' (EVA), Luxembourg

Konstantinos Kouremenos

Department of Chemistry, Umea University, Sweden

Lennart Martens

Department of Medical Protein Research, VIB and Department of Biochemistry, Ghent University, Belgium

Luis P. Fonseca

Instituto Superior Técnico, Centro de Engenharia Biológica e Química,
Institute for Biotechnology and Bioengineering, Lisboa, Portugal

Luisa Brito

Laboratório de Microbiologia, Instituto Superior de Agronomia, Tapada da
Ajuda, Lisbon, Portugal

Luisa Mannina

CNR, Istituto di Metodologie Chimiche, Rome, Italy

Manuel Avilés Sanchez

Department of Cell Biology and Histology, School of Medicine, University of
Murcia, Spain

Mar Vilanova

Misión Biológica de Galicia, Consejo Superior de Investigaciones Científicas,
Pontevedra, Spain

Marcello Donini

ENEA -Casaccia Research Center, UTBIORAD-FARM, Biotechnology
Laboratory, Italy

Marco Lemos

GIRM & ESTM - Polytechnic Institute of Leiria, Peniche, Portugal

Marcus Mau

King's College London, UK

Maria Álava

Departamento de Bioquímica y Biología Molecular y Celular, Facultad de
Ciencias, Universidad de Zaragoza, Spain

Maria De Angelis

Department of Soil, Plant and Food Science, University of Bari Aldo Moro, Italy

María de la Fuente

Legume group, Genetic Resources, Mision Biologica de Galicia-CSIC,
Pontevedra, Spain

Maria M. Malagón

Department of Cell Biology, Physiology and Immunology, IMIBIC,
Universidad de Córdoba, Spain

Maria Gabriela Rivas

REQUIMTE/CQFB, Departamento de Química, Faculdade de Ciências e
Tecnologia, Universidade Nova de Lisboa, Portugal

María Mayán

INIBIC, LaCoruña, Spain

María Páez de la Cadena

Department of Biochemistry, Genetics and Immunology, University of Vigo,
Spain

Marie Arul

Muséum National Histoire Naturelle, Département RDDM, Plateforme de
spectrométrie de masse et de protéomique, Paris, France

Marie-Pierre Bousquet

Institut de Pharmacologie et de Biologie Structurale, UPS/CNRS, Toulouse,
France

Mario Diniz

Dept. Química-REQUIMTE, Faculdade de Ciências e Tecnologia, Universidade
Nova de Lisboa, Portugal

Mark Davey

Catholic University of Leuven (KU Leuven), Belgium

Marko Radulovic

Institute for Oncology and Radiology, Laboratory of Cancer Cell biology,
Belgrade, Serbia

Martin Hajduch

Department of Reproduction and Developmental Biology, Institute of Plant
Genetics and Biotechnology, Slovak Academy of Sciences, Nitra, Slovakia

Martin Kussmann

Faculty of Science, Aarhus University, Aarhus, Denmark

Martina Marchetti-Deschmann

Institute of Chemical Technologies and Analytics, Vienna University of
Technology, Vienna, Austria

Maxence Wisztorski

University Lille 1, Laboratoire de Spectrométrie de Masse Biologique,
Fondamentale & Appliquée, Villeneuve d'Ascq, France

Meri Hovsepian

Institute of Molecular Biology of Armenian National Academy of Sciences
Yerevan, Armenia

Michalis Nikolaidis

Department of Physical Education and Sports Science at Serres, Aristotle
University of Thessaloniki, Greece

Michel Jaquinod

Exploring the Dynamics of Proteomes/Laboratoire Biologie à Grande Echelle,
Institut de Recherches en Technologies et Sciences pour le Vivant, Grenoble,
France

Michel Salzet

Laboratoire de Spectrométrie de Masse Biologique Fondamentale et Appliquée,
INSERM, Villeneuve d'Ascq, France

Miguel Reboiro Jato

Escuela Superior de Ingeniería Informática, Ourense, Spain

Moncef Mrabet

Laboratory of Legumes (LL), Centre of Biotechnology of Borj-Cédria (CBBC),
Hammam-Lif, Tunisia

Mónica Botelho

Centre for the study of animal sciences (CECA)/ICETA, Porto, Portugal

Monica Carrera

Institute of Molecular Systems Biology, Zurich, Germany

Okay Saydam

Molecular Oncology Laboratory, Division of Neuro-Oncology, Department of
Pediatrics Medical University of Vienna, Austria

Ola Söderberg

Department of Immunology, Genetics and Pathology, Uppsala University,
Sweden

Paloma Sánchez-Bel

Dpto. Biología del estrés y Patología vegetal, CEBAS-CSIC, Murcia, Spain

Pantelis Bagos

Department of Computer Science and Biomedical Informatics, University of
Central Greece, Greece

Paolo Destefanis

Department of Urology, "San Giovanni Battista - Molinette" Hospital, Turin,
Italy

Pasquale Vito

Università del Sannio, Benevento, Italy

Patrice Francois

Genomic Research Laboratory, Service of Infectious Diseases, Department of
Internal Medicine, Geneva

Patrícia Alexandra Curado Quintas Dinis Poeta

University of Trás-os-Montes and Alto Douro (UTAD), School of Agrary and
Veterinary Sciences, Veterinary, Science Department, Portugal

Paul Cutler

F Hoffman La Roche, Basel, Switzerland

Paulo Vale

IPMA - Instituto Português do Mar e da Atmosfera, Lisboa, Portugal

Pedro Baptista

Centre for Research in Human Molecular Genetics, Department of
LifeSciences, Faculdade de Ciências e Tecnologia, Universidade Nova de
Lisboa, Caparica, Portugal

Pedro Rodrigues

Centro de Ciências do Mar do Algarve, CCMAR, Faro, Portugal

Pedro Santos

CBMA-Centre of Molecular and Environmental Biology, Department of
Biology, University of Minho, Braga, Portugal

Pedro S. Lazo

Departamento de Bioquímica y Biología Molecular, Instituto Universitario de
Oncología Del Principado de Asturias (IUOPA), Universidad de Oviedo, Spain

Per Bruheim

Department of Biotechnology, Norwegian University of Science and Technology, Trondheim, Norway

Phillip Cash

Division of Applied Medicine, University of Aberdeen, Scotland

Philipp Hess

Institut Universitaire Mer et Littoral(CNRS - Université de Nantes - Ifremer), Nantes, France

Philippe Castagnone-Sereno

Interactions Biotiques et Sante Vegetale, Sophia Antipolis cedex, France

Pierscionek Barbara

School of Biomedical Sciences, University of Ulster, Cromore Road, Coleraine, BT52 1SA, United Kingdom

Pieter de Lange

DipartimentodiScienzedellaVita, SecondaUniversità degli Studi di Napoli, Caserta, Italy

Qi Zhu

Dept. Electrical Engineering, ESAT/SCD, Katholieke Universiteit Leuven, Heverlee, Belgium

Ralph Fingerhut

University Children`s Hospital, Swiss Newborn Screening Laboratory, Children`s Research Center, Zürich, Switzerland

Ralf Hoffmann

Institute of Bioanalytical Chemistry, Center for Biotechnology and Biomedicine, Faculty of Chemistry and Mineralogy, Leipzig University, Germany

Rawi Ramautar

Leiden/Amsterdam Center for Drug Research, Leiden University, The Netherlands

Ricardo Gutiérrez Gallego

Bioanalysis Group, Neuropsychopharmacology Program IMIM-Hospital del Mar & Department of Experimental and Health Sciences, University Pompeu Fabra, Spain

Roman Zubarev

Department of Medical Biochemistry and Biophysics, Karolinska Institutet, Stockholm, Sweden

Roque Bru Martinez

Plant Proteomics and Functional Genomics Group, Department of Agrochemistry and Biochemistry, Faculty of Sciences, Alicante University, Spain

Rubén Armañanzas

Computational Intelligence Group, Departamento de Inteligencia Artificial, Universidad Politécnica de Madrid, Spain

Ruddy Wattiez

Department of Proteomics and Microbiology, University of Mons (UMONS), Belgium

Rune Matthiesen

Institute of Molecular Pathology and Immunology, University of Porto, Portugal

Ruth Birner-Gruenberger

Medical University Graz, Austria

Sabine Luthje

University of Hamburg, Biocenter Klein Flottbek, Hamburg, Germany

Sadin Özdemir

Department of Biology, Faculty of Science and Arts, Siirt University, Turkey

Salvador Ventura

Institut de Biociències i de Biomedicina, Universitat Autònoma de Barcelona, Spain

Sandra Kraljevic-Pavelic

University of Rijeka, Department of Biotechnology, Croatia

Sebastian Galuska

Institute of Biochemistry, Faculty of Medicine, Justus-Liebig-University of

Giessen, Germany

Serge Cosnier

Department of Molecular Chemistry, Grenoble university/CNRS, Grenoble, France

Serhat Döker

Cankiri Karatekin University, Chemistry Department, Cankiri, Turkey

Shan He

Centre for Systems Biology, School of Biosciences and School of Computer Science, University of Birmingham, England

Silvia Mazzuca

Plan Cell Physiology Laboratory, Department of Ecology, University of Calabria, Italy

Simona Martinotti

Dipartimento di Scienze e Innovazione Tecnologica, DiSIT, University of Piemonte Orientale, Alessandria, Italy

Soile Tapio

Helmholtz Zentrum München, German Research Center for Environmental Health, Institute of Radiation Biology, Neuherberg, Germany

Sophia Kossida

Biomedical Research Foundation, Academy of Athens, Department of Biotechnology, Athens, Greece

Spiros D. Garbis

Biomedical Research Foundation of the Academy of Athens, Center for Basic Research - Division of Biotechnology, Greece

Steeve Thany

Laboratoire Récepteurs et Canaux Ioniques Membranaires, UFR Science, Université d'Angers, France

Stefania Orrù

University of Naples Parthenope, Naples, Italy

Stefanie Hauck

Research Unit Protein Science, Helmholtz Center Munich, Neuherberg, Germany

Stefano Curcio

Department of Engineering Modeling, Laboratory of Transport Phenomena and Biotechnology University of Calabria, Italy

Susana Cristóbal

Department of Clinical and Experimental Medicine Faculty of Health Science Linköping University, Sweden

Tàmar García Barrera

Departamento de Química y Ciencia de los Materiales, Facultad de Ciencias Experimentales, Universidad de Huelva, Spain

Theodore Alexandrov

University of Bremen, Center for Industrial Mathematics, Germany

Thole Züchner

Ultrasensitive Protein Detection Unit, Leipzig University, Center for Biotechnology and Biomedicine, Institute of Bioanalytical Chemistry, Germany

Tiziana Bonaldi

Department of Experimental Oncology, European Institute of Oncology, Via Adamello 16, 20139 Milan, Italy

Tomris Ozben

Akdeniz University Medical Faculty Department of Clinical Biochemistry, Antalya, Turkey

Tsangaris George

Proteomics Research Unit, Center of Basic Research II Foundation of Biomedical Research of the Academy of Athens, Greece

Üner Kolukisaoglu

Center for Plant Molecular Biology, Eberhard Karls University Tübingen, Tübingen, Germany

Valeria Bertagnolo

Department of Morphology and Embryology University of Ferrara, Italy

Vera Muccilli

DipartimentodiScienzeChimiche, UniversitàdiCatania, Catania, Italy

Veronica Mainini
Dept. Health Science, University of Milano-Bicocca, Faculty of Medicine,
Monza (MB), Italy
Vicenta Martínez-Zorzano
Department of Biochemistry, Genetics and Immunology
University of Vigo, Spain
Virginie Brun
French Atomic Energy Commission and *French National Institute for Health*
and Medical Research, France
Vittoria Matafora

Biological Mass Spectrometry Unit, San Raffaele Scientific Institute, Milan, Italy
Vladislav Khrustalev
Department of General Chemistry, Belarussian, State Medical University,
Dzerzhinskogo, Minsk, Belarus
Xiaozhe Zhang
Department of Medicine, University of Frioburg, Switzerland
Yuri van der Burgt
Leiden University Medical Center, Department of Parasitology, The
Netherlands

SOUTH AMERICA

Alessandro Farias
Neuroimmunomodulation Group, department of Genetics, Evolution and
Bioagents, University of Campinas - SP - Brazil
Alexandra Sawaya
Department of Plant Biology, Institute of Biology, UNICAMP, Campinas, São
Paulo, Brazil
Andréa P.B. Gollucke
Hexalab/Catholic University of Santos, Brazil
Arlindo Moura
Department of Animal Science - College of Agricultural Sciences - Federal
University of Ceara, Fortaleza, Brasil
Bruno Lomonte
Instituto Clodomiro Picado, Universidad de Costa Rica
Deborah Schechtman
Department of Biochemistry, Chemistry Institute, University of São Paulo,
Brazil
Edson Guimarães Lo Turco
São Paulo Federal University, Brasil
Elisabeth Schwartz
Department of Physiological Sciences, Institute of Biological Sciences,
University of Brasilia, Brazil
Fabio Ribeiro Cerqueira
Department of Informatics and NuBio (Research Group for Bioinformatics),
University of Vicosa, Brazil
Fernando Barbosa
Faculty of Pharmaceutical Sciences of Ribeirão Preto University of São Paulo,
Brazil
Hugo Eduardo Cerecetto
Grupo de Química Medicinal, Facultad de Química, Universidad de la
República, Montevideo, Uruguay
Luis Pacheco
Institute of Health Sciences, Federal University of Bahia, Salvador, Brazil

Mário Hiroyuki Hirata
Laboratório de Biologia Molecular Aplicado ao Diagnóstico, Departamento de
Análises Clínicas e Toxicológicas, Faculdade de Ciências Farmacêuticas,
Universidade de São Paulo, Brazil
Jan Schripsema
Grupo Metabólica, Laboratório de Ciências Químicas, Universidade
Estadual do Norte Fluminense, Campos dos Goytacazes, Brazil
Jorg Kobarg
Centro Nacional de Pesquisa em Energia e Materiais, Laboratório Nacional de
Biotecnologia, Brazil
Marcelo Bento Soares
Cancer Biology and Epigenomics Program, Children's Memorial Research
Center, Professor of Pediatrics, Northwestern University's Feinberg School of
Medicine
Mario Palma
Center of Study of Social Insects (CEIS)/Dept. Biology, Institute of Biosciences,
Univesity of São Paulo State (UNESP), Rio Claro - SP Brazil
Rinaldo Wellerson Pereira
Programa de Pós Graduação em Ciências Genômicas e Biotecnologia,
Universidade Católica de Brasília, Brazil
Roberto Bobadilla
BioSigma S.A., Santiago de Chile, Chile
Rossana Arroyo
Department of Infectomic and Molecular Biology, Center of Research and
Advanced Studies of the National, Polytechnical Institute (CINVESTAV-IPN),
Mexico City, Mexico
Rubem Menna Barreto
Laboratorio de Biologia Celular, Instituto Oswaldo Cruz, Fundação Oswaldo
Cruz, Rio de Janeiro, Brazil
Vasco Azevedo
BiologicalSciencesInstitute, Federal University of Minas Gerais, Brazil

NORTH AMERICA

Adam Vigil
University of California, Irvine, USA
Akeel Baig
Hoffmann-La Roche Limited, Pharma Research Toronto, Toronto, Ontario,
Canada
Alexander Statnikov
Center for Health Informatics and Bioinformatics, New York University School
of Medicine, New York
Amosy M'Koma
Vanderbilt University School of Medicine, Department of General Surgery,
Colon and Rectal Surgery, Nashville, USA
Amrita Cheema
Georgetown Lombardi Comprehensive Cancer Center, USA
Anthony Gramolini

Department of Physiology, Faculty of Medicine, University of Toronto, Canada
Anas Abdel Rahman
Department of Chemistry, Memorial University of Newfoundland and
Labrador St. John's, Canada
Christina Ferreira
Purdue University - Aston Laboratories of Mass Spectrometry, Hall for
Discovery and Learning Research, West Lafayette, US
Christoph Borchert
Biochemistry & Microbiology, University of Victoria, UVic Genome British
Columbia Proteomics Centre, Canada
Dajana Vuckovic
University of Toronto, Donnelly Centre for Cellular + Biomolecular Research,
Canada
David Gibson

University of Colorado Denver, Anschutz Medical Campus, Division of Endocrinology, Metabolism and Diabetes, Aurora, USA

Deyu Xie

Department of Plant Biology, Raleigh, USA

Edgar Jaimes

University of Alabama at Birmingham, USA

Eric McLamore

University of Florida, Agricultural & Biological Engineering, Gainesville, USA

Eustache Paramithiotis

Caprion Proteomics Inc., Montreal, Canada

FangXiang Wu

University of Saskatchewan, Saskatoon, Canada

Fouad Daayf

Department of Plant Science, University of Manitoba, Winnipeg, Manitoba, Canada

Haitao Lu

Washington University School of Medicine, Saint Louis, USA

Hexin Chen

University of South Carolina, Columbia, USA

Hsiao-Ching Liu

232D Polk Hall, Department of Animal Science, North Carolina State University Raleigh, USA

Hui Zhang

Johns Hopkins University, MD, USA

Ing-Feng Chang

Institute of Plant Biology, National Taiwan University, Taipei, Taiwan

Irwin Kurland

Albert Einstein College of Medicine, Associate Professor, Dept of Medicine, USA

Jagjit Yadav

Microbial Pathogenesis and Toxicogenomics, Laboratory, Environmental Genetics and Molecular, Toxicology Division, Department of Environmental Health, University of Cincinnati College of Medicine, Ohio, USA

Jianbo Yao

Division of Animal and Nutritional Sciences, USA

Jiaxu Li

Department of Biochemistry and Molecular Biology, Mississippi State University, USA

Jiping Zhu

Exposure and Biomonitoring Division, Health Canada, Ottawa, Canada

Jiri Adamec

Department of Biochemistry & Redox Biology Center, University of Nebraska, Lincoln Nebraska, USA

Jiye Ai

University of California, Los Angeles

John McLean

Department of Chemistry, Vanderbilt University, Nashville, TN, USA

Joshua Heazlewood

Lawrence Berkeley National Laboratory, Berkeley, CA, USA

Kenneth Yu

Memorial Sloan Kettering Cancer Center, New York, USA

Laszlo Prokai

Department of Molecular Biology & Immunology, University of North Texas Health Science Center, Fort Worth, USA

Lei Li

University of Virginia, USA

Leonard Foster

Centre for High-throughput Biology, University of British Columbia, Vancouver, BC, Canada

Madhulika Gupta

Children's Health Research Institute, University of Western Ontario London, ON, Canada

Masaru Miyagi

Case Center for Proteomics and Bioinformatics, Case Western Reserve University, Cleveland, USA

Michael H.A. Roehrl

Department of Pathology and Laboratory Medicine, Boston Medical Center Boston, USA

Ming Zhan

National Institute on Aging, Maryland, USA

Nicholas Seyfried

Emory University School of Medicine, Atlanta, USA

Olgica Trenchevska

Molecular Biomarkers, Biodesign Institute at Arizona State University, USA

Peter Nemes

US Food and Drug Administration (FDA), Silver Spring, USA

R. John Solaro

University of Illinois College of Medicine, USA

Rabih Jabbour

Science Application International Corporation, Maryland, USA

Ramesh Katam

Plant Biotechnology Lab, Florida A and M University, FL, USA

Robert L. Hettich

Chemical Sciences Division, Oak Ridge National Laboratory, Oak Ridge, USA

Robert Powers

University of Nebraska-Lincoln, Department of Chemistry, USA

Shen S. Hu

UCLA School of Dentistry, Dental Research Institute, UCLA Jonsson Comprehensive Cancer Center, Los Angeles CA, USA

Shiva M. Singh

University of Western Ontario, Canada

Susan Hester

United States Environmental Protection Agency, Durnam, USA

Terry D. Cyr

Genomics Laboratories, Centre for Vaccine Evaluation, Biologics and Genetic Therapies Directorate, Health Products and Foods Branch, Health Canada, Ontario, Canada

Thibault Mayor

Department of Biochemistry and Molecular Biology, Centre for High-Throughput Biology (CHiBi), University of British Columbia, Canada

Thomas Conrads

USA

Thomas Kislinger

Department of Medical Biophysics, University of Toronto, Canada

Wan Jin Jahng

Department of Biological Sciences, Michigan Technological University, USA

Wayne Zhou

Marine Biology Laboratory, Woods Hole, MA, USA

Wei Jia

US Environmental Protection Agency, Research Triangle Park, North Carolina, USA

Wei-Jun Qian

Pacific Northwest National Laboratory, USA

William A LaFramboise

Department of Pathology, University of Pittsburgh School of Medicine Shadyside Hospital, Pittsburgh, USA

Xiangjia Min

Center for Applied Chemical Biology, Department of Biological Sciences Youngstown State University, USA

Xiaoyan Jiang

Senior Scientist, Terry Fox Laboratory, BC Cancer Agency, Vancouver, Canada

Xu-Liang Cao

Food Research Division, Bureau of Chemical Safety, Health Canada, Ottawa, Canada

Xuequn Chen

Department of Molecular & Integrative Physiology, University of Michigan,
Ann Arbor, USA

Ye Fang

Biochemical Technologies, Science and Technology Division, Corning
Incorporated, USA

Ying Qu

Microdialysis Experts Consultant Service, San Diego, USA

Ying Xu

Department of Biochemistry and Molecular Biology, Institute of
Bioinformatics, University of Georgia, Life Sciences Building
Athens, GA, USA

SPECIAL ISSUE
**PROCEEDING ABSTRACTS OF THE 4th INTERNATIONAL
CONGRESS ON ANALYTICAL PROTEOMICS (ICAP 2015)**

The content published in this issue was previously selected by the scientific committee to be presented at the 4th International Congress on Analytical Proteomics, from 7 - 9 September 2015 in Caparica, Almada, Portugal. Only the Proceeding Abstracts with a copyright transfer agreement were reproduced.

4th International Congress on Analytical Proteomics (ICAP 2015)

Caparica - Almada, Portugal - 7th – 9th September 2015

Committees

Conference Chairs

José Luis Capelo Martínez

UCIBIO/REQUIMTE, Chemistry Department, University NOVA of Lisbon, Portugal, PhD

Carlos Lodeiro Espiño

UCIBIO/REQUIMTE, Chemistry Department, University NOVA of Lisbon, Portugal, PhD

Hugo Miguel Santos

UCIBIO/REQUIMTE, Chemistry Department, University NOVA of Lisbon, Portugal, PhD

Elisabete Oliveira

UCIBIO/REQUIMTE, Chemistry Department, University NOVA of Lisbon, Portugal, PhD

Scientific Committee

Patricia A. Curado Q. Dinis Poeta

Veterinary Science Department, CEVAC, University of Trás-os-Montes e Alto Douro, Portugal, PhD

Jens Coorsen

School of Medicine, University of Western Sydney, Australia, PhD

Florentino Fdez-Riverola

Informatics Department, University of Vigo, Ourense, Spain, PhD

Julia Lorenzo Rivera

Biochemistry Department, University Autònoma of Barcelona, Spain, PhD

José-Luis Capelo-Martínez

UCIBIO/REQUIMTE, Chemistry Department, University NOVA of Lisbon, Portugal, PhD

Carlos Lodeiro-Espiño

UCIBIO/REQUIMTE, Chemistry Department, University NOVA of Lisbon, Portugal, PhD

Gilberto Igrejas.

University of Trás-os-Montes and Alto Douro. Portugal, PhD

Kay Ohlendieck

Maynooth University, Ireland, PhD

Wolfgang Maret

King's College London, Metal Metabolism Group Division of Diabetes and Nutritional Sciences, London, UK, PhD

Marco Zezzi Arruda

University of Campinas, Campinas, Brasil, PhD

Conference Secretariat

Hugo Miguel Santos

FCT-UCIBIO/REQUIMTE, Chemistry Department, University NOVA of Lisbon, Portugal, PhD

Ricardo Alves

Proteomass FCT-UNL, University NOVA of Lisbon, Portugal, MsD

Tomás Miranda

Computer Science Department FCT-UNL, University NOVA of Lisbon, Portugal, MsD

JOURNAL OF INTEGRATED OMICS

A methodological Journal

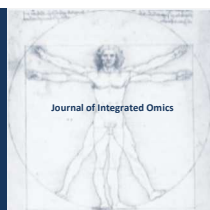
SPECIAL ISSUE: PROCEEDING ABSTRACTS OF THE 4th INTERNATIONAL CONGRESS ON ANALYTICAL PROTEOMICS (ICAP 2015)

CONTENTS OF VOLUME 5 | ISSUE 2 | DECEMBER 2015

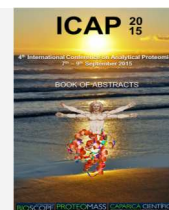
One gene, many changes: a long history involving soybean.....	1
Proteomic profiling of neuromuscular disorders.....	2
The proteomic proficiency to identify and elucidate antimicrobial resistance bacteria.....	3
Protein of interests for serological diagnosis in Bird fancier's Lung.....	4
A case report of cyclosporine complexed with human calcineurin remaining in the gut after 25 years of its consumption	5
Improved efficiency of <i>in situ</i> tryptic proteolysis for fresh frozen and FFPE tissue.....	6
Development and evaluation of method for metalloproteins detection involved in bipolar disorder	7
Using protein biomarkers to understand the biological mechanisms underlying beef sensory qualities.....	8
Some proteins of possible -clinical interest found in samples of human intestinal mucus (HIM).....	9
Proteomic alterations in transgenic and non-transgenic soybean leaves after plant cultivation in the presence of silver nanoparticles.....	10
Rice Proteomics: Method Development of 2D-PAGE for Malaysian Upland Rice	11
Effect of resistance inducers and <i>Hemileia vastatrix</i> infection on protein patterns of <i>Coffea arabica</i> leaves.....	12
A quest for innovative and robust biomarkers using MS-based proteomics	13
Matrix metalloproteinases (MMPs) in buck blood chronically infected by arthritis encephalitis virus in Northeast of Brazil	14
Identification of immunoreactive microbial proteins in the field of allergic diseases, and application for serodiagnosis.....	15
<i>Caenorhabditis elegans</i> based proteomic analysis during bacterial infections	16
Quantitative analysis of the mitochondrial proteome and phosphoproteome in the yeast <i>Saccharomyces cerevisiae</i>	17
Dynamism of protein expression in gametes and early mammalian embryos analysed by a holistic approach and multiplexed SRM.....	18
A new view of brain function: quantifying protein responses to learning and memory with reverse phase protein arrays.....	19
Dysregulation of IFN alpha and IL-10 in central nervous system in transgenic porcine Huntington's disease model.....	20

Targeted quantitative proteomics in pharmaceutical sciences.....	21
Quantitative mass spectrometric measurement of multiple serum apolipoproteins: ready for validation and clinical application.....	22
Proteomic analysis and μ -, δ - and κ -opioid receptor content in brain cortex of rats exposed to increasing doses of morphine for 10 days; comparison with animals sacrificed 20 days since the last dose of morphine	23
Advantages and disadvantages of chromatographic approaches to study bacterial metalloproteomes.....	24
Mechanisms underlying metabolic disorders deciphered using quantitative proteomics of non-sequenced species	25
Aptamers in cancer research	26
Quantitative proteomics in human drug metabolism	27
Scorpion venom proteomics and transcriptomics	28
Comparative proteomic profiling of brain tissues of rat with different behavioral characteristics during emotional stress	29
Cell-free lysates for the production of difficult proteins	30
Linear Algebra Analysis and Quantification of Arginine Dimethylation in Histone Modification using Liquid Chromatography–Tandem Mass Spectrometry-Based Targeted Proteomics	31
Integrated Proteomic and Metabolomic Approaches Reveal Energy Disruption in Epilepsy.....	32
Methods for the preparation of acute myeloid leukemia patient samples for proteomic and phosphoproteomic analysis	33
Comparison of 2D proteomic maps revealed properties of <i>Ambrosia artemisiifolia</i> sub-pollen particles accounting for more severe asthma symptoms than its whole pollen grains.....	34
Effect of a quorum sensing inhibitor on the <i>Pseudomonas aeruginosa</i> PAO1 proteome.....	35
Immune responses of silkworm, <i>Bombyx mori</i> against infection by an endoparasitoid	36
Proteomic characterization of Colorectal Cancer tissues reveals tumor stage- and EGFR-specific CRC biomarkers	37
Probing the structure and conformational changes in complex biological systems using zero-length crosslinking and mass spectrometry	38
The O ₂ -Affinity and Cooperativity of Hemoglobin Are Regulated by the Effector-Linked 4D-Structural Changes (Protein Dynamics) rather than the Ligand-Linked 3D-Structural Changes	39
Flagellar proteins and their key roles in human diseases: Once neglected, still underestimated	40
Effects of heavy metals on <i>Cyanothece</i> sp. CCY 0110 growth, extracellular polymeric substances (EPS) production, ultrastructure and protein profiles	41
Brain ubiquitin mitochondrial subproteome and its changes induced by neurotoxins and neuroprotectors	42
The Resonant Recognition model to design short bioactive therapeutic peptides: does it really work?.....	43
Knots in proteins – a tangled tale	44
Comparative proteomic profile between transgenic and non-transgenic soybean seeds by 2D-PAGE	45
Comparative evaluation of extraction methods for UPLC-ESI-QTOF MS analysis of lipids from human blood serum	46
Differential protein expression in the testes of ganders under different monochromatic light sources.....	47

Proteomic comparative study of smooth muscle cells isolated from systemic and pulmonary arteries	48
Protein expression profiles of serum samples bipolar disorder patients mapped by 2-D DIGE coupled with nanoLC-MS/MS..	49
Western blotting - detection of a specific protein in MCF7 and MCF7-Z tissue	50
The quantitative proteomic study of human saliva samples obtained from caries-free and caries-susceptible people.....	51
The quantitative comparison of the human tooth pulp obtained from caries-free and caries-susceptible people.....	52
<i>In vivo</i> determination of the CYP4A and COX5A expression and catalytic activity in rat hepatic microsomes after drug administration	53
Label free proteomics from rat liver microsomes in drug dependence.....	54
Sodium effect on agonist binding to δ -opioid receptors in isolated plasma membranes from forebrain cortex of rats exposed to increasing doses of morphine (10 - 50 mg/kg) for 10 days.....	55
Do or do not. There is no try.....	56
Proteomics analysis of the peritoneal dialysis effluent: a longitudinal study	57
Differential expression of egg white proteins in different breeds of chickens and their association with physical characteristics of hard-boiled eggs	58
Methods for the preparation of acute myeloid leukemia patient samples for proteomic and phosphoproteomic analysis.....	59
Proteomics analysis of the peritoneal dialysate effluent reveals the presence of calcium-regulation proteins and acute inflammatory response	60
Evaluation of alternative strategies depletion proteins for biomarker discovery in patients serum with bipolar disorder.....	61
Evaluation of proteome alterations induced by cadmium stress in sunflower(<i>Helianthus annuus</i> L.) cultures using 2-D DIGE and nESI-Q-Tof.....	62



JOURNAL OF INTEGRATED OMICS
A METHODOLOGICAL JOURNAL
[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

One gene, many changes: a long history involving soybean

Marco Aurélio Zezzi Arruda* ^{1,2}

¹Spectrometry, Sample Preparation and Mechanization Group – GEPAM, ²Institute of Science and Technology for Bioanalytics, Institute of Chemistry, Department of Analytical Chemistry, University of Campinas - Unicamp, PO Box 6154, 13803-970, Campinas, Brazil.

*Corresponding author: zezzi@iqm.unicamp.br

Available Online: 31 December 2015

ABSTRACT

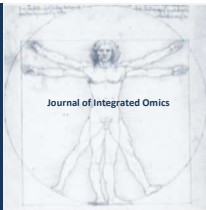
Exogenous genes are successfully introduced into plants since the 80's, and that one, which confer the resistance of the soybeans to glyphosate, the most used herbicide in the world, the first reported in the literature [1]. Since then, genetically modified organisms (GMOs) are being exponentially produced, so that 170 million hectares is nowadays related to crops engineered in at least 28 countries [2]. This fact not only reinforce the importance of such organisms, but also shows a good appeal to sustainability once that these GMOs produce enhanced characteristics such as high amount of fibers, proteins, are resistant to pests and/or insects, among others [2].

In this way, our research group is deeply interested in studying, since long time ago, the effects that a transgenic modification has on the proteome, the metabolome, the ionome, besides others of an organism [3-5], also evaluating different generations of plants. In this way, enzymatic evaluation, proteomics, and bioaccessibility will be presented for investigating some aspects regarding soybean [*Glycine max* (L.) Merrill] seeds and plants. Platforms like 2-D DIGE, ICP-MS, and ESI-MS/MS will be presented, as well as some results regarding stress production and the cultivation of this culture in the presence of nanoparticles.

Acknowledgements: FAPESP, CNPq, CAPES

References:

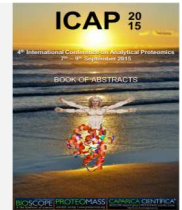
- [1] Monsanto do Brasil. http://www.monsanto.com.br/produtos/sementes/soja_roundup_ready/soja_round_ready.asp. [accessed at June, 24th, 2015]
- [2] Gilbert N. *Nature* 497(2013)21
- [3] Mataveli, LVR, Fioramonte, M, Gozzo, FC, Arruda, MAZ. *Metallomics* 4(2012)373
- [4] Barbosa HS, Arruda SCC, Azevedo RA, Arruda MAZ. *Anal Bioanal Chem* 402(2012)299.
- [5] Arruda SCC, Barbosa HS, Azevedo RA, Arruda MAZ. *J Proteom*, 93(2013)107



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Proteomic profiling of neuromuscular disorders

K. Ohlendieck*, S. Murphy, A. Holland

Muscle Biology Laboratory, Maynooth University, National University of Ireland, Maynooth, Co. Kildare, Ireland. *Corresponding author: kay.ohlendieck@nuim.ie

Available Online: 31 December 2015

ABSTRACT

Purpose: Neuromuscular disorders are associated with a great variety of pathophysiological triggering mechanisms, including primary genetic defects, acquired autoimmune processes, traumatic tissue injury, drug-induced abnormalities, cytotoxic insults and disuse-related atrophy, as well as changes during natural aging. In order to establish robust biomarker signatures for developing improved diagnostic, prognostic and therapy-monitoring test systems that can differentiate between individual muscular diseases, systematic evaluations of proteome-wide changes in pathological muscle specimens have been carried out.

Experimental description: Mass spectrometry-based proteomic profiling was used to compare skeletal muscle samples from normal versus pathological samples. Both, animal models of common neuromuscular disorders and human biopsy material was used in large-scale comparative studies. Subcellular fractionation procedures, fluorescence two-dimensional gel electrophoresis and/or liquid chromatography were employed for the efficient separation of the assessable muscle proteome. The independent verification of proteomic findings was carried out by enzyme assays, protein binding experiments, immunoblotting surveys and immunofluorescence microscopy.

Results: The systematic profiling of pathological skeletal muscle specimens has resulted in the establishment of a variety of novel biomarker candidates for the detailed characterization of muscular dystrophy, motor neuron disease, myotonia, diabetes-associated muscle weakness and sarcopenia of old age. Identified proteins are associated with cellular signalling, ion homeostasis, excitation-contraction coupling, the contractile apparatus, the cytoskeleton, the extracellular matrix and the cellular stress response.

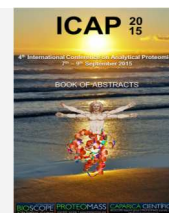
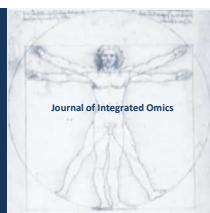
Conclusions: The newly identified proteomic biomarker candidates can now be further characterized and tested for their suitability as diagnostic, prognostic and therapy-monitoring indicator molecules in the field of neuromuscular pathology.

Keywords: biomarker, diabetes, motor neuron disease, muscular dystrophy, myotonia, sarcopenia of old age.

Acknowledgements: Research was funded by the Irish Higher Education Authority, the Deutsche Duchenne Stiftung aktion benni & co e.V., Muscular Dystrophy Ireland and the Hume scholarship programme of Maynooth University. The Programme for Research in Third Level Institutions PRTL Cycle 5 is co-funded by the Irish Government and the European Union under Ireland's EU Structural Funds Programme 2007-2013.

References:

- [1] Ohlendieck K. Proteomic identification of biomarkers of skeletal muscle disorders. *Biomarkers in Medicine* 2013;7:169-86.
- [2] Brinkmeier H, Ohlendieck K. Chaperoning heat shock proteins: proteomic analysis and relevance for normal and dystrophin-deficient muscle. *Proteomics Clinical Applications* 2014;8:875-95.
- [3] Holland A, Dowling P, Meleady P, Henry M, Zweyer M, Mundegar RR, Swandulla D, Ohlendieck K. Label-free mass spectrometric analysis of the mdx-4cv diaphragm identifies the matricellular protein periostin as a potential factor involved in dystrophinopathy-related fibrosis. *Proteomics* 2015 (in press).



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

The proteomic proficiency to identify and elucidate antimicrobial resistance bacteria

G. Igrejas^{*1,2,3,4}, A. Gonçalves^{1,2}, T. Santos^{1,2}, C. Marinho^{1,2}, S. Correia^{1,2}, R. Monteiro^{1,2}, L. Pinto^{1,2}, H. Radhouani^{1,2}, S. Ramos^{1,2}, J. L. Capelo^{3,4}, H. M. Santos^{3,4} and P. Poeta^{3,4,5}

¹Department of Genetics and Biotechnology, University of Trás-os-Montes and Alto Douro, Vila Real, Portugal. ²Functional Genomics and Proteomics Unit, University of Trás-os-Montes and Alto Douro, Vila Real, Portugal. ³BIOSCOPE Group, REQUIMTE-CQFB Chemistry Department, Faculty of Science and Technology, University NOVA of Lisbon, Lisbon, Portugal. ⁴ProteoMass Scientific Society. Faculty of Sciences and Technology. Campus de Caparica, Caparica. Portugal. ⁵Veterinary Science Department, University of Trás-os-Montes and Alto Douro, Vila Real, Portugal. *Corresponding author: gigrejas@utad.pt

Available Online: 31 December 2015

ABSTRACT

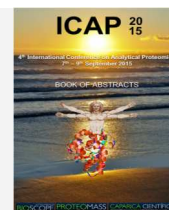
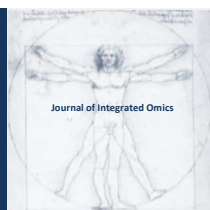
Purpose: To consider the contribution of proteomics to the study of antimicrobial-resistant (AMR) bacteria from clinical and environmental origin.

Experimental description: Proteomic approaches have been considerably improved during the past decade and have been used to overview and investigate the differences in protein expression profiles of cells grown under a broad spectrum of growth conditions and with different stress factors including antibiotics. The Functional Genomics and Proteomics Unit based at the University of Trás-os-Montes e Alto Douro (Vila Real, Portugal), using gel-based proteomic methodologies (e.g. 2D-GE, MALDI-TOF/MS, LC-MS/MS) has aimed to elucidate the distinctive characteristics of AMR bacteria from different sources, either clinical or environmental.

Results: From *Salmonella* Typhimurium and methicillin-resistant *Staphylococcus aureus* human clinical strains to *Enterococcus* spp. and *Escherichia coli* environmental strains recovered from wildlife species the work done so far has allowed to overview the proteome of several strains during induced stresses and to identify the most recurrent and deviations in the identified proteins associated biological processes. The research done allowed identifying antimicrobial proteins directly related with antimicrobial resistance or to bacterial virulence that complemented previously published genomic data where no specific mechanisms of resistance/virulence have been elucidated. Other identified proteins shown to be indirectly related with antimicrobial resistance or to bacterial virulence like chaperone proteins linked with aminoglycosides resistance, membranar proteins previously shown to increase the level of resistance to tetracycline or associated with increased virulence. Moreover, comparative proteomics studies presented an overview of the complex changes in expression and metabolism that occur when AMR bacteria is stressed with specific antibiotics. For instance, the abundance of chaperone, porin and export proteins showed to be particularly affected during antibiotic stressing conditions, which indicates that the stress response and transport functions are essential and directly influence the antibiotic resistance response of AMR strains.

Conclusions: The proteomic profiling allows to obtain an overview of the proteins present under specific stress conditions while the comparative proteomics methodologies allows to investigate differences in protein expression profiles of cells grown under different stress factors. Proteomic studies of stress-induced metabolism changes in bacterial strains from clinical and environmental origins might show how different strains are related and how different environments act as reservoirs.

Keywords: Proteomics, Gel-based Methodologies, Antimicrobial Resistance, Clinical Bacteria, Environmental Bacteria.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Protein of interests for serological diagnosis in Bird fancier's Lung

Adeline Rouzet^{*1}, Gabriel Reboux^{1,2}, Coralie Barrera^{1,2}, Bénédicte Rognon^{1,2}, Jean-Charles Dalphin^{1,3}, Paul De Vyust⁴, Laurence Millon^{1,2}, Sandrine Roussel^{1,2}.

¹Research team UMR/CNRS 6249 Chrono-environnement, University of Franche-Comté, 25030 Besançon (France). ²University Hospital of Besançon Parasitology – Mycology department, 25030 Besançon (France). ³University Hospital of Besançon, Pneumology department, 25030 Besançon (France). ⁴ULB Erasme Hospital of Brussels, Pneumology department, 1070 Brussels (Belgium). *Corresponding author: adeline.rouzet@gmail.com

Available Online: 31 December 2015

ABSTRACT

Purpose: Hypersensitivity pneumonitis (HP) is an inflammatory interstitial lung disease resulting from exposure to a wide variety of antigens contained in organic particles dust. Bird fancier's lung (BFL) is a form of HP caused by repeated inhalation of avian antigens, mainly located in droppings, blooms and feathers. Several candidate substances have been reported such as IgA, IgG and pigeon intestinal mucin, but none of them have been clearly identified. Diagnosis of HP requires an association of clinical, radiological and biological markers including serological test. However, immunological tests routinely used lack of specificity and standardization. The aim of this study was to investigate immunogenic proteins in pigeon dropping by comparative western blot analysis and to identify them by mass spectrometry.

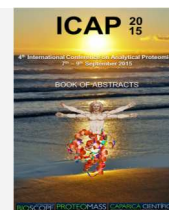
Experimental description: To evaluate antibody-mediated immunity, two-dimensional western blotting using *Columba livia* dropping were performed. The membranes were probed with sera from 10 patients with BFL, 2 healthy exposed controls and 2 controls without exposure to birds. 14 candidate proteins were excised from the two-dimensional-gel Electrophoresis and identified by liquid chromatography coupled to tandem mass spectrometry (LC-MS/MS).

Results: First of all, none of sera from control and exposed subjects presented specific antibodies against proteins from pigeon dropping extract. We identified 6 proteins which may be grouped into three categories based on their putative functions. The heavy chain V-III, the lambda chain and the polymeric receptor of immunoglobulin was respectively bound by antibodies from at least 90%, 80% and 80% of patient with BFL. Among the proteins of the immune system of birds, we also identified the mucin recognized by antibodies from at least 70% of patients. Pancreatic lipase and chymotrypsinogen, both digestive proteins secreted into the pancreatic fluid were also identified. These proteins were revealed by Western blotting for 80% of patients.

Conclusions: This study allowed the identification of 6 proteins of interest for serological diagnosis of BFL. These proteins related to immune and digestive systems of birds. Genes coding for these proteins were cloned and recombinant antigen are currently under development. The performance of these recombinant antigens will be evaluated by ELISA and may allow optimization of the serological diagnosis of BFL.

Keywords: Bird fancier's lung, mass spectrometry, immunoglobulin, western blotting, bird droppings.

Acknowledgements: This work was supported by the Besançon University Hospital through a project entitled APICHU D-PEO (Appel à Projet Interne de recherche Clinique ou biologique du Centre Hospitalier Universitaire). The authors thank breeders in the Franche-Comté region for their collaboration in this study.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

A case report of cyclosporine complexed with human calcineurin remaining in the gut after 25 years of its consumption

González-Correa Carlos-Augusto^{*1}, Gómez-Buitrago Paola-Andrea¹, Taborda-Ocampo Gonzalo¹, Acuña-Sornoza Ricardo², Santacoloma-Osorio Mario³, Zezzi-Arruda Marco-Aurelio⁴

¹Research Group on Electrical Bio-Impedance, Doctoral Program on Biomedical Sciences, University of Caldas, Manizales, Colombia.

²Colombian National Coffee Research Center CENICAFE, Chinchiná, Colombia. ³Clinical Department, University of Caldas, Manizales, Colombia. ⁴State University of Campinas, Institute of Chemistry, Campinas, Brazil. *Corresponding author: c.gonzalez@ucaldas.edu.co

Available Online: 31 December 2015

ABSTRACT

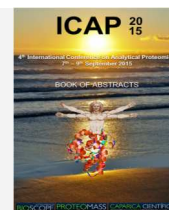
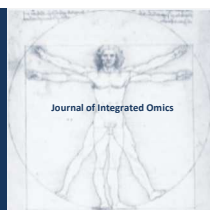
Purpose: To report on how residuals of cyclosporine, an extremely toxic immunosuppressant drug, can remain in the human gut after decades of its consumption and how a cleansing protocol of the colon can remove it.

Experimental description: in our Group we are in the way of implementing a leaning protocol aimed at promoting a beneficial modulation of the intestinal microbiota, increased physical activity and improved alimentary habits. The first part of this protocol is a 6-day intestinal cleansing program which includes: supplemented fasting, 1 daily transanal irrigation and consumption of psyllium (PSY), montmorillonite (MMT), prebiotics and probiotics. This program allows the collection of human intestinal mucus (HIM) samples in good quantity and quality as necessary for proteomic studies, a field we have been working on. Proteins from HIM were treated with a denaturalized buffer, and their separation was achieved by 2D-PAGE. Digested peptides were analyzed with a NanoAcquity UPLC chromatograph coupled to a HDMS spectrometer equipped with a NanoESI source.

Results: As a casual finding, the protein Chain-crystal-structure-of-human-calcineurin complexed-with-cyclosporin-A-and-human-cyclophilin was identified in the sample from a 40 years old woman. When asked about consumption of cyclosporine in the previous days to the cleansing program, this volunteer confirmed that she has taken it under medical prescription, but when she was only 15 y old, after she was diagnosed with Systemic lupus erythematosus (SLE). She remembered well the name of the drug, because she lost her hair and her nails at that time, as adverse effects to the drug. Calcineurin (CN) is a protein phosphatase that activates the nuclear factor of activated T cell cytoplasmic (NFATc), upregulating the expression of interleukin 2 (IL-2), which, in turn, stimulates the growth and differentiation of T cell response. Cyclosporine is an inhibitor of this protein and is commonly used in some autoimmune diseases.

Conclusions: 1. This finding confirm one of the most important functions of HIM as protection against pathobiota, metabolic waste and xenobiotics; 2. The proteomic study of HIM from an individual can be useful to know part of his/her clinical history as well as to help in the implementation of customized medicine; 3. Periodic cleansing of the gut can be advisable for persons that have orally consumed toxics, but probably also for people looking to improve their general well being. MMT, for instance is used in people diagnosed with or being suspected of intoxication with aflatoxins, as it can adsorb these toxins and help to remove them from the intestinal tract.

Keywords: cyclosporine, human intestinal mucus, transanal irrigation.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Improved efficiency of *in situ* tryptic proteolysis for fresh frozen and FFPE tissue

Ekta Patel^{*1}, Malcolm Clench¹, Peter Marshall², Andy West² and Simona Francese¹

¹BMRC, City Campus, Sheffield Hallam University, Howard Street, Sheffield, S1 1WB. ²GlaxoSmithKline, Gunnels Wood Road, Stevenage, Hertfordshire, SG1 2NY. *Corresponding author: Ekta.Patel@student.shu.ac.uk

Available Online: 31 December 2015

ABSTRACT

Purpose: Despite improvements to on-tissue digestion efficiency, methods to further improve both reliability and number of proteins identified when using a bottom up proteomic approach remains. Current investigations focus on improving the yield of tryptic peptides from fresh frozen rat brain tissue and FFPE mouse brain tissue.

Experimental description: Fresh frozen tissue: Following wash steps of ethanol and chloroform, 20µg/mL trypsin solution containing either OcGlu or MEGA-81 was deposited using the SunCollect sprayer (SunChrom, Friedrichsdorf, Germany). Samples were incubated for 4 hours at 37°C.

FFPE tissue: Following tissue deparaffinization using xylene and wash steps of ethanol in increasing concentration, an antigen retrieval step was carried out. 20µg/mL trypsin solution containing either OcGlu or MEGA-8 was deposited using the SunCollect sprayer. Samples were incubated for overnight at 37°C.

MALDI analysis: α-cyano-4-hydroxycinnamic acid (CHCA) with aniline in acetonitrile:water:TFAaq was spotted (10mg/mL) onto the digest regions or sprayed using the SunCollect (5mg/mL) and subjected to MALDI-MSP or MSI (HDMS SYNAPT™ G2 system, Waters Corporation, Manchester) analysis.

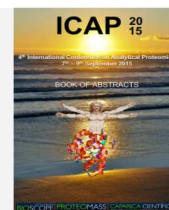
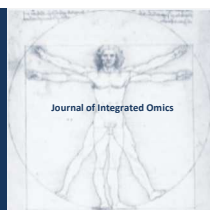
Results: The study concluded that the incorporation of a detergent yielded several hundred peptide peaks with S/N ratio above 3.5 for the digestion of fresh frozen tissue. Most of the peaks were considered to be from the most abundant proteins in brain; the tentatively identified myelin basic protein (MBP). In fresh frozen tissue, peak intensities originating from the detergent MEGA-8 are clearly superior to those of OcGlu. Existing methodologies have been transferred onto the digestion of FFPE tissue to generate a more efficient proteolysis.

Conclusions: In conclusion this study shows that ion population, abundance and protein identification can be further improved, thus opening up the way to more reliable and efficient biomarker discovery and screening.

Keywords: MALDI, *in situ*, Proteolysis, Detergents, Fingermarks.

References:

[1] DJIDJA, M.C., et al. (2009). *Proteomics*. 9: 1-15. 2. PATEL, E., et al. (2015) *JASMS*. 3. FERGUSON, L.S., et al. (2012). *Analyst*. 137: 4686-4692.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Development and evaluation of method for metalloproteins detection involved in bipolar disorder

G. S Pessôa^{*1,2}, J. R. de Jesus^{1,2}, A. Sussulini^{1,2}, J. L. C. Martínez³; M. A. Z. Arruda^{1,2}

¹Spectrometry, Sample Preparation, Mecanization Group, Analytical Chemistry Department, Institute of Chemistry, University of Campinas, P.O. box 6154, 13083970, Campinas, SP, Brazil. ²National Institute of Science and Technology for Bioanalytics, Analytical Chemistry Department, Institute of Chemistry, University of Campinas, P.O. box 6154, 13083970, Campinas, SP, Brazil. ³Bioscope group. Chemistry Department. Faculty of Sciences and Technology. New University of Lisbon. Campus de Caparica., 2829-516. Caparica. Portugal.

*Corresponding author: gpessoa@iqm.unicamp.br

Available Online: 31 December 2015

ABSTRACT

Purpose: Bipolar disorder (BD) is characterized as a chronic psychiatric illness, severe and highly debilitating, with recurrent mood disorders, presenting episodes of mania and depression separated by periods of normal behavior (Sussulini, *et al.*, 2009). The cause of bipolar disorder is unknown, although it is known that a variety of biochemical, genetic, and environmental factors can be involved in the process of developing of the disorder. To date, there is no biochemical test to confirm the disease, and the diagnosis is still based on experience and clinical judgment (Jesus, *et al.*, 2015). The main focus of this work was to propose a metalloproteomic strategy for BD, since this area has been not carried out in the literature.

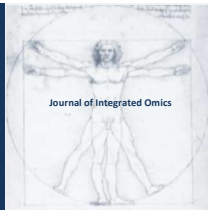
Experimental description: Metal and metalloid ion involved in this pathology were determined by ICP-MS, using a dynamic reaction cell (Perkin Elmer, ELAN DRC-e). For metalloproteomic evaluation, a mild procedure for removal of major abundant protein was applied. For this task, a high performance liquid chromatograph was used to separate the proteins, using a sequential detection of proteins (UV-Vis) and metal ions (hyphenated LC-ICP-MS).

Results: Considering the elemental determination, the DRC strategy was considered as an efficient method in the removal of interferences, since satisfactory results in terms of trueness and precision were obtained, using a certified reference material. Thus, an ionic profile of each group was possible. After the chromatographic separation of proteins and metalloproteins, the removal of major abundant proteins is a main challenging step, since for a metallomic study, the information about tertiary structure must be preserved as intact as possible. The detection of proteins in UV-Vis and metalloproteins in LC-ICP-MS confirmed that our strategy was reliable for a metalloproteomic evaluation.

Conclusions: Differential levels of metal and metalloid ions were observed when comparing the three studied groups, as well as the detection of metalloproteins by LC-ICP-MS. Thus, our method can be considered as reliable for evaluation of BD disorder.

Keywords: bipolar disorder, metallomic, metalloprotein.

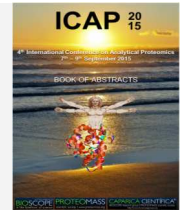
Acknowledgements: FAPESP, FAEPEX, Capes and CNPq



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Using protein biomarkers to understand the biological mechanisms underlying beef sensory qualities

Mohammed Gagaoua^{*1,2,3}, E.M. Claudia Terlouw^{1,2}, Brigitte Picard^{1,2}

¹INRA, UMR 1213 Herbivores, F-63122 Saint-Genès- Champanelle, France. ²Clermont Université, VetAgro Sup, UMR 1213 Herbivores, B.P. 10448, F-63000 Clermont-Ferrand, France. ³Equipe Maquav, INATAA, Université des Frères Mentouri Constantine 1, Route de Ain El-Bey, 25000 Constantine, Algeria. *Corresponding author: brigitte.picard@clermont.inra.fr; mohammed.gagaoua@clermont.inra.fr

Available Online: 31 December 2015

ABSTRACT

Purpose: Comparative proteomic analyses have revealed that numerous proteins are related to meat sensory qualities. They belong to myriad biological pathways: metabolism, structure, heat stress, oxidative stress, apoptosis, proteolysis and others¹. These relationships suggest that these proteins may play key roles in the mechanisms underlying meat qualities. Therefore, they can be used, not only as biomarkers for meat and carcasses quality, but also to unravel these mechanisms. In this study, the central role of Grp75 (*HSPA9*), a member of the Hsp70 family, in the biological mechanisms involved in the conversion of muscle to meat is described.

Experimental description: *Longissimus thoracis* (LT, fast oxydo-glycolytic) muscles of 21 French Blonde d'Aquitaine young bulls were used in this study². pH parameters at 45 min, 3h and 30h *p.m.* and CIE-*L*a*b** colour traits after 24h *p.m.* were determined. Protein biomarkers according to Picard *et al*¹ were quantified using Dot-Blot on muscles sampled 30 min after slaughter. Relationships between colour and pH traits and the biomarkers were determined by means of regression and Pearson correlation analyses using SAS 9.2 software.

Results: Grp75 protein was related to pH and colour parameters. It was also related to numerous biological pathways, likely glycolytic metabolism (Enolase, $r = 0.63$, $P < 0.01$ and LDHB, $r = 0.42$, $P < 0.05$), proteolysis (μ -calpain, a Ca^{2+} dependent protease, $r = 0.54$, $P < 0.05$), structure maintenance (MyLC1-F, $r = 0.52$, $P < 0.05$ and MyBP-H, $r = 0.63$, $P < 0.01$) and oxidative stress (Prdx6, SOD1 and DJ-1, $r = 0.61$ - 0.63 , $P < 0.01$). Grp75 was earlier described as a multi-functional protein. It is a cytoprotective chaperone present in nearly all cellular compartments and assists in the import and folding of mitochondrial proteins and protects cells from glucose deprivation. It is a member of the Hsp70 protein family, and not induced by heat stress but by glucose deprivation, Ca^{2+} influx or agents perturbing glycolysis. At slaughter, after bleeding of the animal, lack of oxygen and nutriment will lead to accumulation of ROS in the mitochondria, the end of ATP production, acidification of the cytoplasm and a deregulation of Ca^{2+} in the muscle cells which may set off apoptosis³. In the present study, Grp75 content soon after slaughter was related to the glycolysis pathway and pH, suggesting that it plays a central role in the conversion of muscle to meat, and consequently, meat sensory qualities. In addition, Grp75 was also reported to link the inositol 1,4,5-trisphosphate receptor (IP3R) to voltage-dependent anion channel (VDAC), presumably enhancing Ca^{2+} trafficking from endoplasmic reticulum (ER) towards mitochondria and cytosol. Possibly, increased Ca^{2+} levels in the cytoplasm activate μ -calpain and/or regulate apoptosis. Our results indicate that Grp75 may play a central role in changes in the structural proteins in the muscle cell, and consequently in meat colour development.

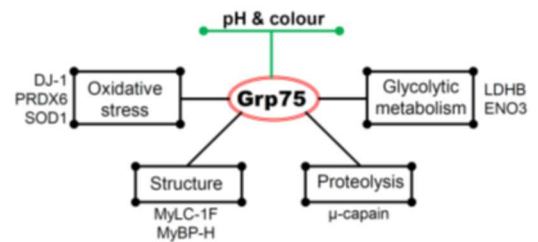
Conclusions: This example illustrates the important role of analytical proteomics in improving knowledge on the determinism of meat sensory qualities.

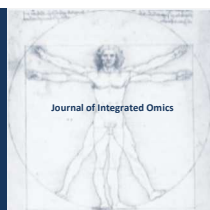
Keywords: Beef meat; Colour; pH; Biomarkers; Validation; Prediction; Biological mechanisms.

Acknowledgements: This work was part of the EU ProSafeBeef project (FOODCT-2006-36241). A grant of the PROFAS B+ program given to M.G. is also gratefully acknowledged.

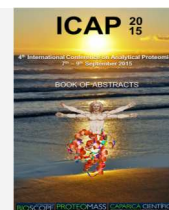
References:

- [1] Picard, B.; Gagaoua, M.; Micol, D.; Cassar-Malek, I.; Hocquette, J. F.; Terlouw, E. M. C., Inverse relationships between biomarkers and beef tenderness according to contractile and metabolic properties of the muscle. *J Agric Food Chem* 2014, 62, 9808-18.
- [2] Gagaoua, M.; Terlouw, E. M. C.; Micol, D.; Boudjellal, A.; Hocquette, J. F.; Picard, B., Understanding Early Post-Mortem Biochemical Processes Underlying Meat Color and pH Decline in the *Longissimus thoracis* Muscle of Young Blond d'Aquitaine Bulls Using Protein Biomarkers. *J Agric Food Chem* 2015. Doi: 10.1021/acs.jafc.5b02615
- [3] Ouali, A.; Gagaoua, M.; Boudida, Y.; Becila, S.; Boudjellal, A.; Herrera-Mendez, C. H.; Sentandreu, M. A., Biomarkers of meat tenderness: present knowledge and perspectives in regards to our current understanding of the mechanisms involved. *Meat Sci* 2013, 95, 854-70.





JOURNAL OF INTEGRATED OMICS
A METHODOLOGICAL JOURNAL
[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Some proteins of possible -clinical interest found in samples of human intestinal mucus (HIM)

Gómez-Buitrago Paola-Andrea^{*1}, González-Correa Carlos-Augusto¹, Taborda-Ocampo Gonzalo¹, Acuña-Sornoza Ricardo², Santacoloma-Osorio Mario³, Zezzi-Arruda Marco-Aurelio⁴.

¹Research Group on Electrical Bio-Impedance, Doctoral Program on Biomedical Sciences, University of Caldas, Manizales, Colombia.

²Colombian National Coffee Research Center CENICAFE, Chinchiná, Colombia. ³linical Department, University of Caldas, Manizales, Colombia. ⁴State University of Campinas, Institute of Chemistry, Campinas, Brazil. **Corresponding author: quimicapao@gmail.com*

Available Online: 31 December 2015

ABSTRACT

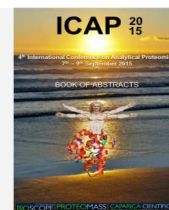
Purpose: To report the identification of some proteins isolated from samples of human intestinal mucus (HIM) with possible clinical interest.

Experimental description: HIM was collected from six volunteers admitted to a leaning program which included a colon cleansing protocol and a rectosigmoidoscopy. Protein extraction was achieved by a denaturalization buffer, separation was performed by gel electrophoresis both with SDS-PAGE and 2D-PAGE. Protein shotgun was done with solution desalinization by SPE and albumin depletion. Digested peptides were analysed using a NanoAcquity UPLC chromatograph coupled to a HDSM espectrometer, equipped with a NanoESI source. Samples were injected to a C₁₈ precolumn with a flow of 5 µL/min water/acetonitrile (97:3) with 0,1% formica cid and then transfered to an analytical column C₁₈ and eluted with a flow of 1 µL/min.

Partial results: 1. A methodology for the analysis of HIM from a single person is reported, from the preparation of the patients for the collection of the sample, till protein characterization of the mucus. This is done using mass spectrometry, using three different techniques: SDS -PAGE, 2D-PAGE y shotgun; 2. 14 proteins of clinical interest have been identified so far: Annexin A2, Calcium-activated chloride channel regulator, Chain crystal structure of human calcineurin complexed with cyclosporin A and human cyclophilin, Complement C3, Disulfide isomerase, Galectin-3 y 4, Inmunoglobulin programmed cell death 1 ligand, LAMTOR3, Muc2, Nuclear Protein 8, RuvB like 2, Tyrosine Protein phosphatase non receptor type 11 and Zymogen granule membrane protein 16.

Conclusion: The proteomic study of HIM can be a very useful tool for the study of different diseases, not only of those directly related to the digestive system, bus also many associated with the immune system.

Keywords: electrophoresis, human intestinal mucus, shotgun proteomics.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Proteomic alterations in transgenic and non-transgenic soybean leaves after plant cultivation in the presence of silver nanoparticles

R. M. Galazzi^{*1,2}, C. A. Lopes Júnior^{1,2,3}, M. A. Zezzi-Arruda^{1,2}

¹Spectrometry, Sample Preparation and Mechanization Group – GEPAM, Institute of Chemistry, University of Campinas – UNICAMP, P.O. Box 6154, 13083-970, Campinas-SP, Brazil. ²National Institute of Science and Technology for Bioanalytics, Institute of Chemistry, University of Campinas–UNICAMP, P.O. Box 6154, 13083-970, Campina-SP, Brazil. ³Department of Chemistry, Federal University of Piauí – UFPI, P.O. Box 64049-550, Teresina-PI, Brazil. *Corresponding author: rodrigo.galazzi@iqm.unicamp.br.

Available Online: 31 December 2015

ABSTRACT

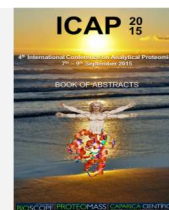
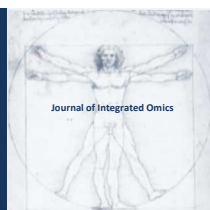
Purpose: The present study evaluates the proteomic map of transgenic (T) and non-transgenic (NT) soybean leaves after plant cultivation with silver nanoparticles (AgNPs) or silver nitrate (AgNO₃), at 50 mg kg⁻¹ silver concentration.

Experimental description: The T and NT soybean plants were cultivated for 21 days in a growth chamber with controlled photoperiod (12 h) and temperature (27 ± 1 °C). For treated plants, AgNO₃ and AgNP solutions were added, resulting, in the end of the cultivation period, a 50 mg kg⁻¹ of silver. The proteins were extracted from leaves with phenol, separated by two-dimensional difference gel electrophoresis (2-D DIGE) to evaluate possible changes in the proteomic map. The differential protein abundances were evaluated by a DeCyder® program, considering a threshold of 2.0 fold (100% variation).

Results: The protein content for control T soybeans leaves was higher than NT soybean leaves (*ca.* 18%). For NT plants exposed to AgNO₃ (NT-AgNO₃) and AgNP (NT-AgNP), the NT-AgNO₃ showed an increase of *ca.* 35% in terms of protein concentration in their leaves. No significant alterations in protein content for NT-AgNP in comparison with the control were observed. However, for T plants treated with AgNO₃ (T-AgNO₃) and AgNP (T-AgNP), the protein concentration in T soybean leaves was changed in both cases. A decrease of *ca.* 6% in T-AgNO₃ compared with control plants was observed, whereas for T-AgNP an increase in the protein content (*ca.* 28%) was observed. Regarding the evaluation of differential abundant proteins, 19 and 11 protein spots are found for NT vs NT-AgNO₃ and NT vs NT-AgNP, respectively, and 22 protein spots are differential for NT-AgNO₃ vs NT-AgNP group. Considering T vs T-AgNO₃ and T vs T-AgNP groups, 8 and 12 protein spots are differential abundant, respectively, while for T-AgNO₃ vs T-AgNP group, 9 protein spots are found. These results corroborates with those of protein concentration, suggesting that changes in the proteomic map were dependent on the genetic modification, as well as the form in which silver is added in the cultivation (AgNP or AgNO₃).

Conclusions: According to the results, the treatment with AgNO₃ and AgNP changes the proteomic map for both T and NT soybean leaves, once that protein spots with different abundances were found. Further experiments will be carried out for identification of these differential proteins by LC-MS/MS.

Keywords: transgenic soybeans; silver nanoparticles; proteomics; 2-D DIGE.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Rice Proteomics: Method Development of 2D-PAGE for Malaysian Upland Rice

Arif, S. M.^{*1}, Rahmat, Z.¹, Ma, N. L.²

¹Department of Biotechnology and Medical Engineering, Faculty of Biosciences and Medical Engineering, Universiti Teknologi Malaysia, 81310, Johor Bahru, Johor, Malaysia. ²Department of Biological Science, Faculty of Science and Technology, Universiti Malaysia Terengganu, 21030 Kuala Terengganu, Terengganu, Malaysia. *Corresponding author: syakila.arif@gmail.com ; zaidah@utm.my

Available Online: 31 December 2015

ABSTRACT

Purpose: Method development is the establishment of analytical method that will be applicable and copacetic for the target analysis. It is one of the critical determinants to ensure the successful of the experiment being carried out. The aim of this study is to develop a suitable 2D-PAGE method that could help in the proteomics analysis of Malaysian upland rice. Although 2D-PAGE for rice proteomics has been established for more than a decade ago, none has been reported for Malaysian upland rice and it is acknowledged no universal protocol ever exists as each plant behaves differently.

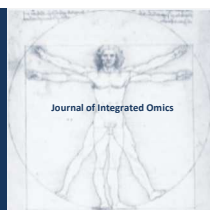
Experimental description: In this study, a cultivar of Malaysian upland rice named Wai with age ranging from 4-16 weeks were extracted by using modified TCA/Acetone based method. The isoelectric focusing (IEF) was performed using 18cm immobiline pH gradient strip, pH range 4-7 with the protocol: 1 h at 100 V (linear), 2h at 500 V (gradient), 12h at 8000 V (gradient), 2h at 8000 V (linear) and 10h at 100 V (slow). The strip was loaded on 12% acrylamide gel for the second phase and electrophoresis was carried out at 100 V for 16 hours and stained with comassie brilliant blue red 250. The protein spot was analyzed using PDQuest software version 8.0.

Results: The suitability of the method developed is determined based on the resolution of the protein spot on the gel and its pattern. The result shows that it is feasible to visualize numbers of protein spot with good intensity for all age range (4-16 weeks) of Wai cultivar which indicates the suitable method for 2D-PAGE analysis of Malaysian upland rice is developed.

Conclusions: This study accommodates new insights for upland rice protein separation, serving as platform that boosts the relevance and importance of the developed 2D-PAGE for further Malaysian upland rice proteomics analysis.

Keywords: Development, Proteomics, Upland Rice, 2D-PAGE.

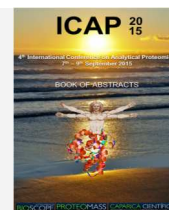
Acknowledgements: We thank the Universiti Teknologi Malaysia (UTM) and Ministry of Education (MOE) for funding this project (Fundamental Research Grant Scheme, Vote: 4F366 and Research University Grant, Vote: 10J16) and Zamalah for sponsoring the graduate student scholarship.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Effect of resistance inducers and *Hemileia vastatrix* infection on protein patterns of *Coffea arabica* leaves Title

K. Possa^{1,2}, R. Tenente², I. Chaves^{3,4}, C. Pinheiro^{3,5}, J. Renaut⁶, C. P. Ricardo³, M. L. V. Resende¹, L. Guerra-Guimarães^{*2,7}

¹Dep. Fitopatologia, Universidade Federal de Lavras, Lavras, Minas Gerais, 37200-000, Brazil. ²Centro de Investigação das Ferrugens do Cafeeiro/Instituto de Investigação Científica Tropical, Quinta do Marquês, 2784-505 Oeiras, Portugal ³Instituto de Tecnologia Química e Biológica/Universidade Nova de Lisboa, Quinta do Marquês, 2780-157 Oeiras, Portugal. ⁴Instituto de Biologia Experimental e Tecnológica, Quinta do Marquês, 2780-157 Oeiras, Portugal. ⁵Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa, 2829-516 Caparica, Portugal. ⁶Luxembourg Institute of Science and Technology, Environmental Research and Innovation (ERIN) department, 41, rue du Brill, L-4422 Belvaux, Luxembourg. ⁷Linking Landscape, Environment, Agriculture and Food /Universidade de Lisboa, Tapada da ajuda, 1349-017 Lisboa, Portugal. *Corresponding author: leonorguima@gmail.com

Available Online: 31 December 2015

ABSTRACT

Purpose: The coffee leaf rust (*Hemileia vastatrix*) disease causes serious losses in coffee production and quality, and if no chemical control measures are taken huge depreciation of marketing values will occur. For a sustainable coffee production there are increasing societal expectations to reduce pesticide treatments and make use of alternative strategies of plant protection, such as the application of resistance inducers. Using a comparative proteomic analysis of *Coffea arabica* leaves we aimed to understand the cellular mechanisms that are behind the disease control capacity of the resistance inducers Greenforce CuCa or Bion[®], when applied prior to infection with *H. vastatrix*. These resistance inducers have shown efficiency in the control of the disease when compared to a standard fungicide¹

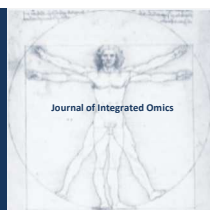
Experimental description: Leaves of *Coffea arabica* cv. Mundo Novo were sprayed with GreenForce CuCa (formulation based on coffee berry husk supplemented with cooper and calcium, developed in UFPA, Brazil) and Bion[®] (acibenzolar-S-methyl-ASM). Three days after treatment, leaves were sprayed with *H. vastatrix* urediniospores. Leaves treated with water or inoculated (without any previous treatment) were used as controls. Samples were collected at 3, 5 and 7 days after treatments, corresponding respectively to 0, 2 and 4 days after inoculation. Protein extracts were obtained by precipitation with trichloroacetic acid in acetone following purification with the 2D clean-up kit (GE Healthcare). After separation of proteins by 2-DE and staining with Colloidal Coomassie blue, the most abundant and visible polypeptides spots were removed from the gel and their identification was accomplished by mass spectrometry (MALDI/TOF/TOF MS) followed by homology search in ESTs coffee databases.

Results: We were able to identify a total of 204 leaf proteins which were mainly involved in photosynthesis, protein metabolism, biotic and abiotic stresses, redox reactions, glycolysis and secondary metabolism; of those proteins 65 changed in abundance between treated and/or infected leave samples. GreenForce CuCa affected mainly photosynthetic proteins, while Bion[®] treatment, affected mainly energy metabolism and stress. The protein profile of leaves treated with Bion[®] resembles more the profile of the healthy control than of the GreenForce CuCa treatment.

Conclusions: The results showed that the resistance inducers affected mainly proteins of the primary metabolism; this metabolic adjustment has contributed to an increased in the resistance of the coffee plant to the pathogen. ¹Monteiro *et al.* 2013. In: VIII Simpósio de Pesquisas dos Cafés do Brasil, Salvador, Bahia, Brazil.

Keywords: Coffee Leaf Rust, Leaf proteome, 2-DE, MALDI-TOF/TOF-MS, GreenForce CuCa, Bion[®]

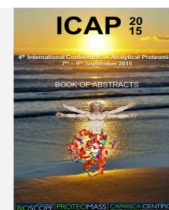
Acknowledgements: This work was supported by Portuguese Funds through FCT (Fundação para a Ciência e a Tecnologia), under the project PTDC/AGR-GPL/109990/2009 and the strategic project PEst-OE/eqb/LA0004/2011 and by a Pos-PhD grant and PhD grant of CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico - Brazil) and INCT-Café (Instituto Nacional de Ciência e Tecnologia do Café, Brazil).



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

A quest for innovative and robust biomarkers using MS-based proteomics

L. Dayon*, A. Núñez Galindo, John Corthésy, S. Oller Moreno, O. Cominetti, M. Kussmann

Molecular Biomarkers Core, Nestlé Institute of Health Sciences, Lausanne, Switzerland. *Corresponding author: loic.dayon@rd.nestle.com

Available Online: 31 December 2015

ABSTRACT

Purpose: The identification of novel biomarkers with proven clinical utility relies on a complex process composed of discovery, qualification, verification, and validation steps. Proteomic biomarker discovery has provided numerous candidates but only few of those have reached the validation stage and even less are used in today's routine clinical practice. Mass spectrometry (MS)-based proteomics is powerful enough to comprehensively characterize biological and clinical samples but offers limited sample throughput, mainly due to complex sample preparation upstream of MS. Proteomic biomarker discovery studies are therefore often performed with a too small number of samples, compromising study design. We have developed dedicated highly-automated workflows to facilitate, accelerate, and improve molecular phenotyping and translation of clinical proteomic discovery findings in human body fluids such as plasma and cerebrospinal fluid.

Experimental description: A highly automated MS-based proteomic discovery approach that comprises immuno-affinity depletion of abundant plasma proteins, buffer exchange, reduction/alkylation, tryptic digestion, isobaric labeling, sample purification and reversed-phase liquid chromatography tandem MS analysis was developed, characterized [1] and finally used to analyze cohorts of patients, ranging from 100 to 1'000 individuals [2].

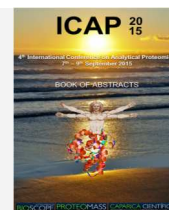
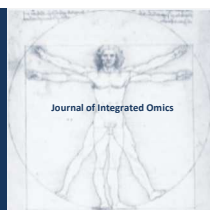
Results: Analytical figures of merit, *e.g.*, precision and trueness, of our approach demonstrated increased robustness compared to to-date published manual or less automated workflows while ensuring sufficient throughput for large-scale studies. Application to clinical biomarker research programs in the fields of metabolic and brain health provided protein biomarker models for stratification of subjects and prediction of relevant clinical readouts.

Conclusions: Our results revealed that the analysis of large number of samples for biomarker discovery in clinical research using today's MS-based shotgun proteomics technology has become feasible. The analytical and biological consistency of our results regarding technical validation but also clinical research demonstrated the value of our approach.

Keywords: Biomarker; Body fluid; Blood; Clinical proteomics; Mass spectrometry; Obesity; Alzheimer's disease.

References:

- [1] Dayon, L.; Núñez Galindo, A.; Corthésy, J.; Cominetti, O.; Kussmann, M., Comprehensive and scalable highly automated MS-based proteomic workflow for clinical biomarker discovery in human plasma. *J. Proteome Res.* 2014, 13, (8), 3837-3845.
- [2] Cominetti, O.; Núñez Galindo, A.; Corthésy, J.; Oller Moreno, S.; Irincheeva, I.; Valsesia, A.; Astrup, A.; Saris, W.H.M.; Hager, J.; Kussmann, M.; Dayon L., Proteomic Biomarker Discovery in 1'000 Human Plasma Samples with Mass Spectrometry: Vision or Reality?. Submitted.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Matrix metalloproteinases (MMPs) in buck blood chronically infected by arthritis encephalitis virus in Northeast of Brazil

R.Q. Bezerra Júnior¹, A.M.X. Eloy^{*2}, J.R. Furtado², Pinheiro, R.R.², Pinheiro, A.A.²

¹Laboratory of Virology, Universidade Estadual do Ceará, Campus Itaperi, Fortaleza, CE, Brazil. ²Empresa de Pesquisa Agropecuária – EMBRAPA Goats and Sheep, Estrada Sobral/Groaíras, Sobral, CE, Brazil. *Corresponding author: angela.eloy@embrapa.br

Available Online: 31 December 2015

ABSTRACT

The objective of this work was to investigate the activity of the MMPs (MMP-2 e MMP-9) in blood serum of chronically infected bucks by CAEV. The experiment was constituted by two groups (n = 5 each group). The first one was composed by five naturally infected bucks (4-5 years) and the second group constituted of five negative bucks (4-5 years) for CAE. Serology was performed using the Western Blotting (WB) and confirmed by Polymerase Chain Reaction (PCR). Blood samples were collected by puncturing the jugular vein from animals and evaluated by zymography (SDS-PAGE) using gelatin as substratum. The gel densitometry analysis showed higher activity ($P < 0.05$) of the pro-MMP-2 and MMP-2 in seropositive group (Fig. 1). The higher activity of the MMPs (MMP-2 and MMP-9) in infected group to CAE suggests a role of these proteases in the chronic inflammatory process in goats, once the key role of MMPs molecules in inflammation are involved in pathophysiological remodeling of the vascular wall, even in animals without apparent disease.

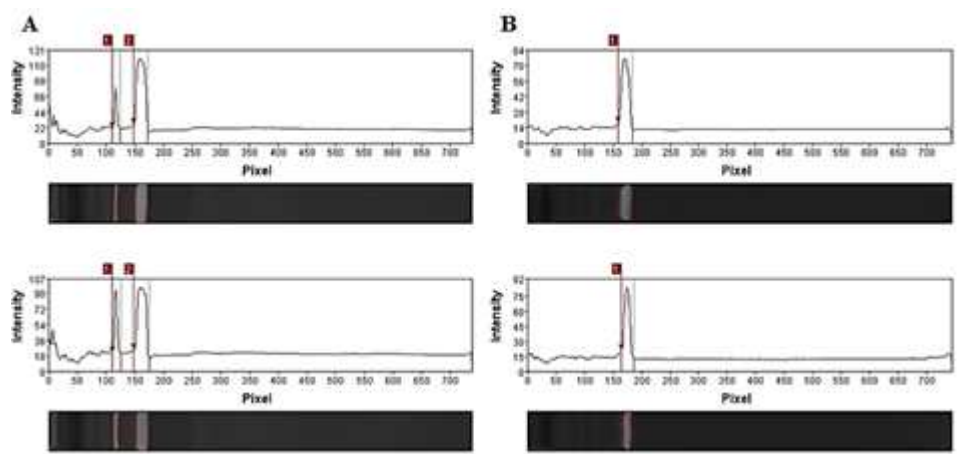
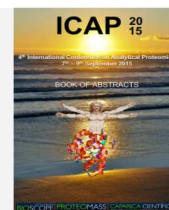
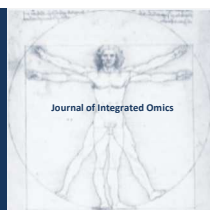


Fig.1 Densitometry of gels zymography showing the MMPs activity in seropositive (A) and seronegative (B) group for CAE.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Identification of immunoreactive microbial proteins in the field of allergic diseases, and application for serodiagnosis

Laurence Millon^{*1,2}, Coralie Barrera¹, Benedicte Rognon^{1,2}, Adeline Rouzet¹, Sandrine Roussel^{1,2}, Michel Monod³, Gabriel Reboux^{1,2}

¹CNRS, University of Franche-Comté, UMR 6249 Chrono-Environnement, France. ²University Hospital of Besançon, Department of Parasitology-Mycology, France. ³ Department of Dermatology, Laboratoire de Mycologie, Centre Hospitalier Universitaire Vaudois Lausanne, Switzerland. *Corresponding author: lmillon@chu-besancon.fr

Available Online: 31 December 2015

ABSTRACT

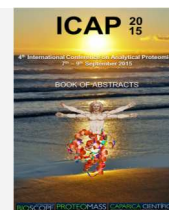
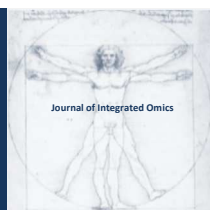
Purpose: Diagnosis of immunoallergenic pathologies due to environmental microorganisms includes detection of circulating specific antibodies. Immunoproteomics have proved to be useful for identifying the immunogenic proteins in several microorganisms linked to allergic diseases.

Experimental description: With this approach, the causative microorganisms are first isolated from the environment of patients. Then the proteins are separated by two-dimensional electrophoresis and revealed by Western blotting with sera of different patients suffering from the disease compared to sera of asymptomatic exposed controls. Immunoreactive proteins are identified by mass spectrometry. Identified immunoreactive proteins found to be specific markers for the disease could be subsequently produced as recombinant antigens using various expression systems to develop ELISA tests.

Results: Using recombinant antigens, standardized ELISA techniques can be developed, with sensitivity and specificity reaching 80% and 90%, respectively. Such techniques have been developed in our lab for diagnosis of Farmer Lung Disease (FLD), which is the most frequent form of occupational hypersensitivity pneumonia. Combinations of recombinant antigens from the 3 main micro-organisms involved in FLD (2 fungi and one actinomycete) are being tested at the moment in a multicenter study. We also have developed an ELISA test for diagnosis Machine Operator Lung, using 2 recombinant antigens from mycobacteria.

Conclusion: Immunoproteomics can be applied to any environmental microorganisms, with the aim of proposing panels of recombinant antigens able to improve the sensitivity and standardization of serologic diagnosis of hypersensitivity pneumonitis, but also other mold-induced allergic diseases such as allergic bronchopulmonary aspergillosis or asthma.

Keywords: allergy; immunoproteomics; serodiagnosis; recombinant antigens.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Caenorhabditis elegans based proteomic analysis during bacterial infections

Krishnaswamy Balamurugan*

Department of Biotechnology, Alagappa University, Karaikudi-630 004, India. *Corresponding author: bsuryar@yahoo.com

Available Online: 31 December 2015

ABSTRACT

Purpose: Infectious disease caused by specific bacteria with increased resistance to available antibiotics raise alarm for better understanding of microbial infections and host susceptibility. Studying host-pathogen interaction is a complex process, the outcome can be defined using information from both host and pathogen perspective in different dimensions. Evolution of molecular tools have enabled to explore the bacterial virulence factors and host immune effector molecules providing in depth knowledge about infection and immune response. Further, suitability of model organisms and advancement of proteomics techniques shed light on host innate immune response against bacterial infections. *Caenorhabditis elegans* is a well-known model for studying bacterial infections for decades and enormous data has been generated using forward and reverse genetics in this model. Using *C. elegans* as a model, the host proteome changes against set of Gram negative pathogens including *Vibrio alginolyticus*, *Proteus mirabilis* and *Pseudomonas aeruginosa* were studied.

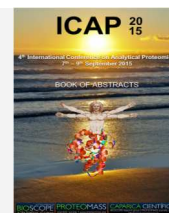
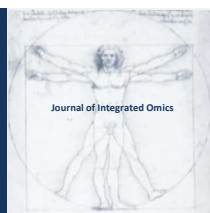
Experimental description: In our studies, regulation of *C. elegans* proteome was monitored against Gram negative bacterial infections using quantitative proteomic approach. Proteins were separated using two-dimensional differential gel electrophoresis (2D-DIGE) and differentially regulated proteins were identified using PMF and MALDI TOF-TOF analyses. The levels of expression of candidate proteins and their specific mRNA were validated using Western blot analysis and quantitative PCR, respectively. The interacting partners were shortlisted by bioinformatics tools and subsequently validated.

Results: Our results suggest that *C. elegans* displayed pathogen specific response by differentially regulating proteins against different pathogens. Identified proteins are found to be key regulators of essential pathways namely, unfolded protein response, MAP kinase and RAS signaling pathways.

Conclusions: For the first time our studies report the role of *C. elegans* proteins PDI-2, DAF-21 and EEF-2 in regulating host immune system against bacterial infections.

Keywords: *Caenorhabditis elegans*, *Vibrio*, *Pseudomonas*, *Proteus*, Proteomics.

Acknowledgements: Financial support from DBT, DST, CSIR and ICMR to Dr. K. Balamurugan is kindly acknowledged.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Quantitative analysis of the mitochondrial proteome and phosphoproteome in the yeast *Saccharomyces cerevisiae*

Claire Lemaire^{*1}, Margaux Renvoisé¹, Ludovic Bonhomme², Marlène Davanture³, Benoit Valot², Michel Zivy³

1.CNRS-UMR9198, Institute for Integrative Biology of the Cell (I2BC), Commissariat à l'Energie Atomique et aux Energies Alternatives (CEA), Centre National de la Recherche Scientifique (CNRS), Université Paris-Sud B3S, CEA-Saclay, Bâtiment 532, 91191 Gif-sur-Yvette, France. ²INRA, PAPPSO, UMR de Génétique Végétale, Gif-sur-Yvette, France. ³CNRS, PAPPSO, UMR de Génétique Végétale, Gif-sur-Yvette, France. *Corresponding author: claire.lemaire@cea.fr

Available Online: 31 December 2015

ABSTRACT

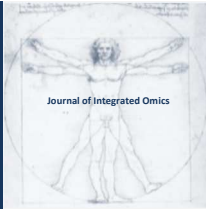
The mitochondrion is an organelle with multiple functions, the most important of these being to provide energy to the cell as a whole in the form of ATP, generated by oxidative phosphorylation catalyzed by the respiratory enzymes in the mitochondrial inner membrane. In humans, deregulation of mitochondrial functions, particularly with regard to the respiratory chain, is associated with several pathologies, including neurodegenerative diseases, neuromuscular diseases, type II diabetes and cancer. The activity of the respiratory enzymes may be modulated in response to metabolic demand and various types of stress. Several levels of regulation may be conceived, including changes in protein expression magnitudes, reversible interaction with effectors, and post-translational modifications, such as phosphorylation. The steadily increasing number of identified mitochondrial phosphoproteins suggests that reversible protein phosphorylation could be an important level of regulation in mitochondria. However, this hypothesis cannot be tested without quantitative data on variations in the abundance of mitochondrial proteins and their level of phosphorylation under different growth conditions. The yeast *Saccharomyces cerevisiae* is a powerful tool for studying various energetic and physiological states as it is a facultative aerobe which can grow on either fermentative or respiratory substrates.

We present for the first time a quantitative study of both protein abundance and phosphorylation levels in isolated yeast mitochondria under respiratory (lactate) and fermentative (glucose or galactose) conditions. To focus our analysis specifically on mitochondrial proteins, we performed a subcellular fractionation and used LC-MS/MS to overcome the limitations of 2D gel electrophoresis. Protein abundances were quantified using a label-free method. The phosphoproteome was analyzed quantitatively using the multiplex stable isotope dimethyl labeling procedure. For all quantified phosphopeptides, protein abundance was determined, allowing normalization of the data and permitting analysis of the specific variation of phosphorylation status independent of changes in protein abundance.

Label free quantitative analysis of protein accumulation revealed significant variation of 176 mitochondrial proteins including 108 proteins less accumulated in glucose medium than in lactate and galactose media. We also showed that the responses to galactose and glucose are not similar. Stable isotope dimethyl labeling allowed the quantitative comparison of phosphorylation levels between the different growth conditions. This study enlarges significantly the map of yeast mitochondrial phosphosites as 670 phosphorylation sites were identified, of which 214 were new and quantified. Above all, we showed that 90 phosphosites displayed a significant variation according to the medium and that variation of phosphorylation level is site-dependent.

This proteomic and phosphoproteomic study is the first extensive study providing quantitative data on phosphosites responses to different carbon substrates independent of the variations of protein quantities in the yeast *S. cerevisiae* mitochondria. The significant changes observed in the level of phosphorylation according to the carbon substrate open the way to the study of the regulation of mitochondrial proteins by phosphorylation in fermentative and respiratory media. In addition, the identification of a large number of new phosphorylation sites show that the characterization of mitochondrial phosphoproteome is not yet completed.

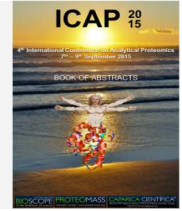
Keywords: Phosphoproteomic, Proteomic, Glucose, Galactose, Lactate, OXPHOS.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Dynamism of protein expression in gametes and early mammalian embryos analysed by a holistic approach and multiplexed SRM

D.D. Deutsch¹, T. Frohlich¹, K. A. Otte¹, E. Wolf^{1,2}, Georg J. Arnold^{*1}

¹Laboratory for Functional Genome Analysis LAFUGA, Gene Center, Ludwig-Maximilians-Universitaet München. ²Molecular Animal Breeding and Biotechnology, Department of Veterinary Sciences, Ludwig-Maximilians-Universitaet München, Veterinaerstrasse, D-80333 Munich, Germany. *Corresponding author: arnold@genzentrum.uni-muenchen.de

Available Online: 31 December 2015

ABSTRACT

Purpose: Early embryogenesis is a highly critical period of mammalian development. Morphology of pre-implantation development has been well studied, but molecular processes, particularly at the level of proteins, are still poorly understood. To identify molecular key players during early development, we performed holistic proteome analyses using the bovine model, which shows crucial similarities to the human reproduction system. Using multiplexed SRM assays, the dynamism of protein expression during embryo development was precisely quantified during all characteristic stages from the oocyte to the expanded blastocysts.

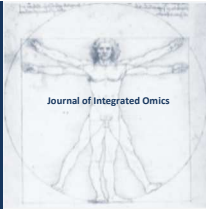
Experimental description: In a holistic approach, four biological replicates of bovine embryos (100 per replicate) at the zygote, 2-cell and 4-cell stage and MII oocytes as a reference were quantitatively analysed by an iTRAQ LC-MS/MS approach on an Orbitrap mass spectrometer. Data processing and bioinformatics analysis comprised MASCOT v2.4, DAVID Bioinformatics Resources 6.7, Self-organizing tree algorithm (SOTA), Cytoscape 3.1.0 plug-ins ClueGO v2.0.8 and CluePedia v1.0.9. SRM based quantifications were performed using batches of 10 zygotes or embryos on a Q-Trap 5500 instrument. GraphPad Prism 6.04 was used for statistical analysis (multiple T-tests).

Results: Among 1072 proteins identified in the iTRAQ approach, 87 showed significant abundance differences in the four stages analysed. The proteomes of 2-cell and 4-cell embryos differed most from the reference MII oocyte. Relevant differences between transcript and protein abundance levels were detected, e.g., for WEE2. Especially affected during development are proteins of the p53 pathway, mitosis, energy and lipid metabolism, and members of “The unfolded protein response”. By Principal Component Analysis (PCA), SRM quantifications comprising a panel of only five proteins were shown to discriminate between all four developmental stages analysed. Using an expanded 27-plex SRM assay, proteins were reliably quantified in nine developmental stages. Absolute protein contents were quantified for nine proteins (n=6).

Conclusions: Despite silence of transcription and translation, the proteome in zygotes and early embryonic stages is highly dynamic. SRM quantification in nine developmental stages revealed high-resolution profiles of the dynamic embryo proteome. Absolute quantification of nine developmentally relevant proteins by SRM revealed protein contents of low femtomole down to attomole range per embryo, and provides an essential data basis for systems biology oriented pathway modelling and prediction algorithms.

Keywords: embryo, oocyte, development, mass spectrometry, SRM, proteomics

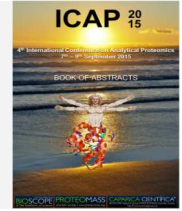
Acknowledgements: This work was supported by a grant from Deutsche Forschungsgemeinschaft (DFG FOR1041 “Germ Cell Potential” AR 362/7-1; WO 685/14-1).



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

A new view of brain function: quantifying protein responses to learning and memory with reverse phase protein arrays

K. J. Gardiner*

Linda Crnic Institute for Down Syndrome, Departments of Pediatrics and Biochemistry and Molecular Genetics, University of Colorado School of Medicine, Aurora, Colorado, 80045, USA. *Corresponding author: gardiner@ucdenver.edu

Available Online: 31 December 2015

ABSTRACT

Purpose: Clinical trials with a diverse array of drugs designed to improve cognition in intellectual disability (ID) have been conducted, are in progress or are planned. Preclinical evaluations of these drugs are generally limited to the study of a single mouse model, use only male mice, and do not generate information on the molecular links between drug action and the functions of the genes causing ID. In addition, analysis of brain expression in learning/memory studies has been typically confined to screening of oligonucleotide arrays or measurement of fewer than 5-10 proteins. Use of quantitative proteomics in preclinical evaluation of drug responses would lead to understanding of the molecular basis of ID and its rescue with drug treatments and could contribute to design of more effective clinical trials.

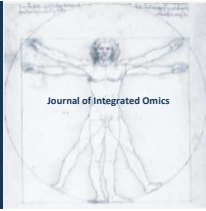
Experimental description: We use the technique of Reverse Phase Protein Arrays (RPPA) in which protein lysates are spotted robotically onto nitrocellulose coated microscope slides. A single slide can accommodate 4000 spots, sufficient for 200 samples in 3-4 replicates of a 5 point sample dilution series. Slides are screened with antibodies to proteins of interest in a protocol of blocking and washing similar to that used in Western blots. RPPA provides advantages of sensitivity, accuracy, throughput, and requirement of minimal sample volume. We investigate changes in protein expression in mouse models of neurological disorders. We generate protein profiles from multiple brain regions, in female and male animals, of different ages, with and without exposure to tests of learning/memory, with and without drug treatment. In a typical experiment, we measure levels of ~100 proteins/protein modifications, including 15 components of both MAPK and MTOR pathways, multiple subunits of glutamate receptors, apoptosis and immediate early gene proteins, in 20-100 mice. For this, RPPA is an ideal technique.

Results: (1) Molecular responses to tests of learning and memory and to drug treatments are more complex and involve a greater diversity of pathways and complexes than previously known. Responses do not propagate linearly within a pathway, likely reflecting the complexities of cross talk among pathways. (2) Gender differences are significant: in hippocampus of untreated control mice, levels of 40% of proteins are significantly higher in females than in males. (3) Gender differences are differentially perturbed in female and male mouse models of neurological disorders, e.g. in one mouse model, in female mice, 20% of hippocampal and 70% of cerebellar proteins are perturbed, but in male mice, only 40% and 20%, respectively, are perturbed. (4) A drug treatment that rescues a deficit in learning/memory does not normalize the protein profile, but instead creates a new pattern of protein expression compatible with successful learning.

Conclusions: RPPA facilitates accurate quantitation of several hundred proteins in hundreds of samples, thus capturing differences among many experimental variables, as well as identifying inter-individual variation. Preclinical evaluations of candidate drugs should include protein expression profiling, in multiple genetic models, in multiple tests of learning/memory, and most importantly, in both female and male mice. Such analyses could lead to identification of effective combinations of drugs with a greater probability of positive outcomes in human clinical trials.

Keywords: protein lysate arrays, antibody, MAP kinase, hippocampus, cerebellum, pathway analysis, mouse models

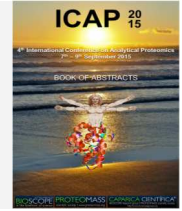
Acknowledgements: Funded by the National Institutes of Health (USA) and the Linda Crnic Institute for Down Syndrome (University of Colorado School of Medicine).



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Dysregulation of IFN alpha and IL-10 in central nervous system in transgenic porcine Huntington's disease model

Karla Jarkovska¹, Ivona Valekova¹, Eva Kotrcova¹, Jan Motlik¹, Suresh Jivan Gadher^{*2}, Hana Kovarova¹

¹Institute of Animal Physiology and Genetics, Academy of Sciences of the Czech Republic, v.v.i., 27721 Libečov, Czech Republic. ²Thermo Fisher Scientific - Life Science Solutions, 7335 Executive Way, Frederick, Maryland 21704, USA. *Corresponding author: Suresh.Gadher@thermofisher.com

Available Online: 31 December 2015

ABSTRACT

Purpose: Huntington's disease (HD) is an inherited neurodegenerative disorder which is progressive and fatal. Any preventive or disease-modifying therapies are not available as yet, hence a better understanding of the earliest changes in brain as well as peripheral cells could be crucial for development of new therapeutic approaches aimed at elimination or repair of the impairing changes caused by huntingtin mutation.

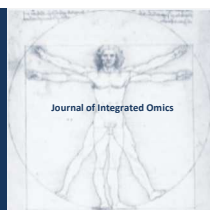
Experimental description: we utilised transgenic porcine HD model bearing an N-terminal fragment of human mutant huntingtin (mHTT) to profile cytokine levels in cerebrospinal fluid, serum and secretomes of microglial cells and blood monocytes in the course of HD using bead based multiplexing Luminex xMAP technology.

Results: among the most pronounced changes was the decline of IFN α in CSF of transgenic animals and this was observed from very early time intervals of HD. IFN α was also decreased in secretome of microglial cells but not blood monocyte in transgenic animals. In addition, IL-10 was lower in CSF as well as microglia secretome. On the contrary, elevated levels of pro-inflammatory IL-1 β and IL-8 were produced by microglial cells of transgenic animals. We further demonstrated higher proportion of the mHTT related to endogenous HTT in microglial cells compared to blood monocytes in transgenic minipigs which may have a causative impact on cytokine production.

Conclusion: in this study, we revealed emerging role of IFN α and IL-10 in central nervous system inflammation and immune response imbalance in HD progression.

Keywords: porcine Huntington's disease model, IFN α , IL10, multiplexing, cerebrospinal fluid, microglial cells.

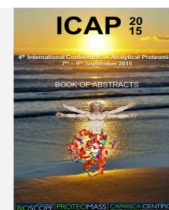
Acknowledgements: this work was supported by the CHDI Research Agreement A-8248, the Operational Program Research and Development for Innovations (EXAM; CZ.1.05/2.1.00/03.0124) and Institutional Research Concept RVO67985904 (IAPG, AS CR, v.v.i). Thermo Fisher Scientific partnered this study to further evaluate the Swine Cytokine Magnetic 7-plex Panel (Invitrogen™ LSC0001M, Thermo Fisher Scientific Inc., formerly LSC0001 from Life Technologies) as well as provided scientific, experimental and statistical analysis support including the algorithm used in this study for data evaluation.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Targeted quantitative proteomics in pharmaceutical sciences

Michael Zhuo Wang *

Department of Pharmaceutical Chemistry, School of Pharmacy, University of Kansas, Lawrence, KS, USA 66047. *Corresponding author: michael.wang@ku.edu

Available Online: 31 December 2015

ABSTRACT

Purpose: Drug metabolizing enzymes (DMEs) and drug transporters (DTs) are important determinants of drug efficacy and adverse effect. Liver is the major organ of drug disposition and hepatic cytochrome P450 enzymes (P450s), flavin-containing monooxygenases (FMOs), and sinusoidal membrane transporters (e.g., OATP1B1 and OCT1) have clinical significance in mediating drug-drug interactions. The goal of this study was to develop LC-MRM-based targeted proteomic methods for absolute quantification of P450s, FMOs, OATP1B1 and OCT1 in human liver samples and evaluate the protein quantification results with orthogonal assays.

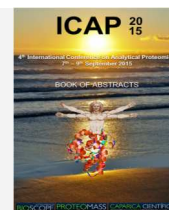
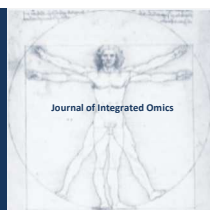
Experimental description: Several panels of individual donor human liver microsomes (HLMs) were either prepared in house or purchased from commercial vendors and used for P450 and FMO quantification. Total membrane proteins were extracted from individual donor human liver tissues using the ProteoExtract Native Membrane Protein Extraction Kit. Signature (surrogate, proteotypic or AQUA) peptides were selected in silico based on previously reported criteria and at least two signature peptides were chosen for each protein. Protein samples were reduced, alkylated, and trypsin-digested using optimized protocols. Absolute protein quantification was achieved by using two types of calibration standards. One contained trypsin digests of recombinant proteins of known concentrations and the other contained synthetic signature peptides of known concentrations.

Results: LC-MRM-based targeted proteomic methods were developed for quantification of human hepatic DMEs (CYP3A4, CYP3A5, CYP3A7, CYP2C9, CYP2C19, CYP4F2, CYP4F3B, FMO1, FMO3, and FMO5) and transporters (OATP1B1, OATP1B3, OATP2B1, OCT1 and P-gp). The absolute protein concentrations of these proteins were determined using HLMs or total membrane proteins. CYP4F enzymes contributed significantly (~15%) to the overall P450 content in HLMs. Ethnic and age-dependent variability in the abundance of these proteins were also observed, e.g., fetal-adult developmental switch between CYP3A4 and CYP3A7, as well as between FMO3 and FMO1. Absolute quantification using protein digests as standards conferred good coherence between different signature peptides of the same protein, whereas signature peptide-dependent quantification was observed using synthetic peptide standards.

Conclusions: LC-MRM-based targeted proteomics provide a multiplexed quantification strategy for DMEs and DTs. Absolute protein quantification of these proteins are expected to improve our understanding of the underlying causes of interindividual variability of drug response. In addition, determination of absolute protein concentrations in human tissues and surrogate models (e.g., sandwich-cultured human hepatocytes and liver-on-a-chip) will enable physiologically-based pharmacokinetic (PBPK) modeling to predict drug exposure in humans prior to clinical trials.

Keywords: targeted proteomics, quantitative pharmacology, drug metabolizing enzymes, drug transporters, quantitative pharmacology

Acknowledgements: This work was supported in part by research grants from Eli Lilly & Co. and NIH.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Quantitative mass spectrometric measurement of multiple serum apolipoproteins: ready for validation and clinical application

Nico P.M. Smit^{*1}, Irene van den Broek¹, Fred P.T.H.M. Romijn¹, Jan Nouta¹, Yuri E.M. van der Burgt², Arnoud van der Laarse¹, Christa M. Cobbaert¹.

¹Leiden University Medical Center, Department of Clinical Chemistry and Laboratory Medicine, Leiden, the Netherlands . ²Leiden University Medical Center, Center for Proteomics and Metabolomics, Leiden, the Netherlands. *Corresponding author: N.Smit@lumc.nl

Available Online: 31 December 2015

ABSTRACT

Purpose: Direct High and Low Density Lipoprotein cholesterol (HDL-C and LDL-C) tests are currently used for cardiovascular disease (CVD) risk classification but suffer from serious drawbacks, especially in dyslipidemic patients [1]. Therefore, we aimed to develop an alternative mass spectrometry (MS) based method for absolute and direct quantitation of the well-defined, clinically relevant serum apolipoprotein counterparts. The MS-based method should have an analytical performance in line with quality specifications, derived from biological variation.

Experimental description: Sample preparation was performed using a modification of the protocol described earlier for apo A-I and B100 [2] using deoxycholate for protein denaturation. Two proteotypic peptides were selected for each of the apolipoproteins (apo) A-I, B100, C-I, C-II, C-III, E as well as additional peptides to distinguish the apo E2, E3 and E4 isoforms [3]. All peptides were detected by multiple reaction monitoring using a 1290 LC-system coupled to a 6490 triple quadrupole mass spectrometer (Agilent) [2,4]. Stable isotope labeled peptides were added before digestion for internal standardization. External calibration was performed using commutable, serum based calibrators. In case of apo A-I and B100, traceability to WHO-IFCC international reference standards is established (www.bipm.org/jctlm). The method is validated according to Clinical and Laboratory Standards Institute guidelines EP15-A and EP9-A2, and the different steps outlined in the EAS-EFLM framework [5]-

Results and Conclusions: Apo A-I and B100 concentrations can be measured by MS with imprecision below the minimal specifications of 5.0 and 5.3 %, respectively [2]. The apo C-II, C-III and E protein concentrations could be measured with imprecisions < 9.2%. LC-MS/MS quantitation of apo A-I, B100, C-II, C-III and E in 54 normo- and 46 hypertriglyceridemic (NTG and HTG) samples correlated well with immunoturbidimetric assays ($R^2 > 0.96$). Good correlations were found between the two peptides of these proteins ($R^2 > 0.95$), and apo E2 and E4 isoforms could be distinguished from the most common E3 isoform. Our method thus allows individual detection and quantitation of six apolipoproteins with additional apo E phenotyping. Using a Bravo workstation and a fully automated protocol for sample preparation about 80 specimens can be analysed in singlicate per day.

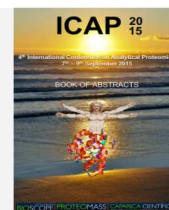
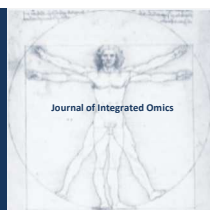
We conclude that our multiplexed apo method is ready for clinical application, both in routine patient care and in clinical or epidemiological studies. Its added value compared to conventional serum HDL-C/LDL-C tests for CVD risk assessment and/or treatment monitoring is currently subject of clinical validation studies.

Keywords: apolipoproteins, cardiovascular disease, traceability, multiple reaction monitoring

Acknowledgements: We thank Agilent Technologies for support and fruitful discussions .

References:

- [1] Langlois M. et al. Atherosclerosis 2014; 233:83-90
- [2] Smit N.P. et al. J Proteomics 2014; 109C: 143-61.
- [3] Martinez-Morillo E. et al. J Proteome Res 2014; 13(2): 1077-87
- [4] Van den Broek I. et al. J Proteome Res 2013; 12: 5760-74.
- [5] Horvath A.R. et al. Clin Chim Acta 2014; 427: 49-57.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Proteomic analysis and μ -, δ - and κ -opioid receptor content in brain cortex of rats exposed to increasing doses of morphine for 10 days; comparison with animals sacrificed 20 days since the last dose of morphine

H. Ujcikova*, K. Stolarova, K. Cechova, P. Svoboda

Laboratory of Biochemistry of Membrane Receptors. Department of Biomathematics. Institute of Physiology of the Czech Academy of Sciences. Videnska 1083. 142 20 Prague 4. Czech Republic. *Corresponding author: ujcikova@biomed.cas.cz

Available Online: 31 December 2015

ABSTRACT

Purpose: The aim of this work was to detect the morphine-induced change of μ -, δ - and κ -OR in rat forebrain cortex as well as the alteration of the total spectrum of proteins by this drug when acting for long period of time.

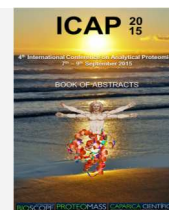
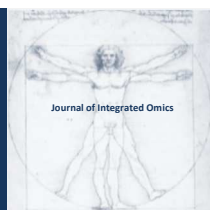
Experimental description: Rats were exposed to increasing doses of morphine for 10 days (10-50 mg/kg) and sacrificed 24 hours (group +M10) or 20 days (group +M10/ - M20) after the last dose of this drug (in parallel with control animals: groups - M10 and - M10/ - M20). The post-nuclear supernatant fraction (PNS) was analyzed for the content of μ -, δ - and κ -opioid receptors by 1D-SDS-PAGE performed under the non-dissociated (- DTT) and dissociated (+DTT) conditions. The individual receptor subtypes were recognized by immunoblot detection with specific antibodies. PNS fraction was also resolved by 2D-ELFO and analyzed by MALDI-TOF MS/MS and *label-free* quantification using the MaxLFQ intensity determination and normalization procedure introduced by Cox et al. in 2014.

Results: (A) The total amount of μ -OR, δ -OR and κ -OR was unchanged by morphine when resolved by 1D-SDS-PAGE under the non-dissociated conditions (- DTT). However, the resolution under the dissociated conditions (+DTT) revealed the presence of multiple protein bands exhibiting the wide range of $M_r \approx 20$ -106 kDa. All these immunoblot signals were also unchanged with the exception of minor isoforms of μ -OR ($M_r \approx 20$ -32 kDa) and δ -OR ($M_r \approx 68$ kDa, \pm DTT) which were decreased in morphine-treated samples, $p < 0.05$. Determination of Na,K-ATPase indicated no difference between control and morphine-treated samples of PNS. This result was verified by binding of selective inhibitor of this enzyme, [³H]ouabain. Noticeably, our data indicated a significant increase of Caveolin-1, $p < 0.05$, but no change of Caveolin-2. (B) Proteomic analysis of the total protein composition by MALDI-TOF MS/MS indicated the morphine-induced change of 27 proteins (+M10). Number of altered proteins was decreased to 14 in rats sacrificed 20 days since the last dose of morphine (+M10/ - M20). The high resolution by *label-free* quantification identified 118 altered proteins (+M10) and this number was decreased to 41 (+M10/ - M20).

Conclusions: The morphine-induced decrease of μ -OR and δ -OR in rat forebrain cortex was surprisingly small; κ -OR was unchanged. Contrarily, depending on sensitivity of method used for proteomic analysis, the 27 (MALDI-TOF MS/MS) or 118 (*label-free* quantification) proteins were altered after 10 days of morphine exposure. In rats sacrificed 20 days after the last dose of morphine, these numbers were decreased to 14 and 41, respectively.

Keywords: morphine, long-term exposure, rat forebrain cortex, post-nuclear supernatant fraction, μ -, δ - and κ -opioid receptors, 1D- and 2D-SDS-PAGE, MALDI-TOF MS/MS, *label-free* quantification, MaxLFQ.

Acknowledgements: This work was supported by the GACR (P207/12/0919, P304/12/G069) and by the Academy of Sciences of the Czech Republic (RVO: 67985823).



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Advantages and disadvantages of chromatographic approaches to study bacterial metalloproteomes

James P. Barnett¹ David J. Scanlan² and Claudia A. Blindauer^{*1}

¹Department of Chemistry, University of Warwick, Coventry CV4 7AL, United Kingdom. ²School of Life Sciences, University of Warwick, Coventry, CV4 7AL, United Kingdom. *Corresponding author: c.blindauer@warwick.ac.uk

Available Online: 31 December 2015

ABSTRACT

Purpose: Although it is now clear that at least a third of all proteins require a metal cofactor for function, it is still not straightforward to predict metal requirements and binding sites of proteins. This is especially true for the plethora of "hypothetical proteins" emerging from genome sequencing that have no biochemically characterised homologues. Indeed, a previous study concluded that "microbial metalloproteomes are largely uncharacterized" [1]. Analytical approaches are therefore needed to begin filling the gaps [2]. We are particularly interested in bacterial zinc proteomes [3], and have been exploring chromatographic separation techniques to fractionate metalloproteomes. This talk will briefly summarise the major conclusions from our work so far.

Experimental description: The marine cyanobacterium *Synechococcus* sp. WH8102 was cultured in artificial seawater. Cells were harvested and lysed, and lysates were fractionated by different chromatographies (size exclusion, anion exchange, IMAC). Fractions were analyzed by 1D SDS-PAGE, peptide mass fingerprinting, and ICP-MS.

Results: Separation by 2D orthogonal chromatography did not lead to the identification of major zinc-requiring proteins. Instead, most intracellular zinc was found in fractions with high levels of phycobilisome proteins. The latter are highly abundant in cyanobacteria, and not only caused problems with the detection of lower abundance proteins, but were also likely to lead to zinc re-distribution [4]. In contrast, similar approaches were able to detect intact Ni-superoxide dismutase [5], highlighting the differences in the bio-coordination chemistry of these two metal ions. Further experiments employing immobilised zinc affinity chromatography led to enrichment of several proteins with zinc-binding ability, including some unexpected, highly interesting new candidates.

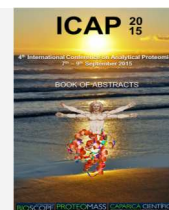
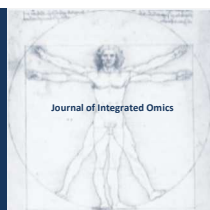
Conclusions: Even the mildest separation conditions may disturb cellular zinc speciation, and care must be taken to avoid misinterpretation of analytical results.

Keywords: metalloproteomics, zinc, 2D-LC, ICP-MS.

Acknowledgements: We thank the Leverhulme Trust (F/00 215/AY) and NERC (NE/F004249/1), Advantage West Midlands and the European Regional Development Fund (Birmingham Science City).

References:

- [1]- A. Cvetkovic, A. L. Menon, M. P. Thorgersen, J. W. Scott, F. L. Poole, F. E. Jenney, W. A. Lancaster, J. L. Praissman, S. Shanmukh, B. J. Vaccaro, S. A. Trauger, E. Kalisiak, J. V. Apon, G. Siuzdak, S. M. Yannone, J. A. Tainer and M. W. W. Adams, *Nature*, 2010, 466, 779–782.
- [2]- J. P. Barnett, D. J. Scanlan and C. A. Blindauer, *Anal. Bioanal. Chem.*, 2012, 402, 3311–3322.
- [3]- J. P. Barnett, A. Millard, A. Z. Ksibe, D. J. Scanlan, R. Schmid and C. A. Blindauer, *Front. Microbiol.*, 2012, 3, 142.
- [4]- J. P. Barnett, D. J. Scanlan and C. A. Blindauer, *Metalomics*, 2014, 6, 1254–1268.
- [5]- J. P. Barnett, D. J. Scanlan and C. A. Blindauer, *Anal. Bioanal. Chem.*, 2012, 402, 3371–3377.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Mechanisms underlying metabolic disorders deciphered using quantitative proteomics of non-sequenced species

F. Bertile*

CNRS – Université de Strasbourg, UMR7178, Institut Pluridisciplinaire Hubert Curien (IPHC), Département Sciences Analytiques, Laboratoire de Spectrométrie de Masse Bio-Organique, 25 rue Becquerel, 67087, Strasbourg, France. *Corresponding author: fbertile@unistra.fr

Available Online: 31 December 2015

ABSTRACT

Purpose: Since the last few years, there is growing evidence that important questions related to human health may simply not be answered by only studies in the mainstream of laboratory murine models. In this context, there is an increasing interest on wild animals as a source of new information. The aim is here to present examples where proteomics applied to exotic species can help understanding how certain animals do support environmental conditions that would be detrimental to human health.

Experimental description: We examined the impact of different conditions (e.g. food scarcity, weightlessness and physical inactivity, high-fat diets) on the proteome of various tissues collected from laboratory rodents and also from wild penguins, space-flown mice, wild bears and selected lines of voles. Because no protein or even genomic sequence was available for most of these species, we performed both classical database searches and de novo sequencing followed by Blast searches to identify a maximum of proteins. Quantitative proteomics was then achieved using a combination of complementary approaches, including 2D-DIGE/MS and label-free MS-based proteomics (spectral counting and/or on the basis of XICs).

Results: In non-sequenced species, the use of de novo sequencing allowed identifying 20% more proteins in comparison with the use of only classical Mascot searches. To obtain this result, de novo sequencing was performed only from high quality spectra as filtered using the Recover module of our software suite (<https://msda.unistra.fr/>). In any studied species, hundreds to thousands of proteins were identified (FDR<1%) then quantified, and automatically classified according to their function. We notably found drastic differences in the plasma proteome of breeding penguins vs. that of laboratory rats in response to prolonged fasting, in the muscle proteome of mice after a 30-day space flight vs. that of bears after 5 months of physical inactivity, and in the brown adipose tissue (BAT) proteome of voles selected for their BAT activity level (high vs. low) in response to a high-fat diet. These proteomic data provided essential metabolic insights in mechanisms related to modern human pathologies, including undernutrition, muscle atrophy, obesity and diabetes.

Conclusions: Studying diversity of animal adaptations/responses to environmental constraints can provide new basic knowledge of importance for biomedicine. To this end, de novo sequencing and quantitative proteomics strategies offers unique possibilities to provide a number of consistent results.

Keywords: exotic species, de novo sequencing, quantitative proteomics, functional annotations, pathway analysis

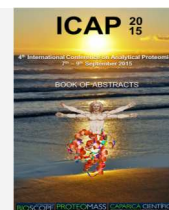
Acknowledgements: This work was funded by the Centre National d'Etudes Spatiales (CNES), the CNRS and Strasbourg University (H2E project; IDEX UNISTRA) and the French Proteomic Infrastructure (ProFI; ANR-10-INSB-08-03).



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Aptamers in cancer research

A. Drabik^{*1}, J. Ner-Kluza¹, Laia Civit², Günter Mayer², Jerzy Silberring¹

¹AGH University of Science and Technology, Kraków, Poland. ²University of Bonn, LIMES Institute, Bonn, Germany. **Corresponding author: drabik@agh.edu.pl*

Available Online: 31 December 2015

ABSTRACT

Introduction: Along with the generation of the term “magic bullet” by Ehrlich a great concept arises of a cancer treatment method, where only tumor cells would be killed by an ideal therapeutic agent.

The targeted therapy has become a main goal for conquer cancer. Most therapeutic agents in current clinical use, such as chemotherapy, radiotherapy, and immunotherapy are not specific for the molecules responsible for human disease, thus distribution of the drug in the organism results in unwanted side effects. Targeted treatment is based on specific ligand recognition.

Aptamers are short single-stranded nucleic acids with a defined three-dimensional shape that allows them to interact with high affinity with a target molecule. They have advantage that they do not induce immunological response. The low immunogenicity and cellular-targeting properties, together with the fact that they can be chemically synthesized in vast amounts might result in suitable drugs with low side effects. They can be used for identification of cell-surface molecules associated with a pathogenic state of a cell.

Purpose: Identification of proteins targeted by aptamers in cancer cells.

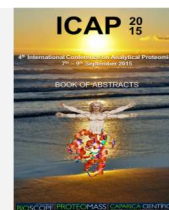
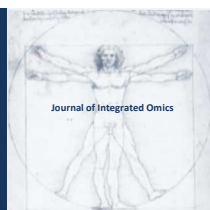
Experimental description: We introduced the proteomic studies of aptamers targets in cancer cell lines PC3 and MCF7. We applied biotinylated forms of aptamers characterized by the strong affinity to streptavidin to isolate target proteins. We have performed traditional bottom-up proteomic approach to analyze the group of proteins that bind to A26 and A33 aptamers that can serve as potential metastases molecular markers and targets for selective chemotherapy. We have found that cell surface receptors and proteins localized inside the cells are capable of binding A26 and A33.

Results: Molecular targets of aptamers in cancer cells were identified. Based on the presented research we indicate the importance of the applicability of aptamers in many technologies including selective diagnostics and therapeutics.

Conclusions: Aptamers can be considered as effective tools, since they possess unique properties to aid in cancer diagnosis, prevention, and treatment.

Keywords: aptamer, proteomics, cancer.

Acknowledgements : This work was supported by the grant "META" No. 5/EuroNanoMed/2012



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Quantitative proteomics in human drug metabolism

Brahim Achour¹, Matthew R. Russell¹, Francesco Lanucara², Amin Rostami-Hodjegan^{1,3}, Jill Barber^{*1}

¹Manchester Pharmacy School, University of Manchester, Stopford Building, Oxford Road, Manchester, UK, M13 9PT. ²Waters Corporation, Stamford Avenue, Altrincham Road, Wilmslow, UK, SK9 4AX. ³Simcyp limited (a Certara Company), Blades Enterprise Centre, John Street, Sheffield, UK, S2 4SU. *Corresponding author: jill.barber@manchester.ac.uk

Available Online: 31 December 2015

ABSTRACT

Purpose: The aim of this study was to develop reproducible, accurate methods for determining the concentrations of enzymes and transporters involved in human drug metabolism.

Experimental description: We have developed QconCAT-based LC-MS/MS methodology for quantifying the cytochrome P450 and UGT enzymes and the drug transporters in human tissue. The QconCATs (artificial proteins consisting of labelled standard peptides, released by proteolysis) initially did not express in *E. coli* and methods for expressing recalcitrant QconCATs were developed as a part of the project. Specifically, we fused the two QconCATs (MetCAT and TransCAT which quantify enzymes and transporters respectively) to proteins with known characteristics. The MetCAT was fused to a QconCAT protein, known to express in inclusion bodies in high yield, whereas the TransCAT was fused to a soluble construct and expressed in cytosol.

The MetCAT has now been used to quantify the enzymes in a panel of 24 livers, and the TransCAT has been used to quantify transporters in four intestinal samples. We have also analysed three liver samples using label-free methodology.

Results: Cytochrome P450 and uridine 5'-diphospho-glucuronosyltransferase enzymes are responsible for much of the metabolism of drugs in the liver, intestine and other tissues. There is, however, substantial inter-individual variability in their concentrations, as well as variation between groups of individuals (ethnic groups, age groups). Transporter proteins are responsible for further variations in how a given dose of a drug may lead to different plasma and tissues in different people and groups.

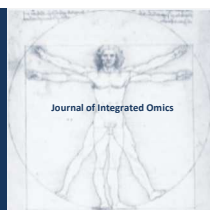
Our results show wide variation in a population between the levels of the different enzymes, for example CYP3A4, which metabolizes many important drugs, is present with a range 10.4–262.1 pmol mg⁻¹ protein. We determined several correlations between enzymes, including a positive correlation between levels of CYP3A4 and CYP2B6.

Most recently, label-free quantification has been shown to give broadly comparable results and, in addition, to give quantification data on other proteins in the samples.

Conclusions: Personalized medicine is crucially dependent on an understanding of the concentrations of drug-metabolizing enzymes and transporters in human tissue. Quantitative LC-MS/MS is a powerful tool for understanding these enzymes and transporters, and QconCAT and label-free quantification methods deliver complementary but consistent information.

Keywords: QconCAT, label-free, quantification (quantitation), cytochrome P450, UGTs.

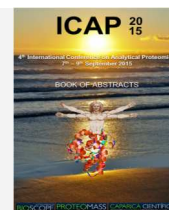
Acknowledgements: Mass spectrometry was performed at the Michael Barber Centre for Mass Spectrometry, University of Manchester.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Scorpion venom proteomics and transcriptomics

Mohamed A. Abdel-Rahman*

Zoology Department, Faculty of Science, Suez Canal University, Ismailia 41522, Egypt. **Corresponding author:*
mohamed_hassanain@science.suez.edu.eg; dr_moh_71@hotmail.com

Available Online: 31 December 2015

ABSTRACT

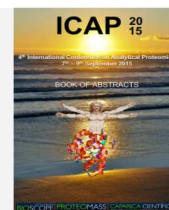
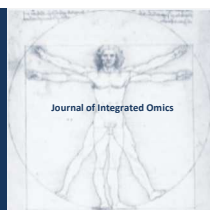
Background: Scorpions (Arthropoda, Arachnida) occupy vast territory of the world and their ability to produce and deliver venoms is an essential factor in this success. Scorpion venom is a very complex mixture of substances, where peptides are the major constituents with various biological and pharmacological properties. Excluding the phenomena of intraspecific diversity, it is estimated that more than 150,000 different peptides/proteins expressed in the venom glands of 1500 different scorpion species.

Experimental description: During the last decade, the cutting edge OMIC technologies of proteomics and transcriptomics have been largely employed to deeply understand structure and molecular diversity of animal venoms from various species including scorpions.

Results: To date, using proteomic analyses, there are 5314 unique molecular masses have been identified from the venom of 27 different scorpions. Mass fingerprinting data clearly showed remarkable differences in the number of venom molecules (ranging from 60 to 665) between scorpion species. Few of the 5314 molecules were fully characterized which mainly belonging to neurotoxins, antimicrobial peptides and bradykinin potentiating peptides. Also, the venom gland proteome of 20 different scorpion species has been revealed using a combination of transcriptomic and bioinformatics methods and resulting in about 73 thousands ESTs. These sequences are corresponding to neurotoxins, antimicrobial peptides, housekeeping proteins, hypothetical proteins and a large number of unassigned types of scorpion venom peptides/proteins.

Conclusions: The main advantages of using transcriptomics in scorpion venomomics are: (i) adding new insights about biological processes taking place in scorpion venom glands; (ii) facilitating the identification of various novel biologically active peptides and (iii) drawing an accurate image about interspecific and intraspecific venom diversity. Recent studies of scorpion venomomics will be discussed, with special focusing on antimicrobial peptides derived from scorpion venoms.

Keywords: Scorpion venomomics, Proteomics, Transcriptomics, Venom gland, cDNA library, Antimicrobial peptides.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Comparative proteomic profiling of brain tissues of rat with different behavioral characteristics during emotional stress.

N.E. Sharanova*, N.V. Kirbaeva

Institute of Nutrition, Moscow, Russia. *Corresponding author: sharanova@ion.ru

Available Online: 31 December 2015

ABSTRACT

Purpose: Evidence from animal studies has demonstrated that the limbico-reticular complex is initially involved in emotional responses. The amygdale and hippocampus are the most multifunctional structures of the limbic system. The reticular formation is directly connected with the emotions due to promote new and significant information to the different brain regions. To identify the differences of proteomic expression in rat brain caused by emotional stress we performed comparative proteomic analysis.

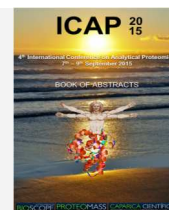
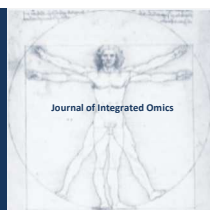
Experimental description: Wistar male rats were divided on 2 groups of behaviorally passive and active animals. Immobilization of rats during 12h (from 9p.m. till 9a.m.) served as a model of acute emotional stress. The protein expression profiles of hippocampus, amygdale, reticular formation and cortex were studied using by two-dimensional electrophoresis and MALDI-TOF.

Results: The proteomic analysis showed the down-regulated expression of GFAP, septine-5, calreticulin, alpha-synuclein, phosphatidylethanolamine-binding protein 1, thiomorpholine-carboxylate dehydrogenase, prohibitin, glutamine synthetase and up-regulated expression of lactate dehydrogenase, F-actin-capping protein subunit alpha-2 in different parts of brain depending on behavioral type of rats and the stage of stress.

Conclusions: Thus, individual behavioral features affect the specific pathway of organism response to the stress and determine an adaptive potential of the organism.

Keywords: brain, emotional stress, comparative proteomics, behavior

Acknowledgements: Prof. S.Pertsov from P.K.Anokhin Research Institute of Normal Physiology, Moscow, Russia.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Cell-free lysates for the production of difficult proteins

F. Bernhard^{*1}, D. Foshag², S. Rupp³, Ernst Thuer⁴, V. Dötsch¹, E. Henrich¹, R. Rues¹

¹Institute of Biophysical Chemistry, Centre for Biomolecular Magnetic Resonance, J.W. Goethe-University, Frankfurt-am-Main, Germany. ²Institute for Interfacial Engineering and Plasma Technology IGVP, University of Stuttgart, Stuttgart, Germany. ³Fraunhofer Institute for Interfacial Engineering and Biotechnology IGB, Stuttgart, Germany. ⁴Comparative Genomics Group, CRG-Centre for Genomic Regulation, Barcelona, Spain. *Corresponding author: fbern@bpc.uni-frankfurt.de

Available Online: 31 December 2015

ABSTRACT

Purpose: Cell-free production of proteins and other biomolecules has emerged during the last decade as an important core technology in the new field of synthetic biology. Lysates of either eukaryotic or prokaryotic organisms can be employed for the synthesis of particularly difficult proteins such as membrane proteins, large multisubunit assemblies or highly oxidized proteins. Cell-free expression provides an open and accessible reaction environment. Additionally, functional properties of the lysate proteomes can easily be modulated in order to generate ideal conditions for the production of high quality samples of individual target proteins.

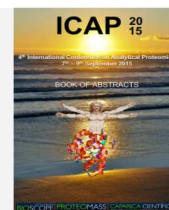
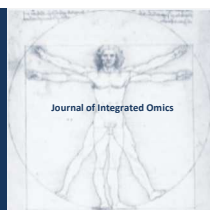
Experimental description: Protocols for the preparation of cell-free lysates have been developed and optimized. Libraries of additives have been screened for the manipulation and optimization of lysate proteomes.

Results: We present strategies for the manipulation of cell-free lysates and reaction conditions for the efficient production of difficult proteins such as G-protein coupled receptors or enzymes. We further exemplify the optimization of protein sample quality by directed fractionation of lysate proteomes and selective inactivation of particular enzyme classes and pathways. Examples focusing on the structural and functional characterization of membrane proteins will be discussed.

Conclusions: Lysate proteomes for the cell-free production of biomolecules can be adjusted according to the requirements of individual targets.

Keywords: cell-free expression, bacterial lysates, membrane protein environments, synthetic biology, artificial proteomes.

Acknowledgements: The work was funded by the Collaborative Research Center (SFB) 807 of the German Research Foundation (DFG) and by the German Ministry of Education and Science (BMBF). Support was further obtained by Instruct, part of the European Strategy Forum on Research Infrastructures (ESFRI).



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Linear Algebra Analysis and Quantification of Arginine Dimethylation in Histone Modification using Liquid Chromatography–Tandem Mass Spectrometry-Based Targeted Proteomics

Yun Chen*

School of Pharmacy, Nanjing Medical University, 818 Tian Yuan East Road, Nanjing, 211166, China. *Corresponding author: ychen@njmu.edu.cn

Available Online: 31 December 2015

ABSTRACT

Purpose: Protein methylation at arginine residues is a prevalent post-translational modification in eukaryotic cells that has been implicated in processes from RNA-binding and transporting to protein sorting and transcription activation. Three main forms of methylarginine have been identified: NG-monomethylarginine (MMA), asymmetric NG,NG-dimethylarginine (aDMA), and symmetric NG,NG-dimethylarginine (sDMA). Current methods are able to predict and determine thousands of methylarginine sites, whereas stoichiometric distinction and quantification of these methylarginine forms, especially aDMA and sDMA are still challenging.

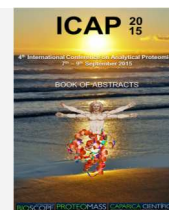
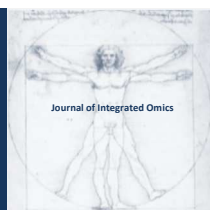
Experimental description: Liquid chromatography coupled with tandem mass spectrometry (LC–MS/MS)-based targeted proteomics is emerging as a promising technique for site-specific quantification of protein modification using proteolytic peptides as surrogates of proteins. However, the routine targeted proteomics assay can not easily distinguish the contribution of aDMA and sDMA. In this study, linear algebra algorithms were employed as an add-on to targeted proteomics to retrieve information on individual dimethylarginine peptides.

Results: A LC–MS/MS-based targeted proteomics assay was first developed and validated for each dimethylarginine peptide. The linear algebra analysis of aDMA and sDMA was achieved from LC-MS/MS of their common spectra using an internal standard and a calculated response factor for each peptide. Finally, we applied this approach to determine the stoichiometry of histone methylation in breast cancer cells and tissue samples

Conclusions: LC –MS/MS-based targeted proteomics assay combined with linear algebra algorithms has a potential for simultaneously determining various methylated forms in protein modification.

Keywords: Protein methylation, aDMA, sDMA, histone, linear algebra, liquid chromatography–tandem mass spectrometry, targeted proteomics

Acknowledgements: The National Natural Science Fund (21175071), the project sponsored by SRF for ROCS, SEM (39), the Jiangsu Six-type Top Talents Program (D), and the Open Foundation of Nanjing University (SKLACLS1102) awarded to Dr. Chen.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Integrated Proteomic and Metabolomic Approaches Reveal Energy Disruption in Epilepsy

J. Adamec^{*1}, R A. Grove¹, C. Boone¹, K. K. Samson², H. M. Roundtree², H. Kylo³, T. Simeone², K. Simeone², D. Madhavan³

¹Department of Biochemistry and Redox Biology Center, University of Nebraska-Lincoln, Lincoln, NE, USA. ²Department of Pharmacology, Creighton University School of Medicine, Omaha, NE, USA. ³Department of Neurological Sciences, University of Nebraska Medical Center, Omaha, NE, USA. *Corresponding author: jadamec2@unl.edu

Available Online: 31 December 2015

ABSTRACT

Purpose: Epilepsy affects ~ 1% of the human population and despite extensive study, the precise nature of epilepsy is poorly understood. Recently we have evaluated the levels and types of post-translational modifications (PTMs) of proteins in surgically removed epileptic brain tissue, which includes seizure onset, irritative and silent zones. The overall goal of this study was to identify enzymes and metabolites responsible for asynchronization of energy metabolism.

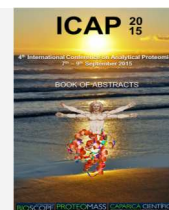
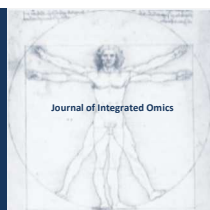
Experimental description: Tissue used included surgical resections from patients with epilepsy as well as a Kv1.1 KO mouse model of familial epilepsy. Non targeted lipid analysis was carried out on a Bruker Solarix FT-ICR by LC/MS. Amino acid analysis was performed using the aTRAQ method on an ABSciex 4000 Q-Trap. Proteome screening for oxidative modification (carbonylation) was performed using 2-D DIGE followed by peptide analysis on a Thermo Velos Pro LTQ. Adenosine energy metabolites were determined through etheno adduct formation and fluorescent detection by HPLC.

Results: Our data revealed tissue specific accumulation of oxidatively damaged proteins in the seizure onset zone. The most significant protein affected was identified as GAPDH, a key enzyme of energy metabolism interconnecting glycolysis (GP), pentose phosphate pathway (PPP) and tricarboxylic acid cycle (TCA). Moreover, higher levels of oxidative stress were observed in onset zones. Together with fact that epilepsies which respond well to the ketogenic diet (KD) have impaired glucose uptake our data suggests that higher level of oxidative stress impacts GAPDH activity, thus reducing levels of TCA intermediates and resulting in temporal asynchronization of GP, PPP and TCA, ultimately decreasing ATP levels. This imbalance can trigger an epileptic seizure. Seizures, on the other hand, can be prevented by supplying cells with TCA intermediates or their precursors such as ketone bodies in KD treatment.

Conclusions: Energy and redox balance is primarily controlled by GP, PPP and TCA. Under normal conditions glucose is transformed to pyruvate by GP, producing a small amount of ATP. The vast majority of ATP is generated in mitochondria through TCA and oxidative phosphorylation which utilizes pyruvate as a substrate. Under oxidative stress, glucose transformation is diverted from GP to PPP resulting in an increase in cellular reducing power through increased NADPH formation. Products of PPP are linked to GP through the enzyme GAPDH. Our data indicates that higher levels of oxidative stress partially inactivate GAPDH, a key enzyme interconnecting GP, PPP and TCA. This reduces the levels of metabolic intermediates available for TCA, leads to asynchronization of GP, PPP and TCA, and to a temporal decrease in ATP levels. Imbalance in the pathways involved in energy metabolism and redox balance, together with low ATP levels, result in neuronal network hyperexcitability. This phenotype can be reduced or prevented by supplying cells with TCA intermediates or their precursors, such as the case in KD treatment.

Keywords: Epilepsy, Energy metabolism, Oxidative damage.

Acknowledgements: This work was supported by Phase III RBC IDEa program funding from NIH



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Methods for the preparation of acute myeloid leukemia patient samples for proteomic and phosphoproteomic analysis

M. Hernandez-Valladares*¹, E. Aasebø¹, F. Berven¹, Ø. Bruserud², F. Selheim¹

¹PROBE, Building for Basic Biology, University of Bergen, Jonas Lies vei 91, N-5009 Bergen, Norway. ²Department of Internal Medicine, Haukeland University Hospital, N-5020, Bergen, Norway. *Corresponding author: Maria.Hernandez-Valladares@uib.no

Available Online: 31 December 2015

ABSTRACT

Purpose: The study of global protein expression in acute myeloid leukemia (AML) patients by mass spectrometry (MS) can help identifying differential expression and post-translational modifications of proteins that could represent disease-related biomarkers for early diagnosis or for improved prognostics and to predict the patient response to different therapeutics. To optimize the proteome and phosphoproteome coverage of samples from AML patients by LC-MS/MS analysis, we have tested several methods with different peptide fractionation and phosphopeptide enrichment strategies.

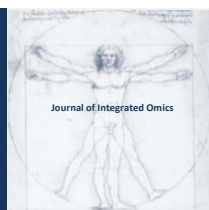
Experimental description: Peptide samples were prepared with an in-solution digestion protocol, using urea or guanidinium hydrochloride as denaturant, and with the filter aided sample preparation (FASP) methodology. Peptide fractionation was carried out with reverse phase/strong cation exchange (SCX) and SCX disks in a stage-tip. Techniques for phosphopeptide enrichment included metal oxide affinity chromatography (MOAC), immobilized metal affinity chromatography (IMAC) and sequential elution from IMAC (SIMAC).

Results: From an initial screening of the different samples on a Linear Trap Quadrupole (LTQ) Orbitrap Elite MS, we found the best proteome coverage with the sequential FASP method, with Lys-C and trypsin as digestion enzymes, which identified and quantified 3100 proteins from 20 µg of sample. With the same strategy, followed by a separate MOAC/TiO₂-beads enrichment of the two peptide pools, we identified and quantified 2.900 phosphorylation sites from only 250 µg of AML patient sample. On a QExactive HF hybrid Quadrupole-Orbitrap MS, 5400 proteins and 4000 phosphosites were identified and quantified from these FASP-prepared samples.

Conclusions: To improve the proteome and phosphoproteome coverage of AML patient samples by MS analysis to discover new biomarkers, testing of different methods can be beneficial. Based on sample preparation optimization in our laboratory, we have chosen the FASP protocol to prepare AML patient samples for MS-based proteomic and phosphoproteomic studies.

Keywords: AML, proteomics, phosphoproteomics, sample preparation.

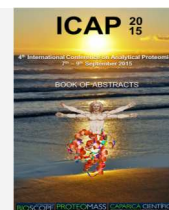
Acknowledgements: The authors would like to thank the Norwegian Cancer Society and Øyvinn Mølbach-Petersens Fond for the funding of this work.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Comparison of 2D proteomic maps revealed properties of *Ambrosia artemisiifolia* sub-pollen particles accounting for more severe asthma symptoms than its whole pollen grains

K. Smiljanic^{*1}, A. Mohamed¹, S. Trifunovic¹, M. Perusko¹, L. Mihajlovic¹, J. Ognjenovic² and T. Cirkovic Velickovic¹

¹Centre of Excellence for Food Molecular Sciences, Faculty of Chemistry, University of Belgrade, Serbia. ²Department of Biochemistry and Molecular Genetics, University of Illinois at Chicago, IL 60607, U.S.A. *Corresponding author: katarinas@chem.bg.ac.rs

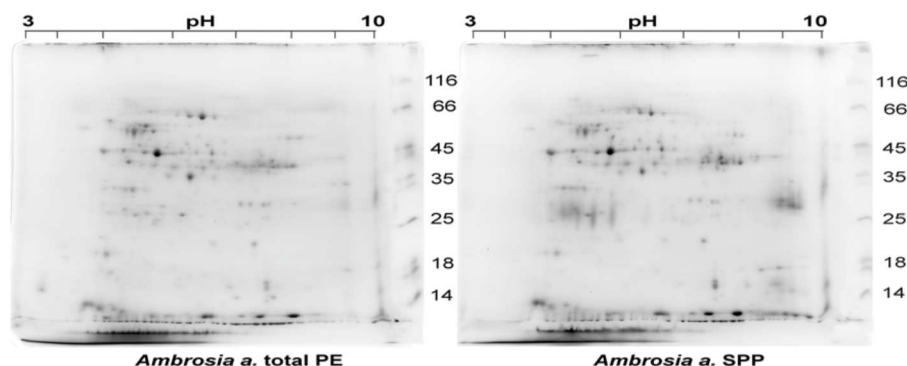
Available Online: 31 December 2015

ABSTRACT

Aims and scopes: It is known that sub-pollen particles (SPP) cause more severe symptoms of asthma than its whole pollen grain counterparts, due to its smaller size and ability to penetrate deeper into the lungs. To reveal other possible causes of such more severe asthma symptoms induced by *Ambrosia artemisiifolia* SPP, its sub-pollen particle and pollen grain proteomes were characterized and compared.

Experimental description: Protein extract of short ragweed (*Ambrosia artemisiifolia*) pollen and its SPP were prepared and subjected to denaturing 2-D electrophoresis. Pollen proteome spots were excised after colloidal coomassie blue brillinat (cCBB) staining and in gel digested for liquid chromatography coupled with high resolution LTQ Orbitrap XL hybrid mass spectrometry. Parallel to that, cCBB stained gels were analyzed and quantified with laser scanner Typhoon 7000 series and Image 2D Master Platinum 7.0 software (GE Healthcare, USA).

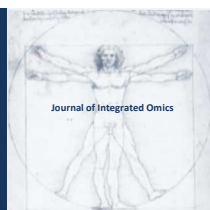
Results: There is statistically significant difference between the contents of major allergen Amb a 1.05 subgroup in the ragweed whole pollen grains and SPP, the latter being richer in Amb a 1.05 (2 times), in major allergen Amb a 11 (5 times), in minor allergens Amb a 4 (7 times) and Amb a 6 (4 times). The 30 kDa basic antigen group in SPP (8 times more abundant) needs further investigation.



Conclusions: Beside its smaller size and hence easier penetrability, short ragweed SPP possess significantly higher load of major *Ambrosia artemisiifolia* allergens, Amb a 1.05 and Amb a 11, minor allergens Amb a 4 and Amb a 6 which could contribute to more severe asthma symptoms caused by SPP.

Keywords: Ambrosia pollen, sub-pollen particles, short ragweed, asthma, Amb allergens.

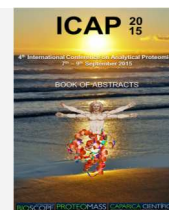
Acknowledgements: This research was carried out with the support from the Ministry of Education and Science of the Republic of Serbia, project no. 172024, and by the European Commission, under the Framework 7, project RegPot FCUBERA, GA No 256716.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Effect of a quorum sensing inhibitor on the *Pseudomonas aeruginosa* PAO1 proteome

Shunmugiah Thevar Karutha Pandian*

Department of Biotechnology, Alagappa University, Science Campus, Karaikudi 630 004, India. *Corresponding author: k_pandian@rediffmail.com

Available Online: 31 December 2015

ABSTRACT

Purpose: Research in the field of quorum sensing (QS) inhibition resulted in the identification of numerous QS inhibitors (QSIs). But their mode of action and impact on proteome of bacterial pathogens remains unclear. Hence, the aim of the present study is to find out the molecular mechanism of QSI and its effect on cytoplasmic proteome of *P. aeruginosa* PAO1.

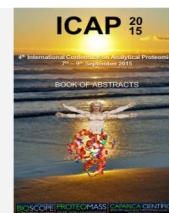
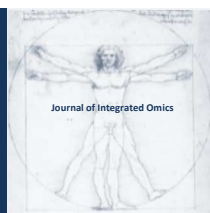
Experimental description: Cytoplasmic proteins of *P. aeruginosa* PAO1 grown in the absence and presence of 5 µg of QSI were extracted by sonication. For 2DE, 600µg each of cytoplasmic proteins from control and treated samples were subjected to isoelectric focusing using immobilized pH gradient (IPG) strips (18 cm, pH 3-10 NL (GE Healthcare)) at 20°C using standard parameters for 18 h. Then the IPG strips were reduced and alkylated for 20 min. After this step, strips were placed on 12 to 15% gradient sodium dodecyl sulphate-polyacrylamide gels (Ettan DALT six, GE Healthcare) for the second dimension and electrophoresis was performed. Protein spots were visualized by colloidal Coomassie brilliant blue G-250. The gels were scanned with Image scanner III (GE Healthcare) and the raw images were analyzed using ImageMaster 2D Platinum 7. Experiments were performed in biological triplicates. More than two fold differentially regulated protein spots were selected for in-gel trypsin digestion and analyzed using MALDI TOF/TOF (AXIMA Performance, Shimadzu.Biotech). The proteins were identified by peptide mass fingerprinting using the software tool MS-Fit (<http://prospector.ucsf.edu>).

Results: Among the detected spots (613), 48 (7.8%) spots are downregulated and 31 (5%) spots are upregulated by more than two fold. Based on statistical significance (ANOVA 0.05), seventeen downregulated and five upregulated protein spots were selected for MALDI TOF/TOF analysis. Downregulated proteins are involved in iron transport, transcriptional regulation, antibiotic resistance, chemotaxis and two component signaling systems. QSI treatment upregulated the expression of histidine kinase, ABC-transporter protein, putative transcriptional regulator, hypothetical proteins G655_17540 and PSPA7_1308.

Conclusions: Since iron transport proteins, two-component response regulators and sensor are involved in the activation of protease, exotoxin A, and pyoverdine biosynthesis proteins in *P. aeruginosa*, downregulation of above said proteins by QSI could be a major event in attenuation of virulence factor production. In addition, QSI also blocked the expression of proteins involved in the pyoverdine and pyochelin synthesis proteins. Most notably, putative iron-sulfur proteins which are known to express highly in biofilms are significantly downregulated upon QSI treatment. In conclusion, QSI inhibits the virulence factor production by targeting the iron homeostasis, pyoverdine and pyochelin biosynthesis pathways.

Keywords: quorum sensing inhibitor; *P. aeruginosa* PAO1; quorum sensing.

Acknowledgements: The financial support from ICMR and CSIR is thankfully acknowledged. The Instrumentation Facility provided by DST (FIST & PURSE) and UGC (SAP-DRS1) and Computational/Bioinformatics Facility provided by DBT (BIF) are gratefully acknowledged.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Immune responses of silkworm, *Bombyx mori* against infection by an endoparasitoid

A.R.Pradeep* Pooja M, Anitha J, Shambhavi P.H., Awasthi A.K, Geetha NM, Ponnuvel K.M and Kanika Trivedy.

Proteomics Division, Seribiotech Research Laboratory, Central Silk Board, CSB-Kodathi Campus, Bangalore – 560035, India. Website: <http://www.sbrl.res.in> *Corresponding author: arpradeepnair@gmail.com

Available Online: 31 December 2015

ABSTRACT

Purpose: In insects it is known that humoral and cellular immunity plays pivotal role in defending the body against microbial and macro-bial infection. Humoral immunity is provided by activation of immune - associated genes and its transcripts whereas the cellular immunity is provided by the cells in the circulating haemolymph, the hemocytes. Recently we showed immuno - competence of the larval integument which forms the primary barrier between environment and internal milieu through proteomics approach. On the other hand, hemocytes spread over the parasitic surface and encapsulate it. However, the uzi fly infection induced toxicity in hemocytes leading to detoxification reactions. We thus elucidated differential immune responses of integumental epithelium and hemocytes induced through infestation by a dipteran endoparasitoid, *Exorista bombycis* in the economically important silk producing insect and a lepidopteran model, *Bombyx mori*.

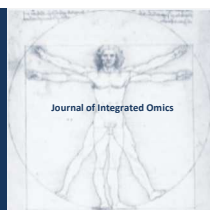
Experimental description: Sub cellular responses of integumental epithelium and hemocytes were analyzed by transmission electron microscopy. Immune responses of integumental epithelium were analyzed by SDS-PAGE, 2D electrophoresis, in-gel digestion and mass spectrometry. Toxic and detoxification responses of hemocytes were analyzed by SDS-PAGE and mass spectrometry and transcriptome responses by microarray analysis. Gene expression was performed by Real - time PCR (qPCR) analysis. All data were analyzed statistically using ANOVA, PCA and regression analysis.

Results: Integumental epithelium showed signs of autophagy and apoptosis and differential expression of associated genes after the infestation. Proteomic analysis revealed enhanced expression of innate immunity components of toll and melanization pathways, cytokines, signaling molecules, chaperones, and proteolytic enzymes demonstrating diverse host responses. qPCR analysis revealed upregulation of genes encoding cytokine, spatzle, BmToll, and NF kappa B transcription factors Dorsal and BmRel. NF kappa B inhibitor cactus showed diminished expression when Dorsal and BmRel were upregulated, revealing a negative correlation ($R^2 = 0.612$). The hemocytes showed signs of toxicity and activation of certain detoxification mechanisms. Microarray and qPCR showed up regulation of genes associated with detoxification mechanism.

Conclusions: Using Proteomic tools, we showed activation of immune proteins in integumental epithelium which is a first time report in any invertebrates. Moreover, endoparasitoid - induced toxicity and activation of detoxification mechanism in hemocytes could be revealed by mass spectrometry and microarray analysis, thereby exposed the differential responses of two tissues to macroparasitic infestation.

Keywords: immunity; detoxification; mass spectrometry; microarray; *Bombyx mori*; parasitism.

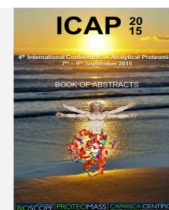
Acknowledgements: To Central Silk Board for permission and laboratory facilities. To Department of Biotechnology, Government of India for funding through research projects to ARP.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Proteomic characterization of Colorectal Cancer tissues reveals tumor stage- and EGFR-specific CRC biomarkers

Manveen K.Sethi¹, Morten Thaysen-Andersen¹, Nicolle H. Packer¹, Mark S. Baker², William S. Hancock², Susan Fanayan^{*2}

¹Department of Chemistry and Biomolecular Sciences, Macquarie University, Sydney, NSW 2109, Australia. ²Department of Biomedical Sciences, Faculty of Medicine and Health Sciences, Macquarie University, Sydney, NSW 2109, Australia. **Corresponding author: susan.fanayan@mq.edu.au*

Available Online: 31 December 2015

ABSTRACT

Purpose: To investigate the total membrane-enriched proteome derived from a cohort of paired tumorigenic and adjacent non-tumorigenic colon tissues from CRC patients, to identify deregulated proteins associated with CRC, the individual TNM stages as well as the EGFR status of the CRC tissues

Experimental description: Membrane proteins were extracted by Triton-X114 phase partitioning, separated by gel electrophoresis, in-gel trypsin digested into 10 fractions and subjected to label-free LC-MS/MS analysis by LTQ orbitrap XL. Identified proteins were analyzed by advanced pathway analysis to identify proteomes and the associated networks, based on tumor stages and EGFR expression

Results: Various cancer associated proteins, including CRC-specific proteins, were found to be significantly up- or down-regulated in CRC tumors relative to adjacent normal tissues. Advanced proteomic analysis identified networks and pathways such as, immunological diseases, cancer and cell to cell signalling to be up-regulated and cellular functions, movements and assembly to be down-regulated in CRC tumor.

We further employed a proteomics-driven approach to provide a comprehensive view of the CRC tissue proteomes and the associated networks, based on tumor stages and EGFR expression.

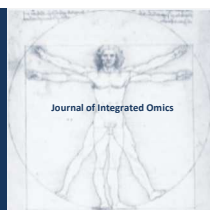
Among proteomic features distinguishing between early and late stage CRC tumors were Increased expression of several cytoskeletal proteins, including cytokeratins (KRT2, KRT18, and KRT19), tubulins (TUBB2A, TUBB2B and TUBB6), actins (ACTB and ACTBL2) and filamins (FLNA and FLNC), which were significantly overexpressed in late stage CRC tumors.

Of note was the up-regulation of the signature CRC metastasis-associated proteins, JUP, EPCAM and SDCBP, as well as cytoskeletal proteins associated with tumor metastasis (ACTB, KRT18, KRT19, TUBB2A, TUBB2B, TUBB6, DES, FLNA and FLNC), observed exclusively in late stage CRC tumors, independent of EGFR expression status. In EGFR⁺ tumors, on the other hand, exclusive EGFR-dependent overexpression of cell adhesion molecules (CEACAM5, 6 and 7) was observed, exclusive of tumor stage.

A close association between EGFR expression status and tumor stage was also observed through similar expression patterns of certain cancer-specific proteins. Among these were the exclusive observation of ERBB2 and its interacting partners (GRB7 and SRC) in EGFR⁺ and late stage tumors (absent in EGFR⁻ and early stage tumors), as well as elevated expression of members of heat shock proteins (HSP90AB1, HSPA1A, HSPD1, HSPA8 and HSPD1) in EGFR⁺ and late stage tumors. Similar expression profiles for cytoskeletal (SPTAN1, SPTBN1, TPM1, TPM3, CAPZA1, CAPZA2, CDH1 and LIM1) and cell adhesion proteins (CTNNA1, CTNNA2, DSG2 and LSGALs4) as well as several glycotransferases (FUT3, FUT8 and ST6GALNAC1), mucins (MUC2, MUC13 and FCGBP) and ribosomal proteins were observed in both EGFR⁺ and early stage CRC tumors.

Conclusions: This work provides a detailed map of the altered proteome and associated protein pathways in CRC. Expression profiles of cancer-associated proteins, including several CRC-specific proteins, involved in tumor growth, progression and metastasis, influenced by EGFR expression status, tumor stage or both, were identified. The insights gained from this study may open avenues for a knowledge-driven search for candidate CRC protein markers.

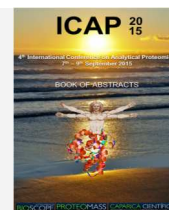
Keywords: colorectal cancer, membrane-enriched proteome, label free shotgun proteomics, epidermal growth factor receptor (EGFR), TNM tumor stage .



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Probing the structure and conformational changes in complex biological systems using zero-length crosslinking and mass spectrometry

David W. Speicher^{*1,2}, Roland Rivera-Santiago^{1,2}, Sandra Harper¹, and Sira Sriswasdi³

¹Center for Systems and Computational Biology, The Wistar Institute, Philadelphia, PA 19104 USA. ²Department of Biochemistry and Molecular Biophysics, University of Pennsylvania, Philadelphia, PA 19104 USA. ³Department of Biological Sciences, University of Tokyo, Tokyo, 113-0032 Japan. **Corresponding author: speicher@wistar.org*

Available Online: 31 December 2015

ABSTRACT

Purpose: Chemical cross-linking and mass spectrometry (CX-MS) combined with molecular modeling is a powerful structural MS strategy. Zero-length cross-linkers are particularly useful for refining and confirming structural models because they provide the most precise distance constraints between cross-linked sites. However, zero-length cross-linked peptides are difficult to identify, and therefore this method has been used far less frequently than longer cross-linkers such as the popular amine-reactive homobifunctional reagents. Due the great value and under-utilization of zero-length cross-link analysis, we have developed methods to enhance its routine application to diverse biological problems.

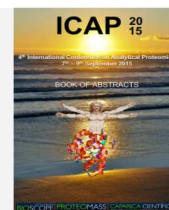
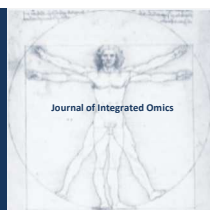
Experimental description: Our group has greatly improved in-depth detection of zero-length cross-links by optimizing mass spectrometry data acquisition for this application, and by developing a software tool, ZXMiner, specifically for zero-length cross-link identification. The strategy utilizes label-free LC-MS/MS quantitative comparisons of tryptic digests from zero-length cross-linked samples and uncross-linked controls to identify putative cross-linked peptide precursors. Spectra from all precursors within 5 ppm of theoretical cross-linked peptides are compared to predicted cross-linked peptide theoretical spectra. Graphical tools facilitate verification of cross-link assignments. Resulting high-confidence peptide identifications are then employed to identify novel interactions and as distance restraints for molecular modeling experiments.

Results: We initially applied this method to large segments of the human red cell protein, spectrin using a Thermo Orbitrap XL mass spectrometer. The data acquisition method and scoring algorithm were subsequently optimized on a Thermo Q Exactive Plus mass spectrometer to achieve improved depth of analysis on complex samples. This method was applied to purified human red cell plasma membranes as a complex physiological model system. Using a 4-hour gradient without sample fractionation, 57 proteins could be identified by five or more peptides, which represents a total search space of almost 5 million Da of unique sequence. Representative results identified 65 cross-linked peptides at an FDR=0. Most cross-links were associated with the most abundant proteins, although 3 cross-links involved several of the least abundant proteins in the database. The most abundant proteins in the membrane are the anion exchanger (AE1) and spectrin, which contained 12 and 34 identified high confidence cross-links, respectively. These data have been used to develop detailed structural models and to map locations of interactions with other membrane proteins.

Conclusions: CX-MS using zero-length cross-linking can be applied to multi-million Da complexes to identify novel interactions and to provide precise distance constraints for molecular modeling and development of experimentally verified structures of large protein complexes.

Keywords: zero-length crosslinking, structural mass spectrometry, protein structure, macromolecular complexes.

Acknowledgements: Funded by grants from the National Institutes of Health and Philadelphia Healthcare Trust.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

The O₂-Affinity and Cooperativity of Hemoglobin Are Regulated by the Effector-Linked 4D-Structural Changes (Protein Dynamics) rather than the Ligand-Linked 3D-Structural Changes

Takashi Yonetani^{*1}, Kenji Kanaori²

¹Univ. of Pennsylvania, Department of Biochemistry & Biophysics, Philadelphia, USA yonetant@mail.med.upenn.edu. ²Kyoto Institute of Technology, Department of Bio-Molecular Engineering, Kyotyo, Japan. *Corresponding author: yonetant@me.com

Available Online: 31 December 2015

ABSTRACT

Introduction: The current, widely-held mechanism of allostery of hemoglobin (Hb) has been that the changes from the R-quaternary/tertiary structures of oxy-Hb to the T-quaternary/tertiary structures of deoxy-Hb exert certain structural constraints on the coordination structure of the heme group, leading to a lower O₂-affinity of the hemes and thus that of Hb [1]. However, we found that there is no causal correlation between the static T/R-quaternary structures and the low/high O₂-affinities of Hb [2]. We explore an alternative mechanism of the regulation of the ligand-affinity and cooperativity in Hb.

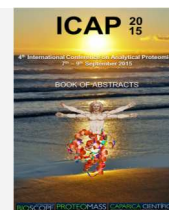
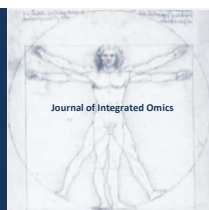
Results and Discussion: The O₂-affinities of free complexes Fe[II]- and Co[II]-protoporphyrins with axial nitrogenous bases in organic solvents are very low ($P_{50} > 10^2 \sim 10^3$ Torr), whereas the apparent O₂-affinities of these metalloporphyrins, which are incorporated in proteins such as apo-myoglobin, apo-Hb, serum albumin, etc., increase substantially to $P_{50} < 10^{-1} \sim 10^4$ Torr, though their coordination structures are apparently unchanged [3]. Such substantial increases in the apparent ligand-affinities of metalloporphyrin-containing proteins are accomplished by preventing/inteferring with the dissociation of the un-bonded ligand by protein matrix, since the interior of globin is nearly fully packed by protein matrix. In Hb and Co-substituted Hb (Co-Hb), the dissociation process of the un-bonded ligand proceeds through the “caged” state [4-6], which can be produced by cryogenic photolysis of the ligated-states at 4.2K and in which the metal-ligand bond is broken and the un-bonded ligand is trapped near the bonding site within the globin moiety. This “caged” state has spectral features distinct from those of either deoxy- or ligated states of the respective hemoproteins. The apparent ligand-affinities (P_{50} , K_T , and K_R , where $P_{50} = 2/[K_T + K_R]$) of Hb and Co-Hb are regulated by heterotropic effectors, as observed by Bohr [7] without detectable changes in either static T-/R-quaternary/tertiary structures of the globin moiety or the coordination/electronic structures of the metalloporphyrin moiety and thus the ligand-affinity of the metalloporphyrins themselves [8-10]. The substantial reduction of the apparent ligand-affinities of Hb and Co-Hb by heterotropic effectors, may be caused by increase in the migration rate of ligands through globin matrix from the “caged” state to solvent, resulting from the effector-linked, enhanced high-frequency (>GHz) thermal fluctuations which increase the transparency of the globin matrix toward small diatomic ligands such as oxygen, CO, and NO [8-10], which may concomitantly reduce the rate of geminate-recombination of the ligand to the metalloporphyrins. Thus, we structurally identify Monod-Wyman-Changeux (MWC)’s “T”-low- and “R”-high- affinity- functional states [11] as high- and low-amplitude, high-frequency (>GHz) thermally fluctuating dynamic states of Hb, respectively, regardless of its static T-/R-quaternary/tertiary structures [10].

Conclusion: The ligand-affinity of Hb is regulated through the 4D structural changes (or protein dynamics) induced by heterotropic effectors [9], rather than the ligand-linked, static T-/R-quaternary/tertiary structural changes, as proposed by Perutz [1]. Thus, the “caged” state of Hb acts as a critical transition state in regulation of the affinity for small diatomic ligands in Hb and Co-Hb [10].

Keywords: Allostery, Protein Dynamics, Cooperativity, Hemoglobin

References:

- [1] M.F. Perutz, Nature 1970 228, 726;
- [2] T. Yonetani, et al., JBC 2002 277, 34508;
- [3] H. Yamamoto, et al., Bioinorg. Chem. 1977 7, 189;
- [4] T. Iizuka, et al., BBA 1974 351, 182;
- [5] T. Iizuka, et al., BBA 1974 371, 126;
- [6] T. Yonetani, et al., JBC 1974 249, 2168;
- [7] C. Bohr, et al., Skand. Arch. Physiol. 1904 15, 401;
- [8] T. Yonetani, M. Laberge, BBA 2008 1784, 1146;
- [9] M. Laberge, T. Yonetani, Biophys. J. 2008 94, 2737;
- [10] T. Yonetani, K. Kanaori, BBA 2013 1834, 1873;
- [11] J. Monod, et al. J. Mol. Biol. 1965, 12, 88.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Flagellar proteins and their key roles in human diseases: Once neglected, still underestimated

D. M. Oliveira^{*1,2}

¹RENORBIO Northeastern Brazil Biotechnology PhD Program. Universidade Estadual do Ceará (UECE). Campus do Itaperi. 60.714.903. Fortaleza, CE. Brasil. ²Nutrition and Health Master's Program. Centro de Ciências da Saúde. UECE. 60.714.903. Fortaleza, CE. Brasil.
^{*}Corresponding author: diana.magalhaes@uece.br

Available Online: 31 December 2015

ABSTRACT

Organelle proteomics, such as the flagellum, has been one of the most suitable approaches used to identify novel components and modifications of a given cell, tissue, organ or even organism. Information is usually gained at a large scale and is very valuable to further understand biological processes of the given compartment. In this presentation I will talk about a particular subset of flagellar proteins, which happen to be amazingly related to protozoa virulence in Leishmaniasis. As all eukaryotic flagella, the *Leishmania* flagellum is a protein nanomachine, but it has a particular way in being arranged to invade human macrophages. Yes, I just said to invade, rather than to be engulfed or phagocytosed by macrophages. *Leishmania* flagellar apparatus turns the protozoan into an “infection motor” by means of complex, intricate surface and shallow subsurface features that provide major biological processes in this trypanosomatid survival and virulence, from motility and cell division, towards invasion and persistence in host cells. Using conventional proteome assays and computational molecular modeling coupled with atomic force microscopy (AFM), my research group has studied specific morphological data resulting from protein structural organization of *Leishmania* spp. flagellum, gathering information not yet explored neither in a nanostructured nor in a pathogenic approach for this species. AFM imaging allows detailed visualizations of the flagellum surface, as can be seen by Figure 1, which, in a 3D-view, shows an undulated surface made of two sided border-like elevations along all the elongated flagellum and the red arrow illustrates one of these; the inner region between these two elevations, a major groove, measured around 200nm in average (redline). Figure 1b also shows a prominent membrane surface of flagellar adhesion zone (FAZ) next to the cell body. The underlying hypothesis of all such studies is that polymerization and depolymerization of parasite actin and actin motor-associated proteins, during both processes of motility and host cell entry, might be key events for successful infection, including the parasite survival within phagosomes. Our reports have revealed an actin-polymerization *Leishmania* protein, profilin, and its partner, formin, as involved in axonemal assembly/disassembly, therefore, flagellar dynamics / remodeling, while *Leishmania* coronins and Arp2/3 complex are believed to participate in the phagosome formation. We must recall that *Leishmania* has differential levels of survival inside phagosomes, suggesting that the survival of parasites is linked to their ability to be targeted to tight/non-cidal and non-degradative compartments. To what extent these different killing pathways (delayed or not) are used in a given phagosome certainly depends on various cellular and environmental conditions (usually a plethora of macrophage receptors at variable proportions that may influence phagosome fate to variable degrees), including interactions with flagellar proteins here emphasized.

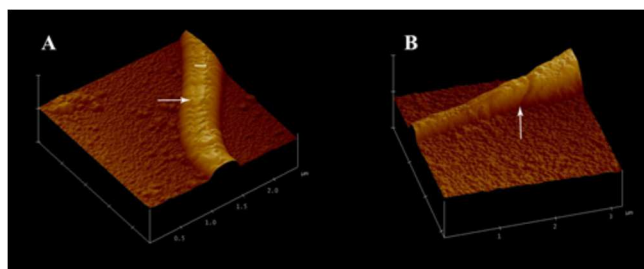
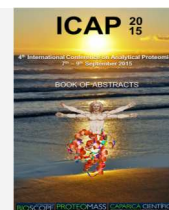
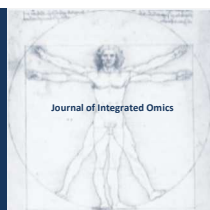


Figure 1. 3D-view images of *L. chagasi* flagellum. (A) Region within the furrow/groove. There is an undulated surface (arrow) made of two sided border-like elevations and a measurement of one groove/furrow whole extension (redline). (B) The adhesion zone (FAZ) to the cell body can be seen (arrow).

Keywords: flagellar proteins; organelle proteome; molecular modeling; atomic force microscopy (AFM); phagosome.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Effects of heavy metals on *Cyanothece* sp. CCY 0110 growth, extracellular polymeric substances (EPS) production, ultrastructure and protein profiles

R. Mota^{1,2,3}, S. B. Pereira^{1,2}, M. Meazzini⁴, R. Fernandes^{1,2}, A. Santos^{1,2,3}, C. A. Evans⁵, R. De Philippis^{4,6}, P. C. Wright⁵, and P. Tamagnini^{*1,2,3}

¹ i3S - Instituto de Investigação e Inovação em Saúde, Universidade do Porto, Portugal. ²IBMC - Instituto de Biologia Molecular e Celular, Univ Porto. Rua do Campo Alegre, 823. 4150-180. Porto Portugal. ³Faculdade de Ciências, Dept. Biologia, Univ. Porto, Rua do Campo Alegre, Edifício FC4, 4169-007. Porto. Portugal. ⁴Dept. Agrifood Production and Environmental Sciences, Univ. Florence. Piazzale delle Cascine 24. I-50144. Florence. Italy. ⁵ChELSI Institute, Dept. Chemical and Biological Engineering, Univ. Sheffield. Hadfield building, Mappin Street. S1 3J. Sheffield. U. K. ⁶Institute of Ecosystem Study, National Research Council (CNR). Via Madonna del Piano, 10. 50019. Sesto Fiorentino. Italy.

*Corresponding author: pmtamagn@ibmc.up.pt

Available Online: 31 December 2015

ABSTRACT

Purpose: The aim of this study was to evaluate the effects of several heavy metals on the growth/survival, EPS production, ultrastructure and protein profiles of the highly efficient extracellular polymeric substances (EPS)-producer cyanobacterium *Cyanothece* sp. CCY 0110.

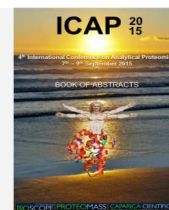
Experimental description: After exposure to different concentrations of metals (Cu²⁺, Pb²⁺, Cd²⁺, or Li⁺), cell growth, production of total carbohydrates and RPS (released polysaccharides) and ultrastructural changes were investigated. Proteomes were compared by two 8-plex iTRAQ studies and results were validated with data from Western blots, O₂ evolution and ROS measurements and enzymatic assays.

Results: The heavy metals affected the cells differently, triggering distinctive responses. Concerning chronic exposure to the metals, the cells were more affected by Cu²⁺ followed by Pb²⁺, Cd²⁺, and Li⁺. The presence of metals led to remarkable ultrastructural changes, mainly at the thylakoid level. The comparison of the proteomes allowed to follow the stress responses and to distinguish specific effects related to the time of exposure and/or the concentration of an essential (Cu²⁺) and a non-essential (Cd²⁺) metal. The majority of the proteins identified with significant fold changes were associated with photosynthesis, CO₂ fixation and carbohydrate metabolism, translation, and nitrogen and amino acid metabolism. Unexpectedly, the amount of released polysaccharides (RPS) was not enhanced by the presence of heavy metals.

Conclusions: This work shows the holistic effects of different heavy metals on the cyanobacterium *Cyanothece* sp. CCY 0110. Overall, the results suggest that during Cu²⁺ chronic exposure the cells adjust their metabolism to invest the spare energy in the activation of metal detoxification mechanisms. In contrast, the toxic effects of Cd²⁺ accumulate over time suggesting that cells might not have the same capacity to deal with this non-essential metal.

Keywords: Cyanobacteria; *Cyanothece*; extracellular polymeric substances (EPS); heavy metals; iTRAQ; proteome.

Acknowledgements: The work was funded by FEDER Funds through the Operational Competitiveness Programme – COMPETE and national funds through FCT – Fundação para a Ciência e a Tecnologia under the project FCOMP-01-0124-FEDER-028314 (PTDC/BIA-MIC/2889/2012) and the scholarships SFRH/BD/84914/2012 and SFRH/BDP/72400/2010. We thank Professor Lucas Stal for providing *Cyanothece* sp. CCY 0110. Sheffield acknowledges the EPSRC (EP/E036252/1).



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Brain ubiquitin mitochondrial subproteome and its changes induced by neurotoxins and neuroprotectors

A.E. Medvedev^{*1}, O.A. Buneva¹, A.T. Kopylov¹, M.V. Medvedeva², O.V. Tikhonova¹, V.G. Zgoda¹

¹Department of Proteomic Research and Mass Spectrometry, Institute of Biomedical Chemistry, 10 Pogodinskaya Street, Moscow, 119121, Russia; ²Moscow State University, Vorobiev Gori 1, Bld.12, Moscow, Russia 119234 Russia. *Corresponding author: professor57@yandex.ru

Available Online: 31 December 2015

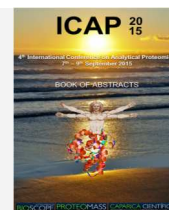
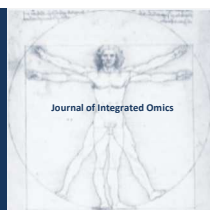
ABSTRACT

Ubiquitin is a 76-residue protein that tags proteins and thus regulates numerous intracellular processes. Incubation of the rat brain mitochondrial fraction with biotinylated ubiquitin *in vitro* followed by avidin-agarose chromatography and mass spectrometry (MS) detection revealed direct ubiquitination of extra- and intramitochondrial proteins. However, the proportion of directly ubiquitinated proteins represented not more than 20% of the total number of identified proteins that specifically bound to avidin-agarose. Profiling of endogenously ubiquitinated proteins of the mitochondrial compartment by means of affinity chromatography on the proteasome subunit S5a conjugated with agarose followed by their MS detection revealed 58 individual proteins identified by a characteristic diglycine fragment of their tryptic peptides. Administration of the neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) inducing symptoms of parkinsonism in mice had a minor effect on the total number of identified proteins (59) but caused qualitative changes in the proteomic profile. Pretreatment with a non-peptide neuroprotector, isatin, decreased MPTP-induced neurotoxicity and significantly reduced the total number of individual ubiquitinated proteins (to 36). Isatin (interacting with a wide range of isatin-binding proteins in the brain and regulating expression of isatin-sensitive genes) administered to control mice also decreased the total number of ubiquitinated proteins (to 30).

Conclusions: The qualitative changes in the ubiquitin mitochondrial subproteome suggest that the neuroprotector effect of isatin is associated with “metabolic immobilization of the brain” induced by interaction of isatin with isatin binding proteins and altered patterns of ubiquitinated proteins of the mitochondrial compartments and inhibition of monoamine oxidase (B) as well.

Keywords: mitochondrial subproteome, affinity-based profiling, ubiquitin, isatin, isatin-binding proteins, MPTP

Acknowledgements: The work was done in the framework of the State Academies of Sciences Fundamental Research Program for 2013-2020 and the Russian Foundation for Basic Research (project nos. 13-04-00161 and 15-04-01545).



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

The Resonant Recognition model to design short bioactive therapeutic peptides: does it really work?

T. Iodeir^{*1}, E. Pirogova²

¹Biosciences, School of Applied Sciences, RMIT University, Bundoora, Victoria, 3083, Australia. ²Biomedical Engineering, School of Electrical and Computer Engineering, RMIT University, Melbourne, Victoria, 3000, Australia. *Corresponding author: taghrid.istivan@rmit.edu.au

Available Online: 31 December 2015

ABSTRACT

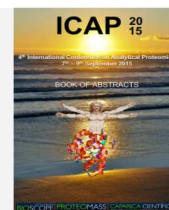
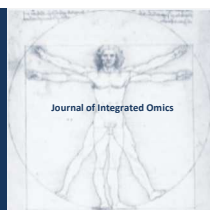
In recent years, more focus has been placed on the role of small molecular weight peptides in clinical medicine, mostly for their ability to penetrate cellular membranes, and to interfere with enzymatic functions or protein-protein interactions. The ability to predict the 3D structures and functions of biological molecules would certainly be useful in designing therapeutic drugs. Hence, the Resonant Recognition Model (RRM), applied in this study, is a physico-mathematical model that incorporates signal processing methodology for structure-function analysis of proteins. The aim of our research is to design and investigate the biological effects of RRM bioactive peptides and their potential to be applied as therapeutic agents.

Experimental description: The RRM approach was used to design 18-22 mer bioactive peptide analogues of natural proteins known to possess cytotoxic effects on cancer cells including Myxoma virus (MV); mammalian interleukins (IL) and tumor necrosis factor (TNF α). The biological effects of RRM-MV; RRM-IL; and RRM-TNF were investigated on mammalian cancer and normal cell lines. The cytotoxic effects were evaluated by qualitative and quantitative cell survival methods. In addition non-bioactive peptides lacking the specific RRM frequencies or peptides with scrambled sequences were also tested as controls. Human apoptosis protein arrays were used to detect the expression levels of pro-apoptotic and anti-apoptotic proteins in treated versus non-treated cancer cells. Glycomic arrays and fluorescent microscopy were used to find the potential cellular targets of these peptides in cancer cells. Furthermore, we also applied the RRM to design short peptides with antimicrobial activity as analogues for known antimicrobial natural peptides like azurocidin (CAP37) and lactoferrins and evaluated their effect on *Escherichia coli*, and Staphylococci including a methicillin resistant *S. aureus* (MRSA) strain.

Results: The bioactive peptides RRM-MV, RRM-IL, and RRM-TNF produced significant apoptotic/necrotic effects on B16 mouse melanoma cells, human melanoma MM96L, and COLO16squamous cell carcinoma, PC3 prostate cancer and MCF7 breast cancer. However, no cytotoxic effects were detected on human red blood cells, normal skin fibroblasts; mouse macrophages, mouse fibroblasts, and Chinese hamster ovary cells (CHO). Yet the non-bioactive, control peptides did not cause any cytotoxic changes in any type of cancer or normal cells. The bioactive peptides were found to be located inside cytoplasmic components in treated cancer cells prior to necrosis stage. Possible cellular targets were predicted to be specific glycoproteins, however the bioactive peptides have a different binding specificity to glycans as Sialy Lewis X, gangliosides and mannoses. Furthermore, the azurocidin peptide analogue had a bacteriostatic antimicrobial effect on all tested bacteria including MRSA.

Conclusions: The RRM is a powerful computational tool to design bioactive peptides with specific biological functions. Our data indicates that each of the RRM designed peptides possessed the desired biological function which was conserved in its unique sequence.

Keywords: Resonant recognition model, RRM-designed therapeutic peptides, anticancer; antimicrobial.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Knots in proteins – a tangled tale

P. Virnau*

Institut für Physik, Staudingerweg 9, Johannes Gutenberg University Mainz, 55128 Mainz, Germany. *Corresponding author: virnau@uni-mainz.de

Available Online: 31 December 2015

ABSTRACT

Ever since Lord Kelvin has conjectured that atoms are composed of knots in the ether, these peculiar objects have stimulated the imagination of mathematicians and natural scientists alike. In recent years the field went through a renaissance and progressed considerably, spurred by the discovery of knotted DNA and proteins.

Even though protein knots are rare, they occur in all kingdoms of life and their topology is typically preserved amongst homologs. Nowadays, knotted protein structures can even be created artificially, and knotted designs could in the future be used in the context of protein engineering. In this talk I will provide an introductory overview to the field and evidence which indicates why the folding of knotted proteins may not be so difficult after all.

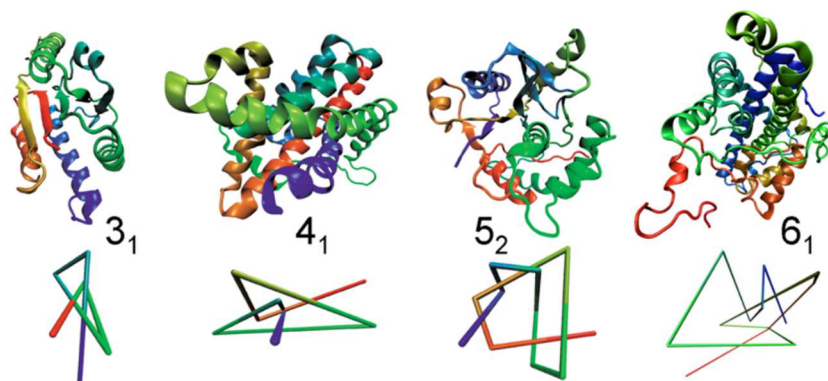
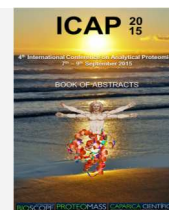
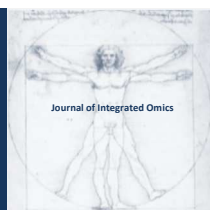


Figure 1. Knotted protein structures. From Bölinger et al, PLoS Comp. Biol. 6, e1000731 (2010).

Keywords: knots in proteins, computer simulations, folding.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Comparative proteomic profile between transgenic and non-transgenic soybean seeds by 2D-PAGE

B. K. de Campos^{*1,2}, M. A. Z. Arruda^{1,2}

¹Spectrometry, Sample Preparation and Mechanization Group –GEPAM, Institute of Chemistry, University of Campinas –UNICAMP, P.O. Box 6154, Campinas, SP 13083-970, Brazil. ²National Institute of Science and Technology for Bioanalytics, Institute of Chemistry, University of Campinas – Unicamp, P.O. Box 6154, Campinas, SP 13083-970, Brazil. **Corresponding author: brunah.campos@hotmail.com*

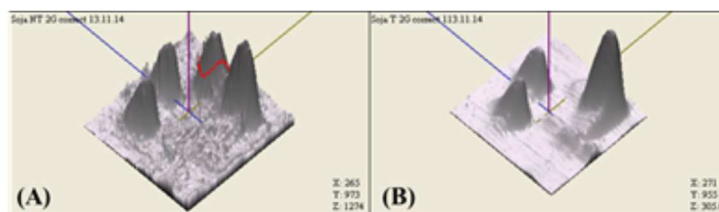
Available Online: 31 December 2015

ABSTRACT

Purpose: Evaluate possible differences in proteomic map between transgenic and non-transgenic soybean seeds varieties (RR 7511 and 8200, respectively) due to change in its genome by the insertion of *cp4EPSPS* gene, which confers to the soybeans tolerance to glyphosate.

Experimental description: A proteomic approach based on 2D-PAGE separation to identify alterations between T and NT profile was used. The soybean seeds proteins were extracted using a buffer (50 mmol L⁻¹ de Tris-HCl, 1,5 mmol L⁻¹ KCl, 10 mmol L⁻¹ DTT, 1,0 mmol L⁻¹ PMSF e 0,1 % SDS (m/v)) and its precipitation performed with ammonium acetate 0,1 mol L⁻¹. The precipitate was solubilized in a specific buffer and quantified with a 2D Quant Kit (according to the manufacturer's instructions). A total of 500 µg of protein, was applied to immobilized pH gradient strips (pH range 4–7) and they were rehydrated at room temperature for at least 12 h. Isoelectric focusing was performed for a total of 14,300 Vh. The second-dimension separation was carried out at 25 mA per gel and 100 W on 1-mm 12% (w/v) SDS polyacrylamide gels. After separation, the gels were stained with colloidal Coomassie brilliant blue. The gels were scanned using an ImageScanner II and the images processed by ImageMaster 2D Platinum 6.0, which allowed the estimation of the number of spots, the pI values and molecular mass of the proteins.

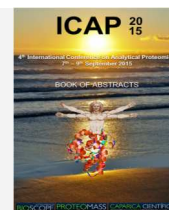
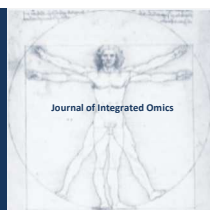
Results: The proteomic profile of T and NT seeds is different, thereby more than 20% of concentration of total protein was found in T seeds. Additionally, throughout the 3D evaluation, 14 different spots were found in the T and only 3 in NT seeds, as exemplified in Figure 1. This indicates that protein synthesis was different between T and NT soybeans. Intensities of the differentials spots were also observed. For a more accurate assessment, a quantitative analysis by 2D-DIGE will be performed to assess these changes. As future prospects, we will also quantify and identify these alterations in the proteomic profile and then propose a biochemical pathway.



Conclusions: The insertion of gene *cp4EPSPS* in soybean may be changing not only their genome but also their proteomic profile, thus most probably affecting future generations of plant. Regarding to the human food safety, no case has been reported however bioaccessibility tests need to be performed.

Keywords: Soybean, Organism Genetically Modified, Transgenic, Proteins, Proteomics, Electrophoresis.

Acknowledgements: The authors thank to FAPESP, CAPES and CNPq for financial supporting.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Comparative evaluation of extraction methods for UPLC-ESI-QTOF MS analysis of lipids from human blood serum

Leite Junior, J. G.¹, Arruda, M. A. Z.¹, Eberlin, M. N.², Sussulini, A.*¹

¹Spectrometry, Sample Preparation and Mechanization Group - GEPAM, National Institute of Science and Technology for Bioanalytics – INCTBio. Institute of Chemistry. Department of Analytical Chemistry. University of Campinas – UNICAMP. Campinas, SP, Brazil. ²Thomson Mass Spectrometry Laboratory. University of Campinas – UNICAMP. Campinas, SP, Brazil. *Corresponding author: sussulini@iqm.unicamp.br

Available Online: 31 December 2015

ABSTRACT

Purpose: The aim of this study was to perform a comparative evaluation of lipid extraction protocols from human blood serum using different chromatographic columns by UPLC-ESI-QTOF MS.

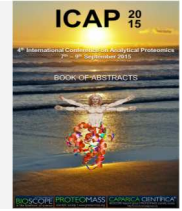
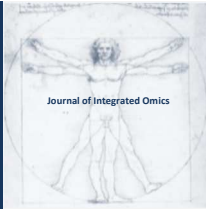
Experimental description: The evaluated extraction procedures included three liquid-liquid extraction techniques optimized for lipid profiling: i) modified Folch method, ii) Bligh and Dyer method, and iii) MTBE method. Modified Folch method consisted in adding 150 µL of methanol (MeOH) to 20 µL of serum sample and vortexing the mixture. Subsequently, 300 µL of chloroform (CHCl₃) were added and the mixture was incubated for one hour in a shaker before we induced phase separation by adding 125 µL of an aqueous solution of NaCl 0.15 mol L⁻¹. The mixture was centrifuged for 10 minutes at 8000 g at 10 °C and the lower (organic) phase was collected. For Bligh and Dyer extraction, 30 µL of serum were mixed with 360 µL of CHCl₃ and 180 µL of MeOH. The phase separation was achieved by adding 120 µL of water, the mixture was centrifuged for 10 minutes at 8000 g at 10 °C and the lower (organic) phase was collected. MTBE method was performed by adding 150 µL of MeOH to 20 µL of serum sample and vortexing the mixture. Subsequently, 500 µL of methyl tert-butyl ether (MTBE) was added and the mixture was incubated for one hour in a shaker before we induced phase separation by adding 125 µL of water. The mixture was centrifuged for 10 minutes at 8000 g at 10 °C and the upper (organic) phase was collected. All collected phase in each extraction was dried to complete dryness with liquid nitrogen, and then reconstituted in the proper mobile phase for lipid profiling analysis. Additionally, the chromatographic separation of the lipids was performed with three different columns (C8, C18, and HILIC), using UPLC-ESI-QTOF MS in positive and negative ionization mode. For all the analyses, a 1290 UPLC (Agilent) and a 6650 QTOF mass spectrometer (Agilent) were used.

Results: The results of this study show which extraction method is more efficient and which chromatographic column is more selective in the lipid profiling analysis of serum samples. It was considered four criteria to compare the extraction protocols: overall extraction coverage by comparing the number of identified peaks and the sum of the peak intensities, reproducibility of lipid quantifications, similarity of metabolite profile based on multivariate statistics and the specificity of extraction methods by comparing the peak abundances for each lipid class.

Conclusions: With the results obtained by lipid profiling analysis, we could compare the efficiency of each type of extraction using different chromatographic columns by UPLC-ESI-QTOF MS. The results from this preliminary study will be further applied in the search for bipolar disorder potential lipid biomarkers.

Keywords: Extraction methods, Lipid profiling, Serum and UPLC-MS.

Acknowledgements: The authors are grateful to FAEPEX for their financial support. The authors also thank Célio F. F. Angolini for helping with the mass spectrometry experiments.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Differential protein expression in the testes of ganders under different monochromatic light sources

C. Y. Cheng¹, Z. X. Zhuang¹, C. J. Chen^{2,3}, S. C. Chang^{1,4}, M. J. Lin^{1,4}, T. Y. Lin⁴, Y. S. Jea⁴, H. L. Chan^{5,6}, S. Y. Huang^{*1,7,8,9}

¹Department of Animal Science, National Chung Hsing University, Taichung 40227, Taiwan. ²Proteomics Core Laboratory, Department of Medical Research, China Medical University Hospital, Taichung 40402, Taiwan. ³Graduate Institutes of Integrated Medicine, China Medical University, Taichung 40402, Taiwan. ⁴Changhua Animal Propagation Station, Livestock Research Institute, Council of Agriculture, Taichung 52146, Taiwan. ⁵Institute of Bioinformatics and Structural Biology, National Tsing Hua University, Hsinchu 30013, Taiwan. ⁶Department of Medical Sciences, National Tsing Hua University, Hsinchu 30013, Taiwan. ⁷Agricultural Biotechnology Center, National Chung Hsing University, Taichung 40227, Taiwan. ⁸Center for the Integrative and Evolutionary Galliformes Genomics, iEGG Center, National Chung Hsing University, Taichung 40227, Taiwan. ⁹Center of Nanoscience and Nanotechnology, National Chung Hsing University, Taichung 40227, Taiwan. *Corresponding author: syhuang@dragon.nchu.edu.tw

Available Online: 31 December 2015

ABSTRACT

Purpose: In modern poultry production, patterns of artificial illumination treatment such as light schedule, intensity, and color are important factors that influence productive performances. The purpose of this study was to investigate whether different monochromatic light sources affect the protein expression in the testes of ganders.

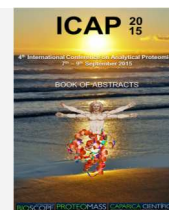
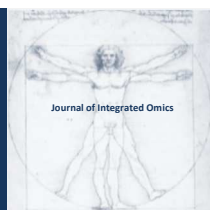
Experimental description: Six male White Roman ganders were used. The ganders were allocated into white or blue light groups. The lighting photoperiod of 7L:17D was applied for 6 weeks as an adjusting period. After the adjusting period, the photoperiod was changed to 9L:15D and maintained for 24 weeks. The ganders were sacrificed at the end of treatment to collect testis samples for proteomic analysis.

Results: The results of two-dimensional difference gel electrophoresis revealed that 37 protein spots were differentially expressed in the testes of ganders with different monochromatic light treatments. A total of 31 spots were identified by MALDI-TOF/TOF. Gene ontology analysis revealed that the differentially expressed proteins were mostly located in extracellular vesicular exosome (27%, 7 proteins), nucleus (11%, 3 proteins), and cytoplasm (11%, 3 proteins). Most of the proteins were with molecular function of protein binding (23%, 6 proteins), ion binding (23%, 6 proteins), carbohydrate binding (19%, 5 proteins), and heterocyclic compound binding (15%, 4 proteins). The majority of the proteins were participated in biological process of biological regulation (27%, 7 proteins) and cellular process (23%, 6 proteins). In addition, proteins related to sperm maturation and capacitation, including hemoglobin, apolipoprotein A-1, and phosphatidylethanolamine-binding protein 1, were downregulated after blue light treatment. The result suggested that blue light might disturb sperm functions and fertility on ganders.

Conclusions: A total of 37 protein spots were differentially expressed in the testes of ganders with blue and white light exposure. Proteins related to sperm maturation and capacitation were downregulated in the testes of blue light treated ganders. The exact role of the differentially expressed proteins needs further investigation.

Keywords: Testes, Monochromatic light sources, Protein expression, Ganders.

Acknowledgements: The authors would like to thank the Council of Agriculture (Contract No. 102AS-2.1.5-L1-L2) and Ministry of Education (under the ATU plan), Taiwan for financial support of this study.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Proteomic comparative study of smooth muscle cells isolated from systemic and pulmonary arteries

M. Žaloudíková¹, A. Eckhardt^{*2}, L. Kulhavá², R. Vytášek¹, K. Karmazín¹, K. Doušová¹

¹Department of Pathophysiology, 2nd Faculty of Medicine, Charles University in Prague, Prague, Czech Republic. ²Institute of Physiology, Academy of Sciences of the Czech Republic, Vídeňská 1083, Prague 4. **Corresponding author: eckhardt@biomed.cas.cz*

Available Online: 31 December 2015

ABSTRACT

Purpose: The cellular compositions of the systemic and pulmonary arterial walls are very similar. However the functional characteristics of these vessels show many differences. One of the most noticeable is in their reaction to hypoxia. We assume this difference can be consequence of the smooth muscle cells (SMC) protein composition of the peripheral arteries. Thus as we assume the definition of their proteomic differences can guide us to the important points with the potential to be a cause of their specific reactions.

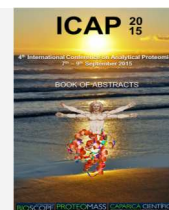
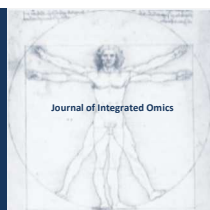
Experimental description: We used 14 adult male Wistar rats (200-250g). Animals lived in normoxia (n=9) or were exposed to isobaric hypoxia (FiO₂=10%) in hypoxic chamber for 4 days (n=5). The rats were euthanased by the intraperitoneal injection of thiopental. Peripheral pulmonary and mesenteric arteries were dissected under the microscopic control. Smooth muscle cells were obtained by the enzymatic digestion of the vascular fragments and the isolated material was subjected to sonication in lysis buffer. Protein mixture was analyzed by one-dimensional gel SDS-polyacrylamide electrophoresis followed by two-dimensional gel SDS-polyacrylamide electrophoresis and identified using nLC-MS/MS. Proteins were identified by correlating tandem mass spectra to IPI and SwissProt databases.

Results: We detected the significant differences in the protein composition between the samples separated from the pulmonary and systemic smooth muscle cells and also between the pulmonary smooth muscle isolated from the rats living in normoxia and exposed to 4 days hypoxia. We detected proteins more abundant in pulmonary SMC (vinculin, annexins, 14-3-3 proteins) and in systemic SMC (disulfide-isomerase, aldehyde dehydrogenase) and we observed the significant increase of collagen VI quantity in the normoxic pulmonary SMC in comparison with animals exposed to hypoxia.

Conclusions: We observed the significant differences of the proteomic profiles between pulmonary and systemic smooth muscle cells and also the significant changes in the normoxic pulmonary smooth muscle cells in comparison with animals exposed to hypoxia. These results bring new findings about extracellular matrix remodeling during hypoxia.

Keywords: smooth muscle cells, pulmonary, systemic artery, proteome, hypertension

Acknowledgements: This work was supported by Czech Science foundation (n. 15-01948S) and with support for long-term conceptual development of research organization RVO:67985823.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Protein expression profiles of serum samples bipolar disorder patients mapped by 2-D DIGE coupled with nanoLC-MS/MS

J.R. Jesus^{*1}, R. M. Galazzi¹, B. K. Campos¹, A. Sussulini¹, C. E. M. Banzatto², J. L. C. Martínez³, M. A. Z. Arruda¹

¹Spectrometry Sample Preparation and Mechanization Group – GEPAM, Institute of Chemistry, University of Campinas - UNICAMP, 13083-970, Campinas, SP, Brazil. ²Department of Psychiatry, Faculty of Medical Science, University of Campinas - UNICAMP, 13083-970, Campinas, SP, Brazil. ³Bioscope group, Chemistry Department, Faculty of Sciences and Technology, New University of Lisbon, Campus de Caparica, 2829-516, Caparica, Portugal. *Corresponding author: jemmyson.jesus@iqm.unicamp.br

Available Online: 31 December 2015

ABSTRACT

Purpose: Bipolar Disorder (BD), which affects up to 3% of the worldwide population, is a mental disease with unclear connections to other similar disorder. This study had aim application approach comparative proteomic to identify potential biomarkers in patient serum samples with bipolar disorder.

Experimental description: This study was approved by the Ethics Committee of the Hospital das Clinicas (University of Campinas, Brazil). Fifty six serum samples (23-57 years old) were collected and classified into five groups: (A) healthy family control (n= 3), (B) healthy no-family control (n= 9), (C) BD patients under treatment with lithium (Li) (n= 14), (D) schizophrenia patients (n= 25) and (E) other diagnostic patients under treatment with Li (n=4). The participants did not have other concomitant diseases such as cancer, endocrinological or metabolic diseases. Abundant proteins (IgG and albumin) depletion were performed using ProteoMiner spin columns. The analysis was performed using two-dimensional fluorescence difference-in-gel electrophoresis (2-D DIGE) coupled with nanoliquid chromatography-tandem mass spectrometry (nanoLC-MS/MS). For the analysis of 2-D DIGE, the samples were compared. As follows: (i) group A vs group B; (ii) group A vs group E; (iii) group B vs group E; (iv) group C vs group E; (v) group E vs group D.

Results: Approximately 50 µg [(pH range 4-70) determined from the 2-D Quant Kit] of each sample and the corresponding amount of the internal pooled standard were labeled with 400 pmol of CyDye DIGE Fluors minimal dyes. The ratios of protein abundance that increased or decreased more than 2.0 fold (t-test, p<0.05) were considered significant. Forty two protein spots were detected as significant difference among groups (41 increased and 1 decreased). In the experiments (i), (ii), (iii), (iv) and (v), were identified 4 protein spots (3 with higher abundance and 1 with lower abundance for B group), 8 (all protein spots with higher abundance for C group), 13 (all protein spots with higher abundance for C group), 9 (all protein spots with higher abundance for D group) and 8 (all protein spots with higher abundance for C group) protein spots presenting significant difference, respectively. All proteins spots presenting significant difference were selected for in-gel trypsin digest and nanoLC-MS/MS sequencing for protein identification. In the next step, the identification of proteins by nanoLC-MS/MS will be made.

Conclusions: In this study, we found that close forty protein spots shown significant differences among the groups compared using 2-D DIGE. As perspective, the proteins will be identified by nanoLC-MS/MS and some proteins will be selected to confirm the comparative proteomic data using immunoassay.

Keywords: Bipolar disorder, comparative proteomic, blood biomarker, 2-D DIGE, nanoLC-MS/MS.

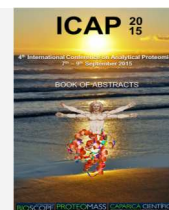
Acknowledgements: FAPESP, CAPES, CNPq, FAEPEX.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Western blotting - detection of a specific protein in MCF7 and MCF7-Z tissue

J. Ner-Kluza^{*1}, A. Drabik¹, Michael Kubbutat², Andreas Lingnau², P. Suder^{1,5}, G. Schroeder⁴, J. Silberring^{1,3}

¹Department of Biochemistry and Neurobiology, Faculty of Materials Sciences and Ceramics, AGH University of Science and Technology, Mickiewicza 30 Ave. 3 0-059 Krakow, Poland. ²ProQinase GmbH, Freiburg, Germany. ³Center of Polymer and Carbon Materials, Polish Academy of Sciences, Marii Skłodowskiej-Curie 34., 41-819 Zabrze, Poland. ⁴Department of Supramolecular Chemistry, Faculty of Chemistry, Adam Mickiewicz University, Umultowska 89b, 61-614 Poznan, Poland. ⁵AGH University of Science and Technology, Academic Centre for Materials and Nanotechnology, Mickiewicza 30, 30-059, Kraków, Poland. *Corresponding author: nerjoanna@gmail.com

Available Online: 31 December 2015

ABSTRACT

Purpose: In the contemporary world, breast cancer is one of the most common and dangerous types of cancer in females. Comparison between two different variants of MCF-7 (MCF-7 vs MCF-7-Z) human breast cancer cell lines used in subcutaneous xenograft tumor models, has shown different tumor growth characteristics in mice. Glycosylated proteins are among post-translational modifications one of the most important types of proteins involved in tumor progression. The aim of the research was to search for the presence of selected glycoproteins, based on quantitative and qualitative analysis in cancer progression.

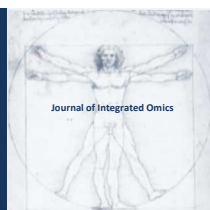
Experimental description: For samples preparation we used the strategy utilizing western blotting and nanoLC-MS/MS analysis. After isolation of proteins from the tissue, glycoproteins were isolated by lectin affinity chromatography. Glycoproteins fractions were separated by the sodium dodecyl sulfate polyacrylamide gel (SDS-PAGE), followed by two complementary analyses. One part of the gel was transferred onto PVDF membrane, and another part was CBB stained. Based on the images obtained after immunostaining, the bands were excised from the CBB-stained gel and analyzed by tandem mass spectrometry coupled to liquid chromatography (LC-MS/MS).

Results: Using LC-MS/MS technique we have succeeded to identify 3000 differentially expressed proteins between MCF7 and MCF7-Z. The selected proteins were associated with cancer metastasis, which plays a vital role in tumor progression, and are involved in cell structure organization. The presence of the selected proteins was confirmed using western blotting.

Conclusions: We have identified the potential biomarkers, which can be important in cancer progression.

Keywords: cell lines MCF-7, glycoproteins, tumor, breast cancer.

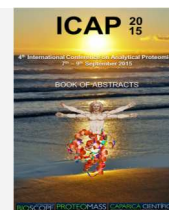
Acknowledgements: The research was supported by the grant EuroNanoMed "META" 05/EuroNanoMed/2012.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

The quantitative proteomic study of human saliva samples obtained from caries-free and caries-susceptible people

L. Kulhavá^{*1,2}, A. Eckhardt¹, M. Jäger¹, S. Pataridis¹, I. Mikšík¹

¹Institute of Physiology of the Czech Academy of Sciences, Videnska 1083, 14220, Prague 4, Czech Republic. ²Department of Analytical Chemistry, Faculty of Science, Charles University in Prague, Hlavova 8, 12843 Prague 2, Czech Republic. **Corresponding author: lucie.kulhava@fgu.cas.cz*

Available Online: 31 December 2015

ABSTRACT

Purpose: The human teeth have been together with the saliva in the center of interest for many years, due to medical problems with them [1]. Only a minor part of the population (ca 10 % of people in the age at ca 30 years) is resistant to tooth caries. In saliva there were detected over 2000 proteins to these days, and important part of them are salivary defense proteins. Our study was aimed to proteomics of saliva and to find the differences in the abundances of the responsible proteins in these tissues between caries-resistant and caries-susceptible people..

Experimental description: The human saliva samples from adult patients (aged 17-35) were collected and divided into two groups with respect to the number of dental caries (caries-susceptible vs. caries-resistant people). The proteins of oral fluids were separated by two-dimensional electrophoresis or by difference gel electrophoresis and the resulting protein maps were quantitatively evaluated. Spots exhibiting statistically significant changes were excised, digested by trypsin, and analyzed by nano-liquid chromatography coupled to a MaXis Q-TOF mass spectrometer with high resolution.

Results: We detected more than several hundreds of proteins in each kind of sample (pulp, dentin, saliva) [2]. Then we compared the individual proteome maps of saliva samples by 2-DE and DIGE and identified the changes in the protein abundances among the individuals. The observed significant differences between caries-resistant and caries-susceptible groups. Special emphasis was put on looking for the abundance changes of proteins that may be involved in formation of dental caries due to their biological functions.

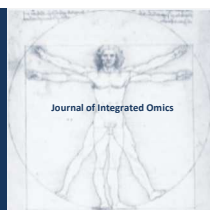
Conclusion: This study comparing proteomes of whole saliva from carious-resistant and carious-susceptible people brings new findings to the saliva protection against tooth caries.

Keywords: human saliva, proteome, two-dimensional gel electrophoresis, DIGE.

Acknowledgements: This work was supported by the Ministry of Health of the Czech Republic Departmental Program for Research and Development (NT14324-3/2013), by the Czech Science Foundation (No. 13-172224S), and with support for long-term conceptual development of research organization RVO:67985823.

References:

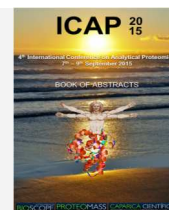
- [1] Jäger M., Eckhardt A., Pataridis S., Broukal Z., Dušková J., Mikšík I., *Physiol Res.* (2014) 63: 141-154.
- [2] Eckhardt A., Jäger M., Pataridis S., Mikšík I., *J. Endod.* (2014) 40(12):1961-1966.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

The quantitative comparison of the human tooth pulp obtained from caries-free and caries-susceptible people

M. Jäger*, A. Eckhardt, S. Pataridis, L. Kulhavá, I. Mikšík

Institute of Physiology of the Czech Academy of Sciences, Videnska 1083, 14220, Prague 4, Czech Republic *Corresponding author: jagr@biomed.cas.cz

Available Online: 31 December 2015

ABSTRACT

Purpose: The majority of the human population worldwide experiences dental caries. Only a minor part of the population (ca. 10 % of people in the age at 30 years) is resistant to development of this oral disorder. The anti-caries resistance can be caused by overall immune system, nourishment, dental care efficiency and/or by many other causes, such as composition of saliva or teeth. Differences in the abundances of the responsible proteins in these tissues between caries-resistant and caries-susceptible people are expected and are in the center of our interest. The aim of our work was to investigate the proteomic profile of human tooth pulp from adult people and compare them mutually.

Experimental description: Sound human teeth extracted for clinical reasons from adult patients (aged 17-40) were divided into two groups with respect to the number of dental caries (caries-susceptible vs. caries-resistant people). The proteins from individual tooth pulp samples were extracted and processed by the method described previously [1]. The proteins were separated either by two-dimensional electrophoresis followed by Coomassie colloidal blue staining or by difference gel electrophoresis and the resulting protein maps were quantitatively evaluated. Spots exhibiting statistically significant changes were excised, digested by trypsin, and analyzed by nano-liquid chromatography coupled to a MaXis Q-TOF mass spectrometer with ultrahigh resolution.

Results: We detected more than several hundreds of spots in the proteome maps of each human tooth pulp sample. Individual proteome maps were compared by both methods (2-DE and DIGE) and changes in the protein abundances were identified among the individuals. The observed differences were statistically evaluated to find the significant changes. Some of these spots corresponded e.g. to peroxiredoxin 1, GTP-binding nuclear protein Ran, apolipoprotein A-I, glutathione S-transferase P, phosphoglycerate kinase 1 or retinal dehydrogenase. These proteins exhibit variety of biological functions: cell communication and signal transduction, transport or metabolism.

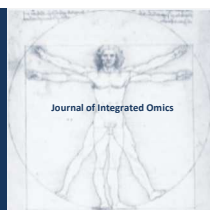
Conclusions: We determined some quantitative differences in the whole proteomes of human teeth pulp samples between carious-resistant and carious-susceptible people. Special emphasis will be engaged in further investigations of these proteins that may be involved in formation of dental caries due to their biological functions.

Keywords: human tooth pulp, proteome, two-dimensional gel electrophoresis.

Acknowledgements: This work was supported by the Ministry of Health Departmental Program for Research and Development (NT14324-3/2013), and with support for long-term conceptual development of research organization RVO:67985823.

References:

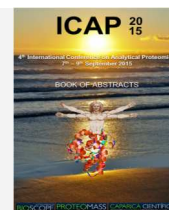
[1] A. Eckhardt, M. Jäger, S. Pataridis, I. Mikšík, J. Endodont. 40 (2014) 1961-1966.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

In vivo determination of the CYP4A and COX5A expression and catalytic activity in rat hepatic microsomes after drug administration

P. Mielczarek^{*1}, M. Smoluch², J. H. Kotlinska³, P. Suder^{1,2}, G. Schroeder⁴, J. Silberring^{2,5}

¹AGH University of Science and Technology. Academic Centre for Materials and Nanotechnology. Mickiewicza 30. 30-059 Krakow. Poland.

²AGH University of Science and Technology. Faculty of Materials Science and Ceramics. Department of Biochemistry and Neurobiology. Mickiewicza 30. 30-059 Krakow. Poland. ³Department of Pharmacology and Pharmacodynamics. Medical University. Chodzki 4a. 20-093 Lublin. Poland. ⁴Department of Supramolecular Chemistry. Faculty of Chemistry. Adam Mickiewicz University. Umultowska 89b. 61-614 Poznan. Poland. ⁵Centre of Polymer and Carbon Materials. Polish Academy of Sciences. Curie-Skłodowskiej 34. 41-819 Zabrze. Poland.

*Corresponding author: przemyslaw.mielczarek@agh.edu.pl

Available Online: 31 December 2015

ABSTRACT

Purpose: Drug dependence influences cell functions and their biological pathways, but the processes do not seem to be clear and understood. Global analysis of liver proteome can contribute to the explanation of drugs' influence on the metabolic processes and may help to explain the whole problem. The rat liver microsomes were taken from control and drugs-addicted rats, and up- and down regulated proteins were evaluated.

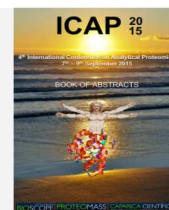
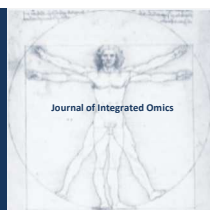
Experimental description: Adult male Wistar rats were treated s.c. with either saline (control) or amphetamine, morphine, or cocaine in saline, for 15 days. Liver microsomes were prepared according to a standard protocol. The proteins were divided into up- and down regulated groups, based on LC-MSⁿ analyses, followed by the Mascot server (Matrix Science) identification, after SDS-PAGE separation. The expression of the selected proteins was determined by Western Blotting and the catalytic activity of cytochromes P450 was also determined.

Results: Two proteins were chosen from up- and down regulated proteins identified by LC-MSⁿ for Western Blotting, cytochrome P450 4A10 (CYP4A) and cytochrome c oxidase subunit Va (COX5A). The enzymatic activity of CYP4A was determined by LC-MSⁿ determination of ω-hydroxylation of lauric acid.

Conclusions: After administration of various drugs (amphetamine, morphine, and cocaine) to the rats, significant changes in liver proteome were observed, also including cytochromes P450 responsible for oxidative metabolism. The expression and catalytic activity of CYP4A and COX5A was determined.

Keywords: cytochrome P450, CYP4A, COX5A, microsomes, drug dependence.

Acknowledgements: The authors acknowledge support for this work by the grant EuroNanoMed "META", 05/EuroNanoMed/2012 and the Polish National Science Center's grant, 2012/07/B/NZ4/01468.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Label free proteomics from rat liver microsomes in drug dependence

M. Smoluch^{*1}, P. Mielczarek², G. Schroeder³, P. Suder^{1,2}, J. Silberring^{1,4}

¹AGH University of Science and Technology. Faculty of Materials Science and Ceramics. Department of Biochemistry and Neurobiology. Mickiewicza 30. 30-059 Krakow. Poland. ²AGH University of Science and Technology. Academic Centre for Materials and Nanotechnology. Mickiewicza 30. 30-059 Krakow. Poland. ³Department of Supramolecular Chemistry. Faculty of Chemistry. Adam Mickiewicz University. Umultowska 89b. 61-614 Poznan. Poland. ⁴Centre of Polymer and Carbon Materials. Polish Academy of Sciences. Curie-Skłodowskiej 34. 41-819 Zabrze. Poland. *Corresponding author: marek.smoluch@agh.edu.pl

Available Online: 31 December 2015

ABSTRACT

Purpose: The drug dependence has a significant impact on the cells' functions and biological processes in mammalian body. In many cases, the mechanism of drug dependence is not fully understood and the knowledge on its influence on the human body is not complete. The presented work shows application of the label free methodology in proteome analysis in drug dependence.

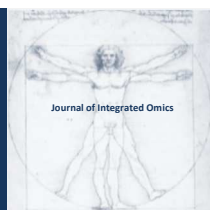
Experimental description: Microsomes were prepared from the Wistar rats after administration of a drug for 15 days. Four groups of animals were analyzed: amphetamine-, morphine-, cocaine-dependent, and control. Proteins were separated by SDS-PAGE and the molecules within the range of 46-58 kDa were trypsin digested and analyzed by LC-MSⁿ. Peptide maps were separated by the reversed phase capillary system, and detected by an Amazon ETD mass spectrometer (Bruker Daltonics, Bremen, Germany). Data were processed by the Profile Analysis (label free approach) and Proteinscape softwares from Bruker Daltonics. Finally, the GO analysis for up-, and down regulated proteins were performed using the PANTHER Gene List Analysis.

Results: Identification of peptides was performed on the basis of CID fragmentation. MS data allowed for proteins identification by the Mascot server (Matrix Science). After label free analyses, the proteins identified in rat microsomes (control and drug dependent) were divided into up-, and down regulated groups. The GO Biological Functions and GO Biological Processes for up and down regulated proteins were defined.

Conclusions: The project presents application of the label free proteomic research in drug addiction with the focus on cytochromes P450 (CYP450), mainly responsible for the metabolic processes in liver. Also, the results show comparison of an influence of different drugs (amphetamine, morphine and cocaine) on proteins present in liver microsomes.

Keywords: label free proteomics, microsomes, cytochromes P450, drug dependence

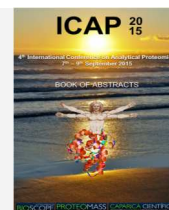
Acknowledgements: The authors acknowledge support for this work by the grant EuroNanoMed "META", number 05/EuroNanoMed/2012.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

[HTTP://WWW.JIOMICS.COM](http://www.jiomics.com)



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Sodium effect on agonist binding to δ -opioid receptors in isolated plasma membranes from forebrain cortex of rats exposed to increasing doses of morphine (10 - 50 mg/kg) for 10 days

M. Vošahlíková*, H. Ujčíková, P. Svoboda

Laboratory of Biochemistry of Membrane Receptors, Department of Biomathematics, Institute of Physiology of the Czech Academy of Sciences, Videnska 1083, 14220 Prague 4, Czech Republic. *Corresponding author: vosahlikova@fgu.cas.cz

Available Online: 31 December 2015

ABSTRACT

Purpose: The aim of this work was to determine the effect of sodium cations, representing the specific allosteric modulators of ligand binding site of opioid receptors (OR), on agonist binding to δ -opioid receptors (δ -OR) in plasma membranes isolated from forebrain cortex of rats exposed to morphine for prolonged period of time.

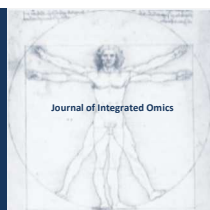
Experimental description: Rats were exposed to increasing doses of morphine (10 - 50 mg/kg) for 10 days and sacrificed 24 hours (group +M10) or 20 days (group +M10/-M20) after the last dose. Control animals were sacrificed in parallel with morphine-treated (groups -M10 and (-M10/-M20)). Plasma membrane enriched fraction (PM) was isolated from rat brain cortex (+M10 and -M10) and analyzed by specific radioligand binding. δ -OR were characterized by saturation binding studies with δ -OR agonists [³H]DADLE and [³H]DPDPE in the presence or absence of 100 mM NaCl. Percoll-purified PM (from all groups) were also resolved by 1D-SDS-PAGE and 2D-ELFO. Subsequently, immunoblot detection with specific antibodies and proteomic analysis was performed.

Results: Maximum number of [³H]DADLE binding sites in PM isolated from morphine-treated rats was 1.4-fold higher than in PM isolated from control rats. Surprisingly, 100 mM sodium chloride had no effect on [³H]DADLE binding in PM prepared from morphine-treated animals (+M10), but, as expected, it did inhibit agonist binding to PM isolated from control animals (-M10). Morphine-induced increase of ligand binding to δ -OR was substantially higher when more specific ligand, [³H]DPDPE, was used for determination of δ -OR in brain membranes. Maximum number of [³H]DPDPE binding sites in morphine-treated membranes was 2.1-fold higher than in control membranes. Thus, the inhibition of agonist binding by sodium was detected in control membranes only. Our results indicated a specific up-regulation of adenylyl cyclases I (8-fold) and II (2.5-fold) and significant increase of proteins functionally related to oxidative stress and apoptosis.

Conclusions: Sodium ions have been described as efficient inhibitors of agonist binding to numerous GPCR including OR, causing the shift of receptor molecules from active (R*) to inactive (R) conformation and the uncoupling of receptor from the cognate G protein, i.e. inverse agonist effect. The low sensitivity of δ -OR to inhibitory effect of sodium in morphine-treated samples of forebrain cortex may be interpreted as disturbance of equilibrium between the active and non-active forms of δ -OR molecules.

Keywords: sodium, morphine, rat forebrain cortex, opioid receptors.

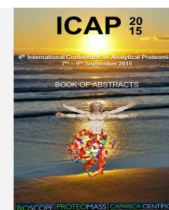
Acknowledgements: This work was supported by the GACR (P207/12/0919, P304/12/G069) and by the Czech Academy of Sciences (RVO: 67985823).



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Do or do not. There is no try

S. Jorge^{*1,2}, J.E. Araújo^{1,2}, H.M. Santos^{1,2}, C. Lodeiro^{1,2}, J.L. Capelo^{1,2}

¹UCIBIO-REQUIMTE, Department of Chemistry, Faculty of Sciences and Technology, University NOVA of Lisbon, 2829-516 Caparica, Portugal. ²PROTEOMASS Scientific Society, Madan Parque, Rua dos Inventores, 2825-182 Caparica, Portugal. **Corresponding author: s.jorge@campus.fct.unl.pt*

Available Online: 31 December 2015

ABSTRACT

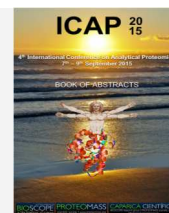
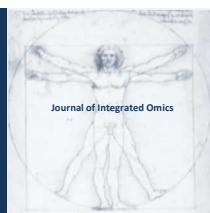
Protein biomarker discovery for medical diagnostic or for pharmacological purposes is an issue of primary importance for the proteomic community. In proteomics, the traditional sample treatments for protein identification through trypsination are tedious and time-consuming. Nowadays, ultrasound energy is gaining momentum to speed up sample digestion as well as different steps of the sample treatment for protein identification. Thus, gel washing, enzymatic digestion and peptide extraction from gels are speeded using USE. However, there is still a lack in the number of samples that can be treated at once^{1,2,3,4}. Herein, we propose the use of a new ultrasonic technology, the Microplate Horn Assembly, to reduce the complexity of classical protocols to identify proteins as well as to increase the number of samples treated a once to an unprecedented level.

Keywords: Proteomics, protein identification, ultrasound energy, protein biomarker discovery.

Acknowledgements: H.M.S. acknowledge the post-doctoral grants from Fundação para a Ciência e a Tecnologia (FCT-MEC) (Portugal) SFRH/BPD/75242/2010. S.J. thanks a research grant from Nova Health-UNL (Portugal) and PROTEOMASS Scientific Society. Financial support from the Scientific PROTEOMASS Association (Portugal) and UCIBIO/REQUIMTE (UID/Multi/04378/2013) are acknowledged.

References:

- [1] J.E. Araújo, E. Oliveira, E. P. Kouvonen, et al. Talanta.2014, 121, 71-80.
- [2] J.D. Nunes-Miranda, C. Núñez, H.M. Santos, et al. Analyst, 2014, 139, 992-995.
- [3] C. Fernández-Costa, C. Ruiz-Romero, F.J. Blanco, H.M. Santos, J.L. Capelo, Talanta, 2013, 106, 163-168.
- [4] G. Vale, H.M. Santos, R.J. Carreira et al. Proteomics, 2011, 11, 3866-3876.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Proteomics analysis of the peritoneal dialysis effluent: a longitudinal study

J. E. Araújo^{*1}, S. Jorge¹, F. Teixeira-Costa², A. Ramos², H. M. Santos¹, J. L. Capelo¹, C. Lodeiro¹

¹UCIBIO, REQUIMTE, Departamento de Química, Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa, 2829-516 Caparica, Portugal. ²Serviço de Nefrologia, Hospital Garcia da Orta, Almada, Portugal. *Corresponding author: jeduardoaraujo88@gmail.com

Available Online: 31 December 2015

ABSTRACT

Purpose: Long-term peritoneal dialysis (PD) leads to morphological and functional alterations in the peritoneum, reducing the lifespan of this dialysis up to five years, and forcing the replacement of PD by other renal replacement therapies. Peritoneum failure does not occur in every patient in the same sequence and to the same extent. This work aims to develop longitudinal studies to unravel the evolution of the proteome of the peritoneal dialysate with time, so biomarkers and molecular profiles for diagnosis and prognosis can be obtained.

Experimental description/ Results: Peritoneal dialysis effluent (PDE) samples from anonymous patients, already at different stages of dialysis were taken. The PDE from the patients was analyzed using 2D gel electrophoresis¹. Gel comparison and statistical analysis using Progenesis SameSpots indicated the gel spots differentially expressed for each patient (P01 (6 proteins overexpressed (↑), 3 underexpressed (↓)); P02 (7(↑), 2(↓)); P03 (12 (↑), 19 (↓)); P04 (4 proteins (↓)); P05 (5 proteins (↑), 2 (↓)). All the detected spots were excised digested and identified by MALDI-MS.

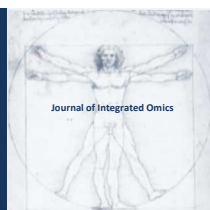
Conclusion: All the identified proteins were studied for their biochemical function. Identification of specific molecular changes can be particularly interesting for the understanding and early detection of long-term peritoneum alterations as well as for the development of new therapies to increase the lifespan of the peritoneal dialysis.

Keywords: Peritoneal dialysis, Peritoneal dialysate effluent, MALDI-TOF-MS, 2D-SDS-PAGE, long-term PD.

Acknowledgements: This work was supported by PROTEOMASS Scientific Society (Portugal). Authors thank LAQV/REQUIMTE (UID/QUI/50006/2013) and UCIBIO/REQUIMTE (UID/Multi/04378/2013) for general funding.

Reference:

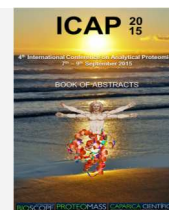
Elisabete Oliveira et al. Clinical Proteomics, 2014, 11:17.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Differential expression of egg white proteins in different breeds of chickens and their association with physical characteristics of hard-boiled eggs

S. Y. Huang^{*1,2,3,4}, S. C. Liu¹, C. Y. Wu⁵, C. J. Chen^{6,7}, Y. P. Lee¹, C. F. Chen^{1,2,3}, J. C. Peng⁵, F. J. Tan¹

¹Department of Animal Science, National Chung Hsing University, Taichung 40227, Taiwan. ²Agricultural Biotechnology Center, National Chung Hsing University, Taichung 40227, Taiwan. ³Center for the Integrative and Evolutionary Galliformes Genomics, iEGG Center, National Chung Hsing University, Taichung 40227, Taiwan. ⁴Center of Nanoscience and Nanotechnology, National Chung Hsing University, Taichung 40227, Taiwan. ⁵Department of Bio-industrial Mechatronics Engineering, National Chung Hsing University, Taichung 40227, Taiwan. ⁶Proteomics Core Laboratory, Department of Medical Research, China Medical University Hospital, Taichung 40402, Taiwan. ⁷Graduate Institute of Integrated Medicine, China Medical University, Taichung 40402, Taiwan. *Corresponding author: syhuang@dragon.nchu.edu.tw

Available Online: 31 December 2015

ABSTRACT

Purpose: The purpose of this study was to explore the differential expression of egg white proteins in different breeds of chickens and their association with physical characteristics of hard-boiled eggs.

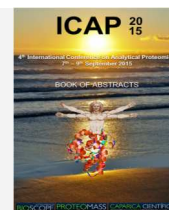
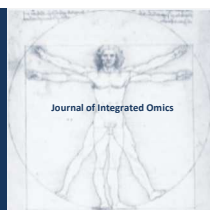
Experimental description: Fresh eggs collected from 31-wk-old ISA Brown, L2 strain Taiwan country chickens (L2 strain TCCs), and Quemoy chickens, were stored at room temperature (25-26°C) for 3 days before further analysis. The eggs were subjected to measuring general traits and physical characteristics of hard-boiled eggs evaluation, including texture profile analysis, stress relaxation, breaking test, and puncture test. The egg white were used for proteomic analysis.

Results: The general egg traits showed that eggs from ISA Brown had higher egg weight, egg length, and egg width than those of L2 strain TCCs and Quemoy chickens ($P < 0.05$). The egg shape index, yolk weight, albumin height, and Haugh unit did not differ among breeds. The physical characteristics of hard-boiled eggs in L2 strain TCCs showed higher hardness, fracturability, gumminess, chewiness, puncture value, viscoelasticity, and toughness than those of the other two breeds ($P < 0.05$). The levels of 16 out of 124 quantified protein spots differed significantly among the three breeds ($P < 0.05$). Fifteen of the differed protein spots were successfully identified by peptide mass fingerprinting and represented 7 distinct proteins, including Hep21 protein precursor, ovoinhibitor, ovalbumin related protein X, ovalbumin, ovotransferrin, serum albumin, and phosphoglucosyltransferase-1. The levels of one and three ovotransferrin spots in L2 strain TCCs were significantly higher than those in ISA Brown and Quemoy chickens. Pearson correlation further showed that the level of ovotransferrin was highly positive-correlated with most of the physical characteristics of hard-boiled eggs.

Conclusions: The result of this study suggested that the reason why the eggs of L2 strain TCCs have better physical characteristics of hard-boiled eggs may result from the higher level of ovotransferrin in their egg white.

Keywords: Chicken eggs, Egg white proteins, Physical characteristics, Hard-boiled eggs.

Acknowledgements: The authors would like to thank the Ministry of Education, Taiwan (under the ATU plan) for financial support of this study.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Methods for the preparation of acute myeloid leukemia patient samples for proteomic and phosphoproteomic analysis

M. Hernandez-Valladares^{*1}, E. Aasebø¹, F. Berven¹, Ø. Bruserud², F. Selheim¹

¹PROBE, Building for Basic Biology, University of Bergen, Jonas Lies vei 91, N-5009 Bergen, Norway. ²Department of Internal Medicine, Haukeland University Hospital, N-5020, Bergen, Norway. **Corresponding author: Maria.Hernandez-Valladares@uib.no*

Available Online: 31 December 2015

ABSTRACT

Purpose: The study of global protein expression in acute myeloid leukemia (AML) patients by mass spectrometry (MS) can help identifying differential expression and post-translational modifications of proteins that could represent disease-related biomarkers for early diagnosis or for improved prognostics and to predict the patient response to different therapeutics. To optimize the proteome and phosphoproteome coverage of samples from AML patients by LC-MS/MS analysis, we have tested several methods with different peptide fractionation and phosphopeptide enrichment strategies.

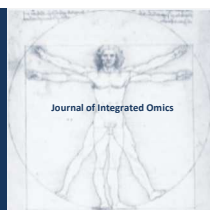
Experimental description: Peptide samples were prepared with an in-solution digestion protocol, using urea or guanidinium hydrochloride as denaturant, and with the filter aided sample preparation (FASP) methodology. Peptide fractionation was carried out with reverse phase/strong cation exchange (SCX) and SCX disks in a stage-tip. Techniques for phosphopeptide enrichment included metal oxide affinity chromatography (MOAC), immobilized metal affinity chromatography (IMAC) and sequential elution from IMAC (SIMAC).

Results: From an initial screening of the different samples on a Linear Trap Quadrupole (LTQ) Orbitrap Elite MS, we found the best proteome coverage with the sequential FASP method, with Lys-C and trypsin as digestion enzymes, which identified and quantified 3100 proteins from 20 µg of sample. With the same strategy, followed by a separate MOAC/TiO₂-beads enrichment of the two peptide pools, we identified and quantified 2.900 phosphorylation sites from only 250 µg of AML patient sample. On a QExactive HF hybrid Quadrupole-Orbitrap MS, 5400 proteins and 4000 phosphosites were identified and quantified from these FASP-prepared samples.

Conclusions: To improve the proteome and phosphoproteome coverage of AML patient samples by MS analysis to discover new biomarkers, testing of different methods can be beneficial. Based on sample preparation optimization in our laboratory, we have chosen the FASP protocol to prepare AML patient samples for MS-based proteomic and phosphoproteomic studies.

Keywords: AML, proteomics, phosphoproteomics, sample preparation.

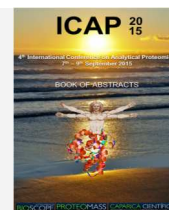
Acknowledgements: The authors would like to thank the Norwegian Cancer Society and Øyvinn Mølbach-Petersens Fond for the funding of this work.



JOURNAL OF INTEGRATED OMICS

A METHODOLOGICAL JOURNAL

HTTP://WWW.JIOMICS.COM



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Proteomics analysis of the peritoneal dialysate effluent reveals the presence of calcium-regulation proteins and acute inflammatory response

E. Oliveira^{1,2}, J.E. Araújo^{1,2}, S. Gómez-Meire³, C. Lodeiro^{1,2}, C. Perez-Melon⁴, E. Iglesias-Lamas⁴, A. Otero-Glez⁴, J.L. Capelo^{1,2} and Hugo M Santos^{*1,2}

¹BIOSCOPE Research Group. UCIBIO-REQUIMTE, Departamento de Química, Faculdade de Ciências e Tecnologia, Universidade NOVA de Lisboa, 2829-516 Caparica, Portugal. ²PROTEOMASS Scientific Society, Madan Parque, Rua dos Inventores, 2825-182 Caparica.. ³SING Group. Informatics Department. Higher Technical School of Computer Engineering, University of Vigo, Ourense, Spain. ⁴Servicio de Nefrología, Complejo Hospitalario Universitario de Ourense, 32004 Ourense, España. *Corresponding author: hmsantos@fct.unl.pt

Available Online: 31 December 2015

ABSTRACT

Purpose: Peritoneal dialysis (PD) is a form of renal replacement used for advanced chronic kidney disease. PD effluent holds a great potential for biomarker discovery for diagnosis and prognosis. In this study a novel approach to unraveling the proteome of PD effluent based on dithiothreitol depletion followed by 2D-SDS-PAGE and protein identification using tandem mass spectrometry is proposed.

Experimental description: In the present study, samples of peritoneal dialysate effluent were collected from six patients from the Complejo Hospitalario Universitario de Ourense (CHUO). Before proteomics analysis the six samples of peritoneal fluid were depleted with DTT and quantified using a Bradford protein assay. In order to analyze the differences among the patients, 2D gels were carried out by triplicate for each patient. A total of 100 µg of protein was loaded onto pH 3–10 strips, and then proteins were visualized with CBB. Gel spots were excised and subjected to in-gel digestion and MALDI-TOF/TOF analysis.

Results: A total of 49 spots were analyzed revealing 25 proteins differentially expressed, among them many proteins involved in calcium regulation.

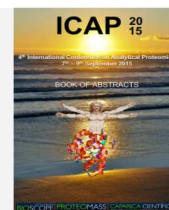
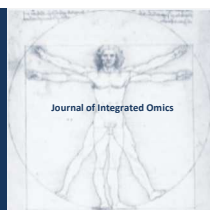
Conclusions: Through this work it has been demonstrated that the proteome of the peritoneal dialysis effluent is far from being well established. A total of 25 proteins differentially expressed have been identified among 6 patients with PD. It is concluded that studies dealing with PDE are difficult to compare if the methodology used to treat the samples and to identify the proteins are not the same. It has been shown that the DTT-based depletion method is a cheap alternative to be considered to other expensive approaches. Remarkably, a group of proteins dealing with calcium metabolism and calcium regulation has been found to be lost through peritoneal dialysate effluent. This finding might be directly linked to the calcification of soft tissues, in patients subjected to PD, and opens new insights into the potential use of PDE as a sample for diagnosis and prognosis of patients with renal failure.

Keywords: Peritoneal dialysis effluent, 2D-Gel Electrophoresis, Protein identification, Proteomics.

Acknowledgements: E. Oliveira and H. M. Santos acknowledge the post-doctoral grants SFRH/ BPD/72557/2010 and SRFH/ BPD/73997/2010 respectively, provided by Fundação para a Ciência e a Tecnologia–Ministério da Educação e Ciência (FCT-MEC, Portugal). J. E. Araújo acknowledges the grant from Fundación Renal Íñigo Álvarez de Toledo (Spain). This work was supported in part by Xunta de Galicia (Spain) under projects 09CSA043383PR and 10CSA383009PR and by PROTEOMASS Scientific Society (Portugal). Authors thanks to REQUIMTE-FCT PEst-C/EQB/LA0006/2013.

Reference:

Oliveira *et al.* Clinical Proteomics, 2014, 11:17.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Evaluation of alternative strategies depletion proteins for biomarker discovery in patients serum with bipolar disorder

J.R. Jesus^{*1}, G. S. Pessôa¹, T. B. Lima², F.C. Gozzo², J. L. C. Martínez³, M. A. Z. Arruda¹

¹Spectrometry Sample Preparation and Mechanization Group – GEPAM. Institute of Chemistry. University of Campinas - UNICAMP. 13083-970. Campinas, SP. Brazil. ²Dalton Mass Spectrometry Group. Institute of Chemistry. University of Campinas - UNICAMP. 13083-970. Campinas, SP. Brazil. ³Bioscope group. Chemistry Department. Faculty of Sciences and Technology. New University of Lisbon. Campus de Caparica. 2829-516. Caparica. Portugal. *Corresponding author: jemmyson.jesus@iqm.unicamp.br

Available Online: 31 December 2015

ABSTRACT

Purpose: Bipolar Disorder (BD) is a mental disease with unclear connections to other similar disorder that affect hundreds of thousand of individuals in the world. Serum may be utilized as a rich source of biomarker of disease. However, the large number of proteins in serum makes the analysis very difficult because high abundant proteins (albumin and Immunoglobulin) tend to mask of those of lower abundance. Depletion of abundant serum proteins may help in the discovery and detection proteins with potential biomarker of diseases, such as bipolar disorder. Thus, this study had aim evaluate alternative procedure depletion abundant proteins to biomarkers discovery in patient serum samples with bipolar disorder.

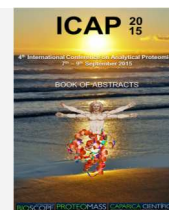
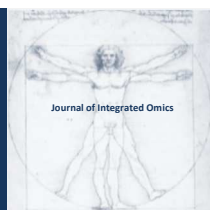
Experimental description: A variety of depletion methods for removal of high abundant proteins from serum have been related in the literature. This work shows the potential of chemical protein depletion, using dithiothreitol (DTT), acetonitrile (ACN), and tandem depletion (DTT/ACN). For protein depletion with DTT, approximately 2.0 µL of DTT (500mM) was added to 20 µL (> 10µg µL⁻¹) of serum human. Following, the mixture was incubated for 60 min at 37 °C until a white precipitate come to light, which is pellet by centrifugation at 13,000 rpm for 40 min. The supernatant was transferred into microtubes and stored at -20 °C until analysis by SDS-PAGE. For protein depletion with ACN, 20 µL (> 10µg µL⁻¹) of serum human was diluted to about 60 µL with Milli-Q water. Then, acetonitrile was added drop by drop to a final concentration of 60% (v/v). Then, samples were vortexed and sonicated in an ultrasonic bath for 20 min. Supernatant was collected after centrifugation at 13,000 rpm for 20 min, and evaporated to dryness in SpeedVac. For tandem depletion, the sera were subjected to a sequential depletion protocol involving the combination of the two precipitation steps previous: protein depletion was first performed with DTT and the supernatant was treated with ACN in order. Following, the depleted protein were analyzed by SDS-PAGE and all proteic bands were, digested (trypsin) and analyzed by nanoLC-MS/MS.

Results: A total de 17 proteins were identified for DTT depletion. Of these, 9 belonged to the group of the major proteins (APOA1, IGHG1, A1AT, C4-A, IGKC, CERUM, APOB, HPT, A1AG1). It was found that this method reduce the presence of albumin in serum human sample. For ACN depletion, it was found that this method reduces the presence of protein above 75 KDa, for example apolipoproteins. The number of proteins identified in the depleted serum by ACN strategies was 16, of which 6 (HPT, A1AG1, ALBU, CERUM, IGKC, A1AT) belonged to the group of the major proteins. 4 (A1AG1, A1AT, CERUM, HPT) were the abundant proteins identified using tandem depletion. Thus, in this way the relative protein in-gel load of other less-abundant proteins was increased.

Conclusions: In this study, we evaluated the application of DTT, ACN and tandem depletion as an alternative procedure of protein major depletion for the search of disease biomarker in serum and accordingly tandem depletion present as an efficient tool.

Keywords: Bipolar disorder, depletion protein, serum human, SDS-PAGE, nanoLC-MS/MS.

Acknowledgements: FAPESP, CAPES, CNPq, FAEPEX.



Special Issue: Proceeding Abstracts of the 4th International Congress on Analytical Proteomics (ICAP 2015)

Evaluation of proteome alterations induced by cadmium stress in sunflower (*Helianthus annuus* L.) cultures using 2-D DIGE and nESI-Q-Tof

C. A. Lopes Júnior^{1,2,3}, R. M. Galazzi^{*1,2}, H. H. F. Koolen^{2,3}, F. C. Gozzo^{2,3}, H. S. Barbosa³, M. A. Z. Arruda^{1,2}

¹Spectrometry, Sample Preparation and Mechanization Group – GEPAM, Institute of Chemistry, University of Campinas, UNICAMP, P.O. Box 6154, 13083-970, Campinas-SP, Brazil. ²National Institute of Science and Technology for Bioanalytics, Institute of Chemistry, University of Campinas–UNICAMP, P.O. Box 6154, 13083-970, Campina-SP, Brazil. ³Department of Chemistry, Federal University of Piauí – UFPI, P.O. Box 64049-550, Teresina-PI, Brazil. ⁴Dalton Mass Spectrometry Group, Institute of Chemistry, University of Campinas – UNICAMP, P.O. Box 6154, 13083-970, Campinas-SP, Brazil. *Corresponding author: rodrigo.galazzi@iqm.unicamp.br

Available Online: 31 December 2015

ABSTRACT

Purpose: The present study evaluates, at a proteomic level, changes in the proteomic map sunflower leaves in the absence or presence (at 50 or 700 mg) of cadmium (as CdCl₂). Experimental description: At the end of the cultivation period (45 days), proteins are extracted from leaves with phenol, separated by two-dimensional difference gel electrophoresis (2-D DIGE), and excised from the gels. The differential protein abundances (for proteins differing by more than 1.8 fold, which corresponds to 90% variation) are characterized using nESI-Q-Tof. After the experiments LC-MS/MS, the identified proteins were evaluated using the String 9.1 program.

Results: The protein content decreases by approximately 41% in plants treated with 700 mg Cd compared with control plants. By comparing all groups of plants evaluated in this study (Control vs Cd-lower, Control vs Cd-higher and Cd-lower vs Cd-higher), 39 proteins are found differential and 18 accurately identified; the control vs Cdhigher treatment is that presenting the most differential proteins. From identified proteins, those involved in energy and disease/defense (including stress), are the ribulose biphosphate carboxylase large chain, transketolase, and heat shock proteins are the most differential abundant proteins. The analysis of the identified proteins using String program revealed that the major physiological processes of the plant affected by cadmium stress were as follows: primary metabolic process, cellular component organization, organic substance metabolic process and metabolic process, cellular process and response to stimulus.

Conclusions: According to the results, photosynthesis is the main process affected by the presence of high Cd concentrations in sunflowers, although the plants are highly tolerant to Cd exposure, as only minor changes were observed in their physiological processes. Therefore, proteomic results from this study are in agreement with those from our ionomic approaches (Lopes Júnior et al., Environ Exp Bot, 107, 2014, 180-186).

Keywords: sunflower; cadmium; proteomics; 2-D DIGE; LC-MS/MS.

Acknowledgements: The authors are grateful to CAPES, FAPESP and CNPq for financial support and fellowships.